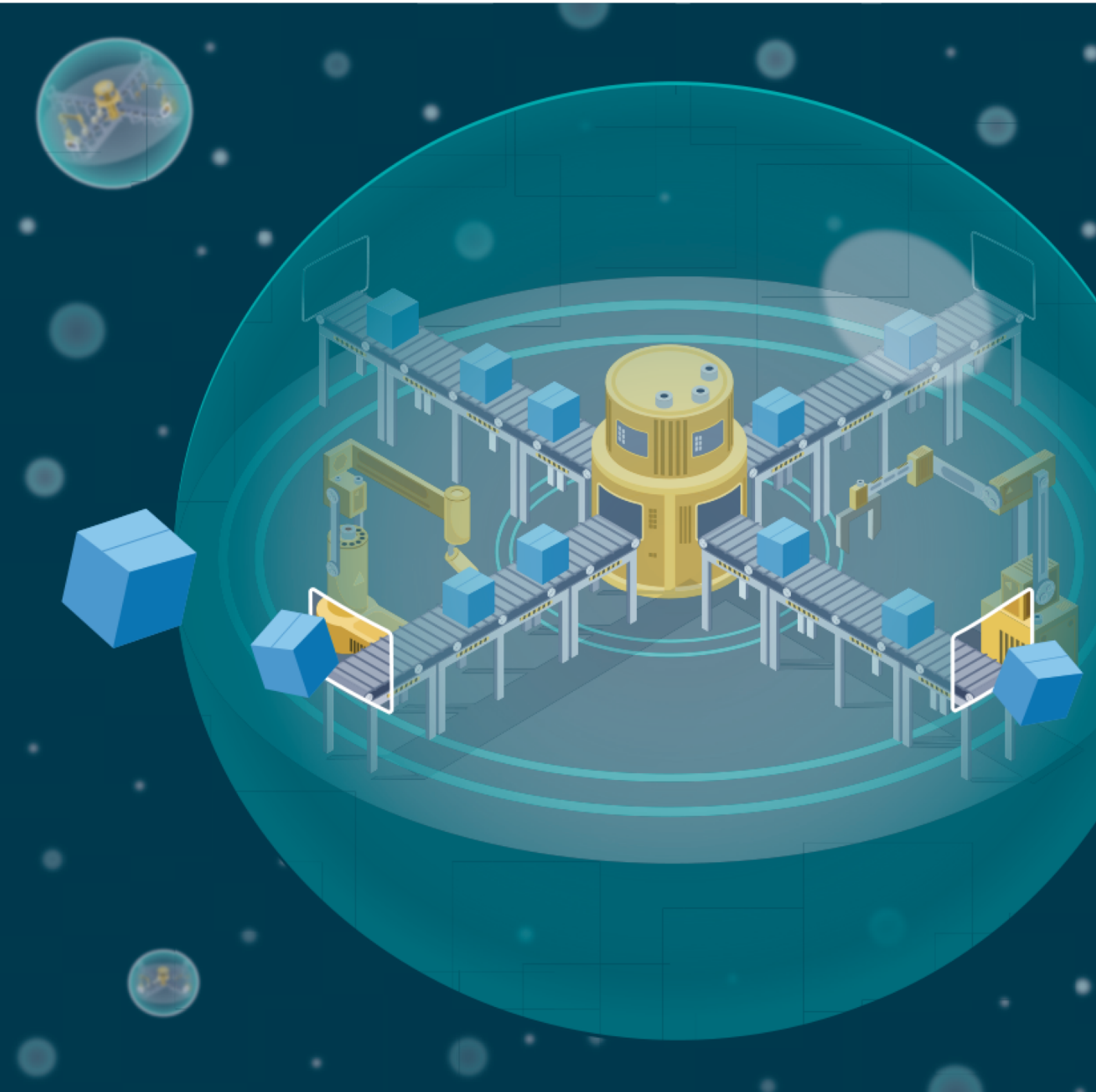


nature reviews rheumatology

March 2026



The future of autologous stem cell transplantation in systemic sclerosis

Charlotte Schimmel & Julia Spierings



Autologous haematopoietic stem cell transplantation revolutionized the treatment of severe systemic sclerosis as the first therapy able to induce long-term remission in this relentlessly fibrosing disease. Nevertheless, questions remain about patient selection, conditioning, and how this treatment fits into the evolving immune-modifying therapeutic landscape. The field should move beyond standardization towards precision therapy.

Systemic sclerosis (SSc) is a heterogeneous autoimmune disorder driven by vascular injury, immune system activation and progressive fibrosis of the skin and internal organs. Over the past two decades, autologous haematopoietic stem cell transplantation (AHSCT) has emerged as the most effective therapy for diffuse cutaneous SSc. Landmark trials, such as the ASTIS and SCOT trials, demonstrated that AHSCT confers substantial survival and quality-of-life benefits over cyclophosphamide-based regimens¹. Both meta-analyses and real-world data from large cohorts, such as the European Society for Blood and Marrow Transplantation, have confirmed these findings^{1,2}.

AHSCT enables immune system ‘reset’ through high-dose immunoblation followed by infusion of autologous haematopoietic progenitor cells, which leads to the reconstitution of a naive, self-tolerant immune repertoire. However, the risk of infection and treatment toxicity is considerably higher in AHSCT than in other immunosuppressive therapies, and treatment-related mortality occurs in up to 10% of patients, owing mainly to cardiopulmonary toxicity and infection¹.

With optimized protocols and careful patient selection, progression-free survival now exceeds 85% at 2 years after treatment. Cardiac screening has substantially improved safety outcomes³. As a result, the role of AHSCT has evolved from a last-resort intervention to a strategic therapy for select patients. The challenge has shifted from proving efficacy to refining who, when and how to treat. Considerable efforts have been made towards standardizing patient selection, conditioning and post-transplantation care. Pre-transplantation assessments routinely include 24-hour Holter monitoring, cardiac MRI, right-heart catheterization and pulmonary evaluation to mitigate risk³. Patients with advanced interstitial lung disease or considerable cardiac dysfunction are typically excluded, although debate continues about the inclusion of those with cardiac involvement – patients who might be both at higher risk and in greatest need of intervention.

Conditioning regimens strive to balance toxicity with the depth of immunodepletion. Cyclophosphamide combined with anti-thymocyte

globulin (ATG), with or without CD34⁺ cell selection, is most commonly used. Comparative analyses suggest that enrichment for CD34⁺ cells improves engraftment and immune system reset but might slightly delay reconstitution and incurs greater cost. Reduced-intensity regimens show promising short-term effectiveness, but confirmation of long-term benefits is needed, as relapse rates might be high³.

Management after transplantation, however, remains heterogeneous. Despite relapse rates being below 15%, some centres administer low-dose mycophenolate or methotrexate as maintenance therapy. From our perspective, the routine use of post-transplantation immunosuppression risks overtreatment and should be avoided. Although the use of such therapies might be warranted in patients with a high risk of relapse, reliable markers for identifying these patients are currently lacking. Efforts in 2025 have aimed to harmonize criteria for response and relapse, which represents an important step towards consistency, but the optimal role of post-transplantation immunosuppression remains undefined^{4,5}.

Research priorities in optimizing immune reset in AHSCT

Mechanistic studies have increasingly elucidated how AHSCT resets the immune system⁶. Transplantation eradicates autoreactive lymphocytes and regenerates a tolerant immune network, but the innate immune system, the dynamics of immune system reconstitution and the relationship with fibrosis and angiogenesis remain incompletely understood. Furthermore, the occurrence of severe cytokine-release syndrome and infections during and after AHSCT is rather unpredictable. Further research into the immunological repertoire and immune system dynamics will enhance understanding of these processes, guide patient selection and management, and help prevent these complications.

Emerging single-cell and immune repertoire analyses reveal heterogeneous recovery trajectories – some marked by skewed T cell receptor repertoires or functionality and levels of autoantibodies that are predictive of early relapse or poor outcome^{6,7}. Comparable progress in biomarker discovery, including transcriptomic signatures and serum cytokine profiles, might further enhance the evaluation of immune system reset quality⁶. Integrating such immunological metrics into clinical monitoring could enable risk-adapted conditioning and post-transplantation management.

In addition, integrated multidimensional data (clinical, genetic and immunological) is needed for the development of predictive models that guide treatment intensity and timing. Adaptive conditioning could tailor chemotherapy and ATG dosing to patient-specific factors such as comorbidities, immune kinetics and pharmacogenomic profiles⁸; for example, ATG dosing based on lymphocyte counts, or testing cytochrome P450 variants that influence cyclophosphamide metabolism and cardiac toxicity risk, offer opportunities for individualizing regimens and reducing complications.

Another research priority is the timing of interventions in SSc. Analyses from the European Society for Blood and Marrow Transplantation

and the Cochrane Library consistently show better outcomes with earlier transplantation^{1,2}. Once fixed-organ fibrosis develops, reversibility decreases and transplantation risk rises sharply. Hence, consensus is growing to perform AHST within the ‘window of reversibility’, which is typically within 5 years of disease onset and prior to irreversible cardiopulmonary damage. Prospective validation of timing thresholds remains an urgent knowledge gap and is currently being investigated in the randomized UPSIDE trial⁹.

Although AHST is increasingly used in severe SSs, the landscape of immunomodulation is rapidly evolving. Conceptually, AHST achieves a broad immune system reset, whereas emerging cellular therapies aim for selective immune deletion and modification. Among these, B cell-directed chimeric antigen receptor (CAR) T cell therapy has demonstrated efficacy in the treatment of refractory autoimmune diseases. Given the important role of B cells in SSs and the potential of rituximab, CAR T cell therapy could offer a precision-based strategy for achieving thorough and long-term B cell depletion¹⁰. Early data from patients with SSs suggest the potential for treating inflammatory manifestations, possibly for those with advanced disease or those who are not eligible for AHST. CAR T cell therapy might also be better tolerated than high-dose chemotherapy followed by AHST in this population, but more safety data are needed to confirm this observation. Furthermore, cytokine-release syndrome seems to be a frequent adverse event in CAR T cell therapy that requires more investigation. Moreover, the high cost and logistical complexity of manufacturing CAR T cells limits widespread implementation of this therapy.

‘Off-the-shelf’ therapies, including allogeneic CAR T cells, CAR natural killer cells and induced pluripotent stem cell-derived CAR T cells or generating CAR T cells *in vivo*, aim to reduce some of these barriers. Other innovations, such as bispecific antibodies (including bispecific T cell engagers), allogeneic mesenchymal stromal cells, CAR regulatory T cells and chimeric autoantibody receptor T cells, are still in the early stages of clinical development but might ultimately expand the therapeutic landscape by offering antifibrotic, immunomodulatory and tolerance-restoring effects in SSs.

Further research needs to be done on the (long-term) efficacy and safety of these emerging cellular therapies in patients with SSs. The most promising path forward will probably combine insights from AHST and novel therapies, using immunological profiling and clinical characteristics to identify the patients who will benefit from systemic immune system reboot versus targeted immune cell ablation.

A new vision for AHST in SSs

AHST remains the benchmark treatment for re-establishing immunotolerance in severe diffuse cutaneous SSs, a uniquely effective therapy proven to halt disease progression and induce sustained remission. Among the available interventions, it stands as the most successful strategy for reversing aggressive autoimmunity and preventing irreversible organ damage.

However, the field now faces a pivotal transition. Although AHST has matured into a standardized procedure, the next phase must focus on precision, prediction and personalization. The challenge is to determine which patients will benefit most; those for whom AHST should be

prioritized early, and those who might instead respond to less intensive approaches such as targeted DMARDs or biologic drugs.

Emerging immunological treatments, including CAR T cells, bispecific T cell engagers and related strategies, are highly promising but remain supported by limited evidence in SSs. Their cost, complexity and early-stage data underscore the need for rigorous comparative studies before their integration into standard care. Determining the right therapeutic sequence – whether AHST followed by maintenance or stepwise escalation from biologic to cellular therapy – will shape the next generation of immunological intervention.

Future priorities include refining timing thresholds for transplantation, validating biomarkers that predict immune reset success and rigorously evaluating how novel immunotherapies fit within the evolving therapeutic landscape. By taking these steps forward, AHST can evolve from a general intensive therapy into a precisely targeted recalibration of immune system homeostasis.

Charlotte Schimmel¹ & Julia Spierings^{1,2}✉

¹Department of Rheumatology & Clinical Immunology, University Medical Centre Utrecht, Utrecht, the Netherlands. ²Division of Medicine, Department of Inflammation, Centre for Rheumatology and Connective Tissue Diseases, Royal Free and University College Medical School, University College London, London, UK.

✉ e-mail: J.Spierings@umcutrecht.nl

Published online: 9 December 2025

References

1. Bruera, S. et al. Stem cell transplantation for systemic sclerosis. *Cochrane Database Syst. Rev.* **7**, CD011819 (2022).
2. Henes, J. et al. Autologous stem cell transplantation for progressive systemic sclerosis: a prospective non-interventional study from the European Society for Blood and Marrow Transplantation Autoimmune Disease Working Party. *Haematologica* **106**, 375–383 (2021).
3. Alexander, T. et al. Autologous haematopoietic stem cell transplantation for rheumatic diseases: best practice recommendations from the EBMT Practice Harmonization and Guidelines Committee. *Bone Marrow Transplant.* **60**, 1451–1464 (2025).
4. Del Papa, N. et al. Definition of relapse criteria in patients with rapidly progressive systemic sclerosis treated with autologous haematopoietic stem cell transplantation. *Bone Marrow Transplant.* **60**, 1537–1540 (2025).
5. Spierings, J. et al. Management of disease progression after autologous hematopoietic stem cell transplantation in systemic sclerosis: Results from an international questionnaire-based study. *Semin. Arthritis Rheum.* **71**, 152638 (2025).
6. Kawashima-Vasconcelos, M. Y., Santana-Gonçalves, M., Zanin-Silva, D. C., Malmegrim, K. C. R. & Oliveira, M. C. Reconstitution of the immune system and clinical correlates after stem cell transplantation for systemic sclerosis. *Front. Immunol.* **13**, 941011 (2022).
7. Bankamp, L. et al. Functional autoantibodies in systemic sclerosis: influence of autologous stem cell transplantation and correlation with clinical outcome. *Rheumatology* **62**, 2168–2177 (2023).
8. Chiu, Y. H. et al. Anti-thymocyte globulin exposure in patients with diffuse cutaneous systemic sclerosis undergoing autologous haematopoietic stem cell transplantation. *J. Scleroderma Relat. Disord.* **8**, 241–246 (2023).
9. Spierings, J. et al. A randomised, open-label trial to assess the optimal treatment strategy in early diffuse cutaneous systemic sclerosis: the UPSIDE study protocol. *BMJ Open* **18**, e044483 (2021).
10. Distler, J. H. W. et al. Emerging therapies for the treatment of systemic sclerosis. *Nat. Rev. Rheumatol.* **21**, 612–625 (2025).

Acknowledgements

The authors thank J.M. van Laar for discussing and proofreading this manuscript.

Competing interests

The authors declare no competing interests.

Research highlights

Gout

Novel approach mitigates immunogenicity of uricase treatment

Although uricase (urate oxidase) is a potent urate-lowering therapy, its use for the treatment of gout is limited by the formation of anti-drug antibodies (ADAs). Combination therapy with pegloticase and an immunosuppressant can reduce immunogenicity but typically requires frequent dosing and does not reliably prevent ADA formation. Research now published in *Science Advances* shows that in animal models, coadministration of erythrocyte-conjugated uricase with a single dose of cyclophosphamide can effectively treat gout by inducing durable immunotolerance.

“Conjugating uricase to erythrocytes ensures efficient, physiological targeting to the spleen, where splenic myeloid cells capture and process the antigen within a naturally tolerogenic clearance pathway,” explains corresponding author Xiaofei Gao.

In the study, coadministration of erythrocyte-conjugated uricase and a single dose of cyclophosphamide was able to prevent ADA formation in mice, even after repeated challenge with mouse erythrocyte–uricase conjugates or free uricase. This effect was not replicated by combination treatment with free uricase plus cyclophosphamide or by treatment with cyclophosphamide alone.

The researchers also showed that the induction of immunotolerance by

erythrocyte-conjugated uricase enhances the bioavailability and urate-lowering efficacy of uricase-based therapy in rodent models. Mechanistically, this tolerance induction required splenic uptake of erythrocyte-conjugated uricase, particularly by macrophages, and was associated with the expansion of splenic effector regulatory T cells.

Together, the findings suggest that erythrocyte-conjugated uricase could address a longstanding barrier to the use of uricase for the treatment of uncontrolled gout. The authors also suggest the findings highlight the broader potential of this approach. “Looking ahead, we anticipate that combining erythrocyte-mediated spleen targeting with transient immunosuppression could serve as the basis of a versatile tolerance-induction platform, with potential applications not only in gout but also in autoimmune diseases and immune-related toxicities where durable and selective immune tolerance is urgently needed,” Gao notes.

Sarah Onuora

Original article: Nie, X. et al. Single coadministration of erythrocyte-uricase conjugate and immunosuppressant induces durable immune tolerance for gout therapy. *Sci. Adv.* <https://doi.org/10.1126/sciadv.aea5196> (2025)

Related article: Schlesinger, N. et al. Mechanisms and rationale for uricase use in patients with gout. *Nat. Rev. Rheumatol.* **19**, 640–649 (2023)

Myositis

Monocyte-driven vasculopathy distinguishes dermatomyositis from CLE

Dermatomyositis is an autoimmune disease in which cutaneous inflammation can be difficult to distinguish from that observed in cutaneous lupus erythematosus (CLE). Although type I interferons contribute to pathology in both diseases, the mechanisms linking skin inflammation to the prominent vascular abnormalities characteristic of dermatomyositis (including nailfold capillary changes) have remained unclear. In a new study, single-cell RNA sequencing provides a cross-disease comparison that offers fresh insight into shared and disease-specific pathways.

The researchers profiled paired lesional and non-lesional skin, alongside matched peripheral blood, and compared these datasets with CLE and healthy skin. The analysis revealed an interferon-rich environment in dermatomyositis marked by enrichment for IL-18 and vascular endothelial growth factor pathways.

Several cell populations were expanded in dermatomyositis relative to their abundance in CLE, including Langerhans cells in non-lesional skin and peripheral helper T cells in lesional skin. Most striking, however, were the endothelial cells, which expressed markers of activation and a pronounced senescence signature in dermatomyositis. The study also uncovered a dermatomyositis-enriched subset of hyperinflammatory CD14⁺ monocytes in both the blood and skin. Ligand–receptor analysis and functional assays indicated that these monocytes promote endothelial cell apoptosis and dysfunction.

Notably, the JAK1 inhibitor upadacitinib reduced monocyte-induced endothelial apoptosis in vitro and decreased endothelial senescence signatures and inflammatory pathway activation in ex vivo-treated dermatomyositis skin samples. These effects were not fully reproduced by inhibition of TWEAK alone, which suggests that broader suppression of JAK1-dependent cytokine signalling is needed to disrupt the pathogenic interactions between monocytes and endothelial cells.

“the JAK1 inhibitor upadacitinib reduced monocyte-induced endothelial apoptosis”

“Clinically, we have known that JAK inhibition might be helpful for dermatomyositis overall, but now we are tying some specific mechanisms to this improvement,” explains J. Michelle Kahlenberg, corresponding author on the study. “We now need to better understand this pro-inflammatory monocyte population and whether it is associated with any particular disease phenotype. Better understanding of vascular–monocyte interactions in dermatomyositis will also be important.”

Jessica McHugh

Original article: Osborne, G. A. et al. Dermatomyositis is characterized by JAK1-mediated monocyte-driven vasculopathy and inflammation. *Sci. Transl. Med.* **17**, eaea9007 (2025)

Research highlights

Osteoarthritis

Denosumab targets synovial inflammation to attenuate knee OA

RANKL is a crucial regulator of abnormal bone remodeling in osteoarthritis (OA). In a phase IIa trial, targeting the RANK–RANKL pathway using denosumab was shown to improve erosive hand OA. However, the exact mechanisms through which this drug attenuates OA are unclear. Now, a study in *Nature Communications* provides insight into these mechanisms in the context of knee OA.

The authors first showed that high expression of RANK and RANKL in the synovium, particularly in fibroblast-like synoviocytes (FLSs), is associated with OA in mice and humans. Across three rodent models of OA (post-traumatic, inflammatory and age-related models) denosumab treatment reduced immune cell infiltration and abnormal bone remodelling and improved synovitis and pain. Similar results were also observed when using denosumab to treat knee OA in beagle dogs.

RNA sequencing analysis of whole joints from mice with OA indicated that FLSs were the primary target of denosumab. FLSs produce numerous pro-inflammatory molecules that perpetuate synovial inflammation and denosumab treatment inhibited these responses.

In vitro analyses validated these results: FLSs stimulated with RANKL secreted pro-inflammatory cytokines and extracellular matrix degrading enzymes. Treating chondrocytes and macrophages with media from these FLSs promoted inflammatory phenotypes in both cell types. Treatment with denosumab reversed these effects.

The RNA sequencing data revealed *Fstl1* as the most

highly expressed gene in FLSs from mice with OA, and that denosumab downregulated the expression of this gene.

Further research demonstrated that denosumab prevents RANKL from activating the RANK–TRAF6–NF- κ B pathway, thereby preventing the expression of FSTL1. Knocking down the expression of TRAF6 or FSTL1 in FLSs in vitro prevented RANK-induced pro-inflammatory effects.

Corresponding author Wei Tong notes that “this research highlights a previously underappreciated, inflammation-targeted action of denosumab in OA, extending its therapeutic rationale beyond bone protection to include direct modulation of the synovial microenvironment.”

The authors also conducted a small single-arm clinical trial of denosumab in 9 individuals with knee OA. After 6 months, a single treatment with denosumab improved synovitis, joint function and pain compared with baseline.

“Moving forward, we aim to address the limitations of systemic denosumab administration, such as low joint exposure and systemic adverse effects, while overcoming the challenges of intra-articular injections (such as infection risk, repetitive procedures and poor patient compliance),” comments Tong.

Holly Webster

Original article: Hu, Y. et al. Denosumab attenuates knee osteoarthritis progression by inhibiting synovial inflammation via the RANK/TRAF6/FSTL1 signalling. *Nat. Commun.* **16**, 11394 (2025)

Related article: Wittoek, R. et al. RANKL blockade for erosive hand osteoarthritis: a randomized placebo-controlled phase 2a trial. *Nat. Med.* **30**, 829–836 (2024)

Rheumatoid arthritis

Vagus nerve stimulation shows clinical benefits for RA in pivotal trial

The results of the pivotal RESET-RA trial demonstrate that active stimulation of the vagus nerve using an implantable device offers a safe and effective non-pharmacological option for the treatment of rheumatoid arthritis (RA).

In the trial, a vagus nerve-targeted neuroimmune modulation device was implanted into 242 people with moderately to severely active RA who had an inadequate response or intolerance to treatment with one or more biologic or targeted synthetic DMARD. Participants were randomly assigned to receive active or sham stimulation during a 3-month controlled phase; all patients were then eligible to receive open-label active stimulation through 12 months.

The primary end point of the study was met, as rates of ACR20 response were higher in the active stimulation arm than in the sham arm at 3 months

(35.2% versus 24.2%). In the open-label stimulation period, the proportion of responders further increased to 50.0% of all participants at 6 months and 52.8% at 12 months. Moreover, 44.8% of participants achieved a state of low disease activity or remission by DAS28-CRP criteria at 12 months.

As well as decreasing joint inflammation, the vagus nerve-stimulation treatment reduced progression of bone erosions in a subgroup of participants determined to be at high risk for structural damage at baseline. The active stimulation treatment, which was delivered for 1 minute daily, was generally well tolerated, and 78.1% of all participants reported being ‘somewhat to very satisfied’ with the therapy at 6 months.

Sarah Onuora

Original article: Tesser, J. R. P. et al. Vagus nerve-mediated neuroimmune modulation for rheumatoid arthritis: a pivotal randomized controlled trial. *Nat. Med.* **32**, 369–378 (2026)

Engineered sialylated IgG1 Fc as a dose-sparing alternative to IVIG

Sruthi Vijaya Retnakumar & Jagadeesh Bayry

 Check for updates

Intravenous immunoglobulin (IVIG) is a cornerstone of autoimmune disease therapy, but its use is constrained by high costs and limited supply. A sialylated IgG1 Fc variant with enhanced affinity for the inhibitory Fcγ receptor FcγRIIB could offer an effective dose-sparing alternative to IVIG, potentially transforming treatments for autoimmune diseases.

REFERS TO Jones, A. T. et al. The anti-inflammatory activity of IgG is enhanced by co-engagement of type I and II Fc receptors. *Science* **390**, ead2927 (2025).

Intravenous immunoglobulin (IVIG) is a preparation of pooled normal IgG derived from the plasma of thousands of healthy donors. IVIG has been a mainstay in clinical practice for several decades, initially introduced as a replacement therapy for people with immunodeficiencies and later widely used – at high doses (1–2 g kg⁻¹ body weight) – to treat an array of autoimmune and inflammatory disorders owing to its immunomodulatory and anti-inflammatory effects. Despite its established efficacy, the clinical use of IVIG is challenged by limited supply, high dosing requirements and high costs, prompting efforts to develop effective, affordable and sustainable alternatives to this complex biologic therapy^{1,2}. Research now demonstrates that a recombinant sialylated IgG1 Fc variant with enhanced affinity for inhibitory Fcγ receptor IIB (FcγRIIB) effectively treats autoimmune disease models at substantially lower doses than conventional IVIG therapy, while uncovering new mechanisms that drive the anti-inflammatory activity of sialylated Fc³.

The immunomodulatory activity of IVIG is explained by several mutually non-exclusive mechanisms, targeting many components of the immune system and mediated by its F(ab')₂ and Fc fragments^{1,2}. Mechanisms proposed to underlie the Fc-mediated action of IVIG include the competitive blockade of activating type I Fc receptors (the canonical FcγRs), which reduces immune complex-driven inflammation; saturation of the neonatal Fc receptor, leading to accelerated clearance of pathogenic autoantibodies; and upregulation of the inhibitory receptor FcγRIIB, which serves to restrain pro-inflammatory responses. Furthermore, experimental studies have demonstrated that terminal sialic acid residues in the IgG Fc domain are crucial⁴. Whereas non-sialylated Fc adopts an open conformation, which makes it accessible to canonical type I FcRs, sialic acid glycans confer a closed Fc conformation that occludes type I FcR binding and instead enables binding to type II FcRs, which are represented by a family of C-type lectin receptors that includes dendritic cell-specific intercellular adhesion molecule

3-grabbing non-integrin (DC-SIGN) and CD23 (ref. 5). However, the precise contribution of sialylation and the requirement for DC-SIGN in IVIG binding and activity have been debated, and cross-species differences in FcγR expression and affinity for human IgG complicate extrapolation from animal models to human disease⁶.

To address these issues, Jones et al.³ used recombinant sialylated IgG1 Fc fragments (sFc) in FcγR-humanized mice to confirm the role of Fc sialylation and type I FcγR engagement in the anti-inflammatory effects of IVIG. The sFc showed anti-inflammatory activity prophylactically in a serum transfer-induced arthritis model at a 25-fold lower dose than IVIG, whereas non-sialylated Fc or a mutant IgG1 Fc that cannot engage type I FcγRs did not protect mice from arthritis³ (Fig. 1).

Blockade of activating FcγRs was one of the first theories proposed to explain the therapeutic mechanism of IVIG, especially because high doses of IVIG are required. However, there has not been formal proof-of-concept for this mechanism, and it has subsequently been shown that the inhibitory receptor FcγRIIB is essential for the therapeutic effect of IVIG in several models of autoimmune diseases, which shifted researchers' focus to the modulation of inhibitory signalling as a prevailing mechanism of IVIG therapy⁷. To further explore these ideas, Jones et al.³ engineered different sFc variants with altered affinity to the inhibitory receptor FcγRIIB or the activating receptor FcγRIIA and investigated whether these variants can protect against inflammation at a lower dose than wild-type sFc. In the serum transfer-induced arthritis model, the V11 sFc variant, which has a higher affinity (~37-fold) than wild-type sFc for FcγRIIB, showed protective efficacy comparable to that of wild-type sFc even at one-tenth the dose, whereas the GA sFc variant, which has a higher affinity (~10-fold) than wild-type sFc for FcγRIIA, was ineffective at this lower dose, indicating that blockade of the activating receptor FcγRIIA was not implicated in the anti-inflammatory actions of sFc. Although removal of sialylated glycans by neuraminidase treatment reduced the efficacy of V11 sFc, mice treated with this variant were nonetheless significantly protected from inflammation, indicating that sFc has sialylation-independent effects. In addition, the researchers demonstrated the protective effect of V11 sFc in a FcγR-humanized mouse model of multiple sclerosis, supporting the broad translational potential of this strategy across multiple autoimmune diseases.

Another key finding from the study is the synergistic cooperation between type I and type II Fc receptors in mediating the anti-inflammatory effects of sFc. The findings demonstrate that DC-SIGN directly interacts with the ectodomains of type I FcRs through a glycan-dependent mechanism, thereby enhancing the binding of sFc to FcγRIIB. This coordinated engagement of FcγRIIB and DC-SIGN underpins the anti-inflammatory activity of sFc and can be further augmented by engineering sFc variants with increased affinity for FcγRIIB³.

The engineered sFc developed by Jones et al.³ is an elegant therapeutic approach with the potential for broader application in various autoimmune diseases. In terms of clinical outcomes, the engineered sFc recapitulates the anti-inflammatory effects of IVIG at markedly lower

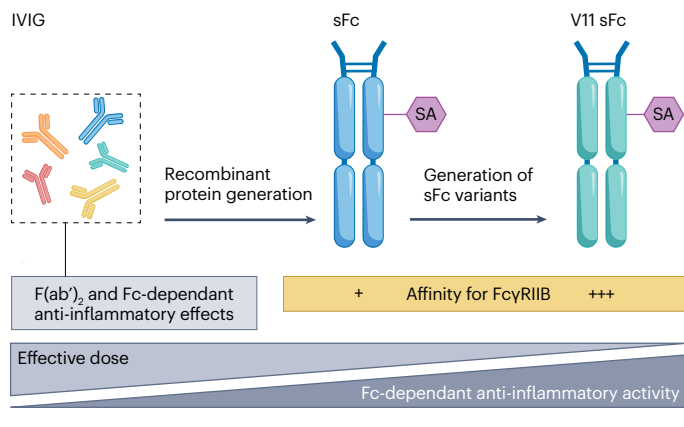


Fig. 1 | Engineered sialylated IgG1 Fc as a dose-sparing alternative to IVIG. Recombinant sialylated IgG1 Fc fragments (sFc), engineered by mutagenesis to have increased affinity for the inhibitory receptor FcγRIIB, reproduce the anti-inflammatory effects of intravenous immunoglobulin (IVIG) at substantially lower doses in FcγR-humanized mouse models of autoimmune diseases. SA, sialic acid.

doses, thus reducing the dependence on large numbers of healthy donors and intricate fractionation procedures and unlocking prospects for scalable and reliable therapeutic production. It is important to note that other Fc-derived molecules, such as an IgG1 Fc hexamer that mimics the action of IVIG, are also in the clinical development⁸. Furthermore, the engineering of monoclonal antibodies to enhance the interaction of Fc with FcγRs or complement proteins is a rapidly evolving area that has already led to several approved treatments for cancer and other diseases, further reinforcing the clinical safety and translational potential of these molecular modifications⁹.

Dissection of the mechanisms of action of IVIG in humans and experimental models has revealed several important features. Some mechanisms are mediated by the Fc region, whereas others depend on the F(ab')₂ fragments. Notably, certain effects require the presence of intact IgG molecules^{1,2}. For example, activation of the autophagy pathway in innate immune cells depends primarily on the F(ab')₂ fragments¹. By contrast, regulation of immune metabolism is mediated by both Fc and F(ab')₂ fragments but is dependent on the sialylated glycans of IgG¹⁰. Together, the evidence indicates that the therapeutic benefits of IVIG do not arise from a single pathway but instead from a set of mutually non-exclusive mechanisms acting together. In this context, it remains uncertain whether recombinant sFc alone can fully reproduce the cellular and molecular actions of IVIG, or how any differences in the molecular mechanisms, regulatory pathways, or target cells might influence long-term clinical outcomes. Addressing these issues will require detailed mechanistic studies, carefully designed clinical trials, and systematic head-to-head comparisons with conventional IVIG across several indications.

In summary, the work by Jones et al.³ provides strong preclinical evidence that enhancing the affinity of sFc for the inhibitory receptor FcγRIIB substantially enhances its anti-inflammatory effects and offers a potential candidate for clinical development for the treatment of autoimmune diseases. Furthermore, by leveraging the use

of FcγR-humanized mouse models, the study sheds clear light on disputed theories regarding the role of various FcγRs and sialylation in the anti-inflammatory effects of IVIG.

Sruthi Vijaya Retnakumar¹ & Jagadeesh Bayry^{1,2} ✉

¹Institut National de la Santé et de la Recherche Médicale, Centre de Recherche des Cordeliers, Sorbonne Université, Université Paris Cité, Paris, France. ²Department of Biological Sciences & Engineering, Indian Institute of Technology Palakkad, Palakkad, India.

✉ e-mail: bayry@iitpkd.ac.in

Published online: 5 January 2026

References

- Bayry, J. et al. Intravenous immunoglobulin: mechanism of action in autoimmune and inflammatory conditions. *J. Allergy Clin. Immunol. Pract.* **11**, 1688–1697 (2023).
- Schwab, I. & Nimmerjahn, F. Intravenous immunoglobulin therapy: how does IgG modulate the immune system? *Nat. Rev. Immunol.* **13**, 176–189 (2013).
- Jones, A. T., Marino, A. E., Martynyuk, T., Bournazos, S. & Ravetch, J. V. The anti-inflammatory activity of IgG is enhanced by co-engagement of type I and II Fc receptors. *Science* **390**, eadv2927 (2025).
- Schwab, I. et al. Broad requirement for terminal sialic acid residues and FcγRIIB for the preventive and therapeutic activity of intravenous immunoglobulins in vivo. *Eur. J. Immunol.* **44**, 1444–1453 (2014).
- Pincetic, A. et al. Type I and type II Fc receptors regulate innate and adaptive immunity. *Nat. Immunol.* **15**, 707–716 (2014).
- Temming, A. R. et al. Human DC-SIGN and CD23 do not interact with human IgG. *Sci. Rep.* **9**, 9995 (2019).
- Samuelsson, A., Towers, T. L. & Ravetch, J. V. Anti-inflammatory activity of IVIG mediated through the inhibitory Fc receptor. *Science* **291**, 484–486 (2001).
- Spirig, R. et al. rIgG1 Fc hexamer inhibits antibody-mediated autoimmune disease via effects on complement and FcγRs. *J. Immunol.* **200**, 2542–2553 (2018).
- Abdeldaim, D. T. & Schindowski, K. Fc-engineered therapeutic antibodies: recent advances and future directions. *Pharmaceutics* **15**, 2402 (2023).
- Rambabu, N. et al. Regulation of immune cell metabolism by therapeutic normal IgG intravenous immunoglobulin. *J. Allergy Clin. Immunol.* **156**, 418–432 (2025).

Competing interests

The authors declare no competing interests.

The obesity–inflammation axis in psoriatic disease: mechanisms and therapeutic strategies

Rebecca H. Haberman¹, Alexis Ogdie², Joseph F. Merola^{3,4}, Jose U. Scher^{1,5} & Lihi Eder⁶✉

Abstract

Obesity constitutes a substantial burden in psoriatic disease that affects approximately half of patients. Importantly, increased adiposity and psoriatic disease are strongly linked, with obesity functioning as both a possible trigger and a disease modifier. Obesity predisposes individuals to develop psoriasis and is likely to drive, at least partially, the progression from psoriasis to psoriatic arthritis. For people with psoriasis or psoriatic arthritis, obesity is associated with lower rates of remission and poorer responses to treatment. Several mechanisms probably underlie this relationship, including systemic and local pro-inflammatory properties of adipose tissue, increased biomechanical stress on joints and entheses, gut dysbiosis and synergistic effects of osteoarthritis. Notably, weight loss can improve both psoriatic disease course and response to therapy; however, current approaches (such as dietary interventions or bariatric surgery) are difficult to implement. Glucagon-like peptide-1-based therapies are an effective strategy for weight loss in psoriatic disease and might even have additive disease-modifying effects to conventional immunomodulators. Although often overlooked, weight loss intervention and obesity management should be included as an integral part of psoriatic disease treatment algorithms.

Sections

Introduction

Clinical link between psoriatic disease and obesity

Mechanisms that link obesity to inflammation in psoriatic disease

Weight loss interventions in psoriatic disease

Unmet needs and future research

Proposed strategies to address obesity in psoriatic disease

Conclusions

¹Division of Rheumatology, Department of Medicine, New York University Grossman School of Medicine, New York, NY, USA. ²Division of Rheumatology, Department of Medicine, Perelman School of Medicine at the University of Pennsylvania, Philadelphia, PA, USA. ³Department of Dermatology, UT Southwestern Medical Center, Dallas, TX, USA. ⁴Division of Rheumatology, Department of Medicine, UT Southwestern Medical Center, Dallas, TX, USA. ⁵Colton Center for Autoimmunity, New York University Grossman School of Medicine, New York, NY, USA. ⁶Division of Rheumatology, Department of Medicine, University of Toronto and Women's College Hospital, Toronto, Ontario, Canada. ✉e-mail: Lihi.Eder@wchospital.ca

Key points

- Obesity is a chronic, relapsing disease characterized by excess adipose tissue, which affects up to 50% of individuals with psoriatic disease.
- Obesity and psoriatic disease are probably causally linked. Clinical evidence demonstrates that increased adiposity is associated with higher risk of psoriatic disease development, worse disease severity and decreased therapeutic response.
- Several mechanisms might underlie this relationship, including adipose dysfunction, metaflammation, biomechanical stress, gut dysbiosis, pain amplification and exacerbation of obesity-related comorbidities.
- Weight loss can improve psoriatic disease outcomes. Glucagon-like peptide-1 receptor agonists might have additional anti-inflammatory functions, providing a synergistic effect to existing immunomodulatory therapies.
- Addressing obesity is crucial in the care of individuals with psoriatic disease and weight loss should be systematically integrated into psoriatic disease treatment algorithms.

Introduction

Psoriasis is an inflammatory skin condition that affects 2–3% of people worldwide¹. Up to one-third of individuals with psoriasis ultimately develop psoriatic arthritis (PsA), a chronic immune-mediated disease characterized by joint and enthesal pain, stiffness and swelling². Together, psoriasis and PsA are often termed psoriatic disease, to acknowledge their overlapping features and shared pathophysiology. Although key cytokines, such as IL-23, IL-17 and TNF, cause the characteristic inflammatory symptoms of psoriatic disease, the underlying drivers of this immune system dysfunction remain largely unknown and are probably multifactorial. Metabolic, genetic, mechanical and microbial factors have all been implicated in the pathogenesis of psoriatic disease³.

Obesity is a chronic, relapsing, progressive disease, characterized by excess adipose tissue that impairs health⁴. Overweight and obesity are defined as a BMI of 25–30 mg/m² and ≥ 30 mg/m², respectively⁵; additional anthropometric measurements, such as waist circumference, might also be considered. Although it cannot explain all of psoriatic disease susceptibility, the association of obesity with psoriatic disease is particularly intriguing. Advances in the understanding of the pathophysiology of adipose tissue dysfunction have identified obesity as a chronic low-grade inflammatory state, with immune system alterations similar to those observed in psoriatic disease. Additionally, unlike other contributors to psoriatic disease susceptibility and outcomes, obesity might be modifiable, particularly with the emergence of effective therapies such as glucagon-like peptide1 (GLP-1)-based therapies.

In this Review, we examine the influence of obesity on the risk of psoriatic disease development and on disease outcomes. We briefly discuss the possible mechanisms underlying the obesity–inflammation axis in psoriatic disease, including metaflammation, biomechanical stress, gut dysbiosis, pain amplification and adipose-related comorbidities. Finally, we explore the effect of weight loss strategies in psoriatic

disease, propose a framework for the management of obesity in people with psoriatic disease and highlight existing gaps in knowledge.

Clinical link between psoriatic disease and obesity

Overweight and obesity are among the most common comorbidities in psoriatic disease, reported in up to 70% of individuals with psoriatic disease^{6–16}. Individuals with psoriasis are almost two times more likely to have obesity compared with the general population^{17,18}. Furthermore, visceral adipose tissue, which is a predictor of cardiovascular adverse events, can be increased in individuals with psoriatic disease compared with those with equivalent BMIs¹⁹. Importantly, increased adipose tissue functions as both a possible trigger for psoriatic disease development and modifier of disease severity and response to treatment.

Risk of psoriatic disease development

Adiposity increases the likelihood of developing psoriatic disease. In a population-based study, the risk of developing skin psoriasis in people with obesity is nearly double that in people with a normal body weight²⁰. Furthermore, the risk of psoriasis increases with BMI in a dose-dependent manner. In the Nurses' Health Study, compared with individuals with normal weight, individuals with overweight had a relative risk of 1.21 for developing psoriasis, which increased to 1.63 and 2.03 in those with a BMI of 30–34.9 mg/m² and ≥ 35 mg/m², respectively²¹. Measures of central adiposity and weight gain have also been identified as risk factors for psoriasis, independent of baseline BMI^{20–23}.

The same pattern is observed in PsA. In a Norwegian population-based study, the relative risk of PsA in individuals with overweight and obesity were 1.4 and 2.5, respectively²⁴, whereas a US population-based study showed that individuals with a BMI of ≥ 35 mg/m² have a risk of developing PsA more than six times that of individuals with a normal BMI²⁵. In individuals with psoriasis, the relative risk of progressing to PsA in individuals with obesity ranges from 1.2 to 2.7 (refs. 26–28). Although weight loss can be associated with reduced risk of progression to PsA²⁸, higher BMI at 18 years of age was predictive of PsA in a prospective psoriasis cohort²⁹, indicating that early adiposity might have long-term effects on the risk of psoriatic disease.

Obesity and psoriatic disease severity

Obesity is associated with worse skin psoriasis^{30,31}. Individuals with obesity have a 47% higher risk of severe skin involvement than those without obesity⁶. In a Taiwanese study, compared with those with a normal weight, the likelihood (odds ratio) of individuals with obesity having severe psoriasis was 2.7 (ref. 32). A dose-dependent relationship is observed between BMI and the degree of skin involvement; an increase of 0.25 units in Psoriasis Area and Severity Index (PASI) has been reported for each unit increase in BMI³³.

Adiposity is also associated with worse measures of PsA; individuals with obesity are almost 50% less likely than those with a normal BMI to achieve sustained minimal disease activity (MDA)¹⁴. These individuals are also more likely to have sonographic enthesal abnormalities³⁴ and, in those with axial spondyloarthritis (axSpA), increased BMI is linked to new bone formation (that is, syndesmophytes and enthesophytes)^{35–38}. In addition, individuals with obesity have higher tender joint and enthesitis counts and worse pain, physical function and quality of life compared with those without obesity^{39,40}.

Obesity and poor response to treatment

The presence of obesity impedes treatment of skin psoriasis^{41–44}. In two large cohort studies, a BMI of ≥ 30 mg/m² reduced the likelihood of

achieving a $\geq 75\%$ improvement in PASI score (PASI75) by 25% compared with a normal BMI^{45,46}. This effect, however, might be specific to the therapeutic mechanism of action, as BMI affects response to treatment with TNF and IL-17 inhibitors in psoriasis⁴⁶. Meanwhile, several small, real-world studies have shown that obesity has a minimal effect on psoriasis response to IL-23p19 inhibitors or phosphodiesterase-4 inhibitors^{47–51}. Obesity also decreases treatment persistence^{41,52,53}. An assessment of drug survival rates of first-course biologic or targeted synthetic DMARDs (bDMARDs or tsDMARDs) in the treatment of patients with psoriasis showed that individuals with a BMI of ≥ 35 mg/m² are more likely to discontinue their medication owing to ineffectiveness, rather than adverse effects, compared with individuals with a normal BMI⁵⁴. In a meta-analysis of 16 cohorts of people with psoriasis, obesity predicted treatment discontinuation with a hazard ratio of 1.2 (ref. 55).

Obesity also reduces the chance of achieving remission after initiation of bDMARDs or tsDMARDs in patients with PsA^{56,57}. Across several disease states (inflammatory bowel disease, rheumatoid arthritis, psoriasis, PsA or axSpA), obesity is associated with 60% higher odds of non-response to TNF inhibitors compared with those without obesity⁵⁸. A one unit increase in BMI was associated with a 6.5% increase in the odds of non-response to treatment, further demonstrating a dose–response relationship with BMI⁵⁸. Most, but not all, studies of TNF inhibitors in PsA have shown similar results; obesity reduces the chance of achieving remission by 50%^{56,59–61}. It remains unclear if obesity affects all classes of medication. In a large cohort study, individuals with obesity had a ~50% reduction in remission regardless of the mechanism of action of the drug; however, over 90% of the patients were receiving TNF inhibitors, which limits generalizability⁵⁷. Additionally, except for phosphodiesterase-4 inhibitors, which can lead to weight loss, bDMARDs or tsDMARDs do not ameliorate obesity^{62,63}. Although treatment with TNF inhibitors might be linked with decreased cardiovascular events, use of these drugs is also associated with mild weight gain^{63–65}. Studies assessing the effect of obesity on treatment persistence in PsA have shown mixed results^{59,66–68}. This attenuated efficacy of advanced therapies in psoriatic disease might result from immunological and/or pharmacological factors.

Pharmacokinetic factors. Independent of immune effects, obesity alters drug pharmacokinetics. Increased adiposity expands the volume of distribution and might accelerate clearance of monoclonal antibodies, particularly when administered at fixed doses. This change can result in lower serum drug levels and subtherapeutic tissue exposure in individuals with high body mass⁶⁹. Although weight-based dosing of infliximab and dose escalation of ustekinumab in individuals with a body weight of >100 kg offers partial compensation, these strategies might be insufficient. Notably, studies have indicated that BMI is inversely correlated with ustekinumab efficacy, suggesting that adiposity-related factors beyond volume of distribution, such as immune dysregulation, also have a role⁷⁰. Fixed-dose IL-23p19 inhibitors might be similarly affected⁷¹; pharmacokinetic studies have shown reduced drug levels in individuals with a body weight of >100 kg⁷².

Similar to TNF and IL-23 inhibitors, clinical pharmacology data for secukinumab, an IL-17A inhibitor, show lower serum drug levels and diminished response in individuals with a body weight of ≥ 90 kg, leading to recommendations for increased dosing in this subgroup⁷³. Indeed, drug administration every 2 weeks (rather than every 4 weeks) improves outcomes in individuals with obesity. Although dose optimization might mitigate pharmacological challenges, an adjunctive and

potentially synergistic strategy is weight reduction. Dietary interventions leading to moderate weight loss enhance response to TNF inhibitors in individuals with obesity and PsA, which offers dual benefits for inflammatory disease control and cardiometabolic health⁷⁴.

Immune mechanisms. Several biologic therapies for PsA target key pro-inflammatory cytokines, such as TNF and IL-17, which are elevated in obesity. The ‘sink effect’ is a pharmacological hypothesis proposed to explain reduced TNF inhibitor efficacy in obesity, whereby increased adipose-derived TNF functions as a reservoir that binds and sequesters therapeutic monoclonal antibodies and thus diminishes their bioavailability at sites of inflammation (such as the synovium and skin)⁷⁵. Additionally, obesity is associated with upregulation of the IL-23–T helper 17 (T_H17) cell pathway, which has a central role in psoriasis and PsA, resulting in worsening of the skin and musculoskeletal inflammation in people with PsA and obesity^{76,77}. Adipokine imbalances, including increased levels of leptin and IL-6, can enhance T_H17 cell differentiation and IL-17 production, potentially overwhelming the suppressive capacity of IL-17 inhibitors in individuals with obesity and a heightened baseline pro-inflammatory milieu^{76,77}.

Mechanisms that link obesity to inflammation in psoriatic disease

Obesity is increasingly recognized as both a causal contributor and a disease modifier in psoriatic disease (Fig. 1). Chronic metabolic inflammation (that is, inflammation that results from metabolically active organs such as adipose tissue, the liver, brain and pancreas) which is termed ‘metaflammation’^{78,79} and is associated with obesity, creates a systemic, low-grade pro-inflammatory milieu that might lower the threshold for PsA development in genetically or immunologically primed individuals. This state might exacerbate pre-existing synovio-enthesal inflammation through shared immune pathways or trigger musculoskeletal inflammation through biomechanical stress or other mechanisms. Nonetheless, it remains unclear if obesity initiates musculoskeletal inflammation in PsA or instead amplifies pre-existing inflammation that originates at the skin, gut or joint level, thereby promoting chronicity and worsening clinical manifestations. Clarifying this distinction is important, as it might inform the development of preventative strategies aimed at halting the progression of psoriasis to PsA and worsening of PsA phenotype (Fig. 2). Obesity could also influence disease outcomes through other mechanisms. In this section, we review several potential pathways by which obesity modulates psoriatic disease.

Adipose dysfunction and metaflammation in obesity

Obesity is now recognized as a complex, chronic, progressive and relapsing disease, characterized by an excessive accumulation of adipose tissue that adversely impacts health⁸⁰. Although environmental influences, such as dietary habits and physical inactivity have an important role, increasing evidence suggests that genetic and epigenetic factors contribute substantially to the pathogenesis of obesity^{81,82}. A defining feature of obesity is the phenotypic remodelling and altered distribution of white adipose tissue (WAT), which underlies many of the associated metabolic alterations, including insulin resistance, systemic inflammation and dyslipidaemia⁸³.

WAT is a metabolically active endocrine and lipid storage organ that has a pivotal role in energy homeostasis and immune regulation⁸⁴. Obesity arises from sustained energy imbalance, excessive energy intake relative to energy expenditure, leading to triglyceride accumulation

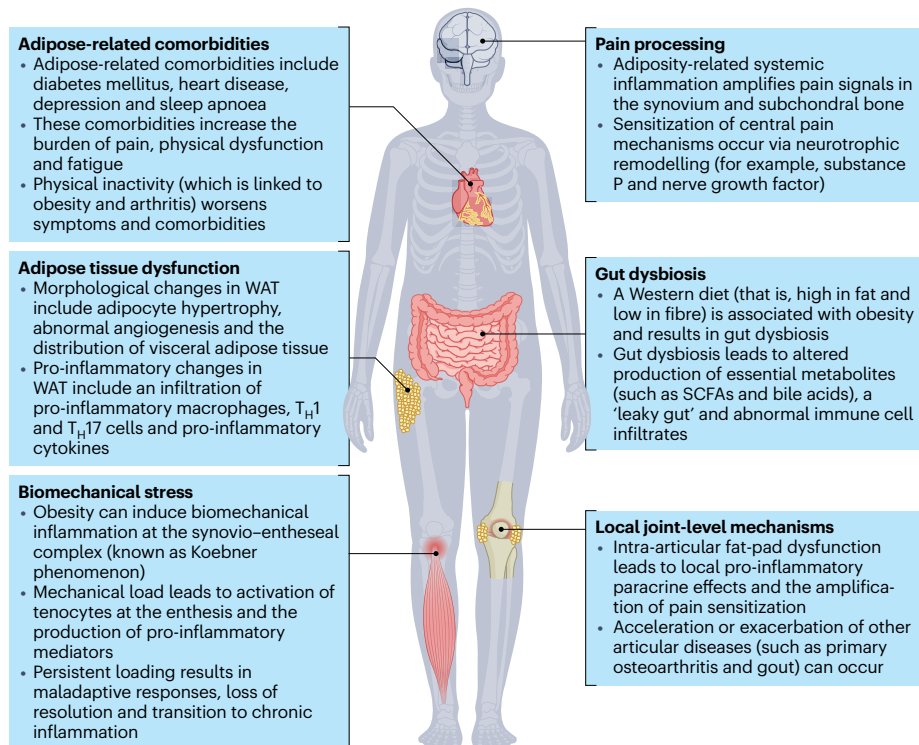


Fig. 1 | Potential mechanistic links between obesity and psoriatic disease. There are several mechanisms by which obesity might affect the onset, severity and symptoms of psoriatic disease, including via adipose-related comorbidities, adipose tissue dysfunction, metaflammation, biomechanical stress, pain processing, gut dysbiosis and local joint-level mechanisms. SCFA, short-chain fatty acids; T_H, T helper; WAT, white adipose tissue.

within adipocytes. With chronic energy surplus, adipose tissue expands beyond its physiological storage capacity, resulting in phenotypic changes in adipose tissue distribution, morphology and function⁸⁵.

Adipose tissue dysfunction exerts systemic effects through both metabolic and immunological pathways (Fig. 3). Metabolic consequences include insulin resistance and altered lipid handling. Concurrently, a shift towards metaflammation occurs. This shift is characterized by an infiltration of immune cells, particularly macrophages, which form pro-inflammatory niches that sustain systemic inflammation and contribute to obesity-related comorbidities, including type 2 diabetes mellitus (T2DM), cardiovascular disease and potentially psoriatic disease⁸³.

Obesity profoundly alters immune system dynamics by promoting the activation of immune pathways relevant to PsA, particularly those involving T_H1 and T_H17 cells and innate immune components^{86,87}. In obesity, macrophages become the predominant innate immune cell population in the dysfunctional WAT, comprising up to 50% of this population⁸⁸. These macrophages shift towards a pro-inflammatory phenotype and secrete pro-inflammatory cytokines, such as TNF⁸⁹. Adipocyte death, a hallmark of WAT dysfunction, promotes macrophage recruitment and the formation of crown-like structures⁹⁰. In parallel, adaptive immune cells within the dysfunctional adipose tissue exhibit skewed profiles, with increased T_H1 and T_H17 cell polarization and diminished regulatory T (T_{reg}) cell responses, promoting sustained inflammation^{86,87,91}. Adipokines are bioactive molecules secreted by adipocytes that mediate several of these pro-inflammatory effects; for example, leptin, which is elevated in obesity and psoriatic disease, activates the JAK-STAT3 pathway, promotes pro-inflammatory macrophage polarization and drives the differentiation and activation of T_H1 and T_H17 cells, thereby amplifying TNF, IL-1 β and IL-6 production^{92,93}.

Metaflammation in PsA

Clinical and translational studies support the role of metaflammation in modulating PsA risk and severity^{27,94}. Individuals with PsA have increased visceral and abdominal adiposity and this adiposity correlates with reduced muscle mass, which correlates with heightened production of pro-inflammatory cytokines and adipokines⁹⁵⁻⁹⁷. Mendelian randomization studies have suggested that metabolic disturbances, such as insulin resistance and hypertriglyceridaemia, confer an increased risk of psoriasis and PsA independent of BMI, reinforcing a causal link⁹⁸⁻¹⁰⁰. Moreover, Mendelian randomization analyses support a bidirectional relationship; psoriasis (and possibly PsA) might contribute to increased adiposity and metabolic dysfunction, potentially via inflammation-induced behaviours, such as poor diet quality and reduced exercise, or systemic metabolic changes, such as inflammation-induced glucose intolerance and atherogenic lipid profile, thus establishing a feedforward loop between psoriatic disease and obesity, termed the 'psoriatic march'^{101,102}.

Findings from animal models of psoriasis underscore the contribution of high-fat diet and metabolic factors to the altered immune state observed in psoriasis. In a high-fat diet mouse model of psoriasis, obese mice exposed to imiquimod developed more severe cutaneous disease and have increased IL-17 responses compared with lean mice¹⁰³⁻¹⁰⁵. Obese mice also show an increase in NLRP3 inflammasome activation, resulting in IL-1 β and IL-18 production, which promotes IL-17-producing $\gamma\delta$ T cells and worsened skin inflammation¹⁰⁵. These findings highlight a potential mechanistic link between diet quality, innate immune activation and T_H17 cell-driven pathology in psoriatic disease.

As described previously, adipokines secreted by dysfunctional adipose tissue modulate both immune and metabolic pathways. In psoriatic disease, adipokine profiles shift towards a pro-inflammatory phenotype;

elevated levels of leptin, resistin, and visfatin contribute to T_H1 and T_H17 cell polarization and T_{reg} cell dysfunction^{106–109}. An immunohistochemistry study further showed that leptin and its receptor are overexpressed in psoriatic skin, with expression levels correlating with disease severity and duration¹¹⁰. Increased leptin levels in PsA, particularly in women, are associated with increased pain and osteoclast precursor activity^{111,112}, which highlights the potential contribution of this increase to musculoskeletal pathology. Animal models offer additional insights into adipokine function. Leptin-deficient mice, despite being profoundly obese, are protected from developing psoriasis and osteoarthritis, underscoring the pro-inflammatory role of leptin^{113,114}. Conversely, the role of adiponectin, an adipokine that is typically anti-inflammatory, remains ambiguous. Despite an inverse association with adiposity, adiponectin levels are paradoxically elevated in PsA and are linked with greater disease activity, suggesting a complex, context-dependent role^{107,115}.

Intra-articular fat pads, such as the infrapatellar fat pad in the knee, have dual mechanical and immunological roles. In addition to functioning as shock absorbers in joints, they secrete adipokines and cytokines that modulate local inflammation. In osteoarthritis, the infrapatellar fat pad, characterized by immune cell infiltration and activation of nuclear factor- κ B (NF- κ B) signalling, has been implicated in knee joint inflammation and pain^{116,117}. Although analogous data are lacking in PsA, intra-articular adipose tissue depots might also contribute to disease pathogenesis through local paracrine immune–metabolic crosstalk.

Obesity and biomechanical stress in PsA

A compelling mechanistic link between obesity and PsA involves mechano-inflammation at the synovio-entheseal complex. Enthesitis, a hallmark lesion in PsA, is considered the primary site of inflammation¹¹⁸. Epidemiological evidence supports the contribution of biomechanical

stress to the onset of PsA^{119,120}. The term ‘deep Koebner phenomenon’ was introduced to describe how repetitive microtrauma in the joints and entheses (from, for example, occupational strain) can trigger PsA, highlighting the role of biomechanical stress in disease initiation¹²¹.

Data from both animal and human studies support the role of mechanical strain in enthesal inflammation. Under mechanical load, tenocytes and local mononuclear cells release pro-inflammatory mediators, including IL-1 β , CCL2 and CXCL1, and also matrix metalloproteinases. Although these responses are essential for tissue repair, persistent or excessive loading, as observed in obesity, might result in maladaptive responses, loss of resolution and transition to chronic inflammation, which are key features of PsA^{122,123}.

Importantly, obesity is associated with both increased mechanical load and systemic pro-inflammatory effects, confounding the distinction between mechanical, metabolic and immune-mediated enthesopathies. This overlap presents a diagnostic challenge, as mechanical and inflammatory enthesitis can appear similar on clinical and imaging assessments. Nonetheless, studies consistently show that obesity is associated with enthesal pathology both in individuals with PsA and in those without PsA^{34,124,125}. For instance, in a rat model of diet-induced obesity, excessive adiposity impaired enthesal healing after surgery and increased tendon inflammation¹²⁶. In humans, individuals with obesity exhibited a greater burden of sonographic enthesal abnormalities, underscoring the mechanical contribution¹²⁴. Among patients with PsA, BMI correlates with higher ultrasound scores for both active enthesitis and structural changes such as new bone formation³⁴.

Reducing biomechanical stress, or ‘off-loading’, might represent a therapeutic strategy for enthesal inflammation. In mouse models, limb suspension to reduce mechanical load attenuated enthesal inflammation and structural damage¹²⁷. These findings, alongside

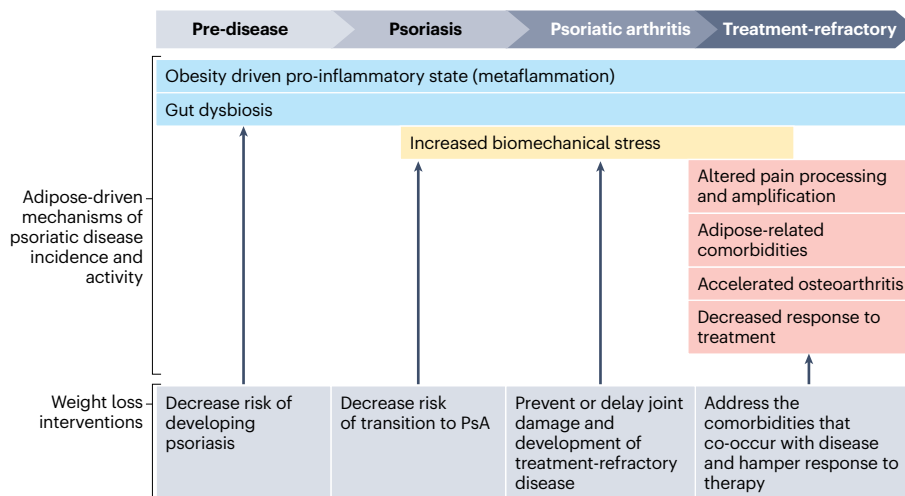


Fig. 2 | Obesity-driven mechanisms across stages of psoriatic disease. Obesity-driven mechanisms probably have a role throughout the continuum of psoriatic disease. The overall pro-inflammatory state, gut dysbiosis and increased biomechanical stress are probably both drivers and modifiers of disease. Obesity-driven changes in pain perceptions, comorbidity burden and response to treatment also function as amplifiers of disease. Given the effect of obesity throughout the disease course, weight loss interventions might be integral in altering the trajectory for patients with psoriatic disease. Prior to disease onset, weight loss can decrease the risk of developing psoriasis and psoriatic arthritis (PsA). Basic and clinical data support a causal relationship

between obesity and PsA, which is exemplified by the ability of bariatric surgery to reduce psoriatic disease risk. Early PsA might also present a ‘window of opportunity’ during which aggressive intervention, including weight loss, might substantially alter long-term outcomes. Lastly, after the development of PsA, weight loss is still vital as it can address and/or reverse altered pain processing and amplification, adipose-related comorbidities (such as fibromyalgia and sleep apnoea), accelerated osteoarthritis and decreased response to therapy. Glucagon-like peptide-1 (GLP-1)-based therapies might be of particular interest as a method for weight loss, along with lifestyle modifications, as preliminary data indicate direct anti-inflammatory and pain-modulating effects.

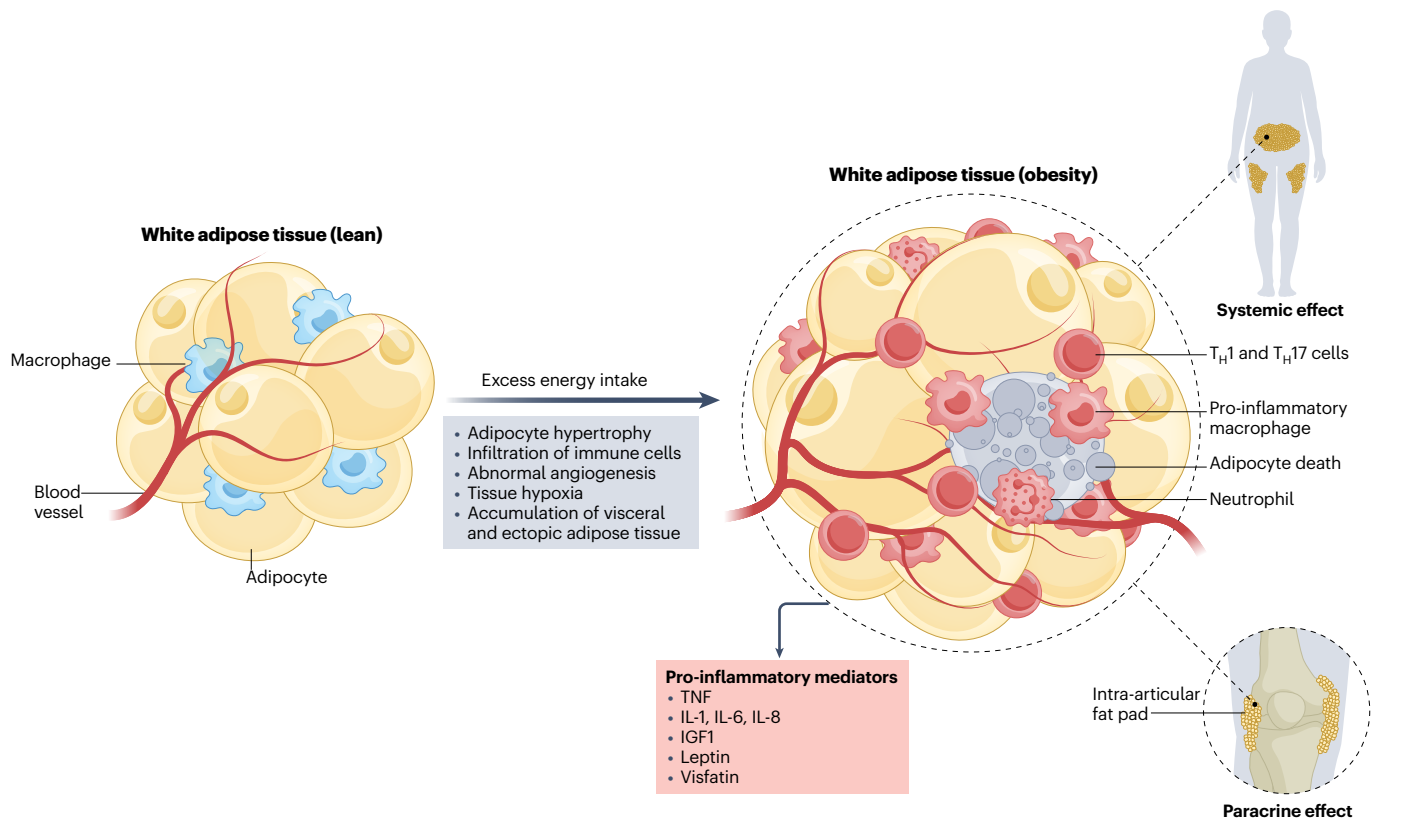


Fig. 3 | Obesity-induced modification of white adipose tissue. Excess energy intake results in the modification of white adipose tissue through adipocyte hypertrophy, infiltration of immune cells, abnormal angiogenesis, tissue hypoxia and the accumulation of visceral and ectopic adipose tissue. These changes promote the activation of T helper 1 (T_H1) and T_H17 cells, skew macrophages

towards a pro-inflammatory phenotype and the secretion of pro-inflammatory mediators such as TNF, IL-6, IL-1, IL-8, leptin, visfatin and insulin-like growth factor 1 (IGF1). These effects are exerted both systemically and locally in the joints via paracrine signalling from the intra-articular fat pad.

clinical data that link weight reduction to improvements in PsA outcomes, highlight weight loss as a promising intervention that targets biomechanical drivers of disease.

Gut dysbiosis in obesity and PsA

The gut microbiota has a crucial role in regulating host metabolism, immune homeostasis and barrier function. Abnormal gut microbial composition, termed gut dysbiosis, has been implicated in the pathogenesis of both obesity and PsA^{128,129} (Fig. 4).

The key role of the gut microbiota in obesity is shown in germ-free mice that are resistant to diet-induced weight gain, which indicates that specific gut microbial communities promote energy harvest and lipid storage¹³⁰. Similarly, the immunological relevance of gut microbial signals is demonstrated using HLA-B27 transgenic rats, which develop spondyloarthritis only in the presence of intestinal microbiota¹³¹. Gut dysbiosis is observed in patients with PsA, suggesting a potential role in triggering or perpetuating disease^{128,132}.

Gut dysbiosis disrupts the production of microbial metabolites and impairs regulatory host–microorganism interactions, which contributes to both metabolic imbalance and immune dysregulation¹³³. A central mechanism by which gut dysbiosis might contribute to both obesity and PsA is intestinal barrier dysfunction, commonly referred to as a ‘leaky gut’^{128,129}. Diets that are high in fat and low in fibre, typically associated

with obesity, favour microbial taxa that downregulate epithelial tight junction proteins, thereby increasing intestinal permeability^{134,135}. This increased permeability enables translocation of bacteria and microbial components, such as lipopolysaccharide (LPS), into the systemic circulation, a phenomenon termed ‘metabolic endotoxaemia’. In obesity, increased levels of circulating LPS activates Toll-like receptor 4 (TLR4) on immune cells, which promotes pro-inflammatory cytokine production and contributes to systemic inflammation and insulin resistance^{136,137}.

In PsA, microbial translocation might also drive inflammation via activation of pattern recognition receptors and inflammasomes in gut-associated antigen-presenting cells, leading to increased IL-23 production and T_H17 cell expansion¹³⁸. Specific microbial peptides might be presented by disease-associated HLA alleles (such as HLA-B27 or HLA-C06) potentially activating autoreactive T cells^{132,139}. Although it remains unclear if systemic T_H17 cell activation in PsA is microbiota-dependent, these mechanisms suggest overlapping pathways with obesity-related metaflammation.

Microbial metabolites might also link gut dysbiosis with PsA and obesity. These metabolites, particularly short-chain fatty acids (SCFAs) and bile acids, function as key mediators of gut–host communication¹⁴⁰. Diets that are high in fat might alter the gut microbiome by reducing SCFA-producing bacterial taxa; this depletion is also observed in people with PsA¹³³. SCFAs, especially butyrate and propionate, have

immunoregulatory effects; they promote T_{reg} cell differentiation and suppress NF- κ B-driven inflammation¹⁴¹. Thus, reduction of these SCFAs might contribute to loss of immune tolerance in both conditions.

Bile acid metabolites might also mediate the effect of microbial composition on systemic immune–metabolic status. Gut bacteria convert primary bile acids into secondary bile acids, which signal through host receptors (such as farnesoid X receptor (FXR) and Takeda G-protein-coupled receptor 5 (TGR5)) to regulate lipid metabolism, glucose homeostasis and immune responses^{129,138}. Gut dysbiosis alters the bile acid pool in obesity, shifting the profile towards pro-inflammatory species^{142,143}. Altered bile acid signatures have been reported in patients with psoriasis and PsA, which were associated with both cutaneous and musculoskeletal disease severity^{142,144}. Together, these data suggest that gut microbiota-driven metabolite imbalances that involve SCFAs and bile acids modulate inflammation and metabolism in PsA and obesity.

Although therapeutic strategies that target the microbiome, such as probiotics, prebiotics, dietary interventions and faecal microbiota transplantation, have shown promise in obesity^{145,146}, results in psoriatic disease remain limited and inconsistent. Trials in psoriasis and PsA using microbiota-modifying approaches have not yet demonstrated robust or durable clinical benefit, probably owing to the complexity and individual variability of host–microbiome interactions^{147,148}.

In summary, gut dysbiosis constitutes a shared pathophysiological mechanism that links obesity and PsA. By impairing barrier function, altering metabolite profiles (such as SCFAs and bile acids) and promoting IL-23– T_H17 cell immune activation, gut dysbiosis contributes to systemic inflammation and metabolic dysfunction common to both conditions.

Obesity and pain amplification

Individuals with obesity frequently report higher pain sensitivity than those with normal weight¹⁴⁹. People with PsA who also live with obesity often experience disproportional levels of pain and physical disability, which might not be fully explained by objective inflammation or joint damage^{14,150}. Similar observations in osteoarthritis led to extensive research that highlighted the role of obesity and adipose tissue dysfunction in amplifying pain independently of structural joint changes. Although specific data in PsA are limited, following studies in knee osteoarthritis, several mechanisms through which obesity can exacerbate pain have been proposed, including increased biomechanical stress, adipose-derived inflammatory mediators and neurotrophic modulation of peripheral and central sensitization pathways¹⁵¹.

Dysfunctional adipose tissue promotes a state of chronic low-grade inflammation that activates Toll-like receptor pathways in the synovium and subchondral bone and sensitizes local nociceptors. Leptin has also

been implicated in pain modulation¹⁵². In osteoarthritis, both serum and synovial leptin levels correlate with pain severity independently of BMI¹⁵². Local adipose tissue depots also contribute to pain severity. The infrapatellar fat pad, which is richly innervated by substance P-positive fibres, exhibits inflammation, fibrosis and increased leptin expression^{116,153}. In humans, MRI-detected alterations in the infrapatellar fat pad have been associated with greater knee pain, underscoring the paracrine effects of peri-synovial adipose tissue on nociception¹⁵⁴.

Obesity also affects neurosensory circuits, promoting increased pain sensitivity through neurotrophic remodelling. Circulating levels of neuropeptides, including substance P, neuropeptide Y, nerve growth factor (NGF) and brain-derived neurotrophic factor (BDNF), are increased in obesity^{155–157}. These molecules lower pressure pain thresholds and enhance temporal summation via activation of receptors (such as transient receptor potential vanilloid 1 (TRPV1) and tyrosine kinase A) on peripheral nociceptors and dorsal root ganglia^{156,157}. Psychophysical testing in individuals with obesity reveals widespread hyperalgesia and features of central sensitization. Importantly, substantial weight loss, either through bariatric surgery, pharmacological therapy or calorie restriction, can reduce pain levels and reduce spinal nociceptive facilitation, suggesting a direct link between excess adiposity and neural amplification of pain^{158–160}.

Obesity-related comorbidities and PsA outcomes

Obesity is a well-established risk factor for a range of comorbid conditions that might exacerbate symptom burden in patients with PsA, independently of underlying musculoskeletal inflammation. These comorbidities include T2DM, congestive heart failure, depression, obstructive sleep apnoea and osteoarthritis, each of which can contribute to chronic pain, stiffness and fatigue^{161,162}. Such overlapping symptomatology might confound the clinical assessment of PsA disease activity and artificially inflate composite outcome measures, even in the absence of active musculoskeletal inflammation. Moreover, obesity and obesity-associated comorbidities are often accompanied by adverse alterations in body composition, including sarcopenia and an increased adipose tissue-to-muscle ratio, which further impair physical function and limit mobility¹⁶³. These non-inflammatory drivers of symptomatology underscore the need for comprehensive assessment strategies in individuals with PsA and obesity.

Weight loss interventions in psoriatic disease

Although weight loss is an effective strategy in the treatment of psoriatic disease, the most common means of weight loss (dietary interventions or bariatric surgery) present challenges. Diet and lifestyle

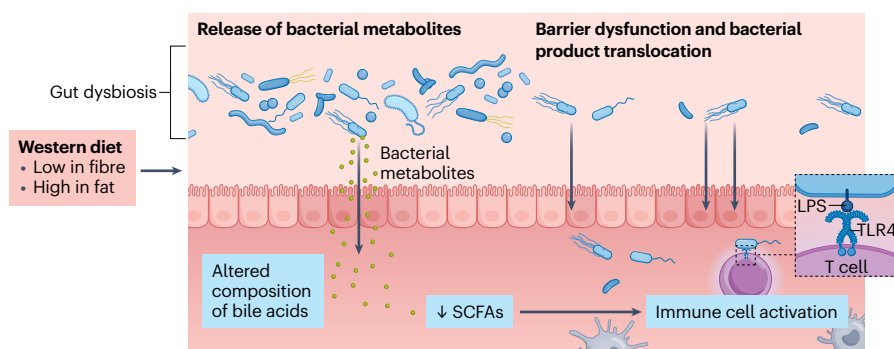


Fig. 4 | Gut dysbiosis in obesity and psoriatic arthritis. Gut dysbiosis has been implicated in both psoriatic arthritis and obesity. A Western diet could affect the gut microbiome composition in turn resulting in abnormal production of metabolites (such as short-chain fatty acids (SCFAs) and bile acids) and subsequent immune cell activation. This diet also contributes to gut barrier dysfunction and interaction of bacterial components with immune cells. LPS, lipopolysaccharide; TLR4, Toll-like receptor 4.

changes can be successful, but often require frequent, intensive interventions, with only around 50–60% of patients achieving their goal weight^{164,165}. Bariatric surgery, including gastric bypass and banding, is very effective, with over 60% of patients maintaining weight loss for over 10 years¹⁶⁶; however, both early complications (bowel obstruction, thromboembolism, gastrointestinal bleed, wound infection and hernia) and late complications (bowel strictures, ulcerations, nutritional deficiencies and malabsorption) can occur, limiting the widespread use of these interventions¹⁶⁷.

Effect of diet-induced weight loss in psoriatic disease

Diet-induced weight loss improves skin psoriasis in individuals with obesity^{168–171}. Although not uniformly observed across all studies, a meta-analysis of seven randomized control trials (RCTs) found that individuals receiving a weight loss intervention had lower PASI scores than those receiving usual care or not receiving a weight loss intervention (–2.5; 95% confidence interval, –3.90 to –1.08) and were almost three times more likely to achieve a PASI75 response^{168,172,173}. Even a limited weight loss might be beneficial for psoriasis outcomes. Individuals with psoriasis who are on a diet and exercise plan, compared with informative counselling alone, had higher reductions in PASI score (48% versus 26%) even though only 30% of the group reached the weight loss goal (≥5% body weight loss)¹⁷⁴.

Evidence also supports the efficacy of diet-induced weight loss in the treatment of PsA¹⁷⁵. In a non-randomized study in 46 individuals with obesity and PsA, a hypocaloric diet for 12–16 weeks resulted in a 19% median body weight loss. MDA increased from 29% to 54% with improvements in active enthesal and joint counts, inflammatory markers, skin psoriasis, physical function, perception of health, pain and fatigue¹⁷⁶. Sustained improvements were reported up to 24 months¹⁷⁷. In another study, 138 individuals with PsA and overweight or obesity on TNF inhibitors were randomized to a hypocaloric diet or a free self-managed diet. Weight loss, regardless of diet type, was associated with MDA in a dose-dependent manner. Compared with individuals who had a <5% body weight loss, individuals with 5–10% and >10% body weight loss were 3.8 times and 6.7 times more likely to achieve MDA, respectively. The DIPSA (Diet Interventions in Psoriatic Arthritis) study randomized 92 individuals with PsA and overweight or obesity who experienced residual musculoskeletal symptoms (Disease Activity in Psoriatic Arthritis (DAPSA) score of >10) to a Mediterranean diet, a low-calorie diet, or control with general dietary recommendations. Modest weight loss was noted across all three groups as well as a considerable improvement in DAPSA score, tender joint count, pain and fatigue; however, no statistically significant differences were found for change in weight and PsA parameters across the groups¹⁷⁸. Weight loss magnitude was associated with improvement in PsA outcomes regardless of dietary intervention¹⁷⁸.

Other smaller studies have also investigated the effect of specific diets for PsA. In one study, 20 participants on a very-low-calorie ketogenic diet showed improvement in DAPSA score that correlated with BMI reduction¹⁷⁹. In another study in 22 participants, a ketogenic diet was compared with a Mediterranean diet. Although the ketogenic diet was associated with greater improvement in PsA disease measures, the ketogenic diet also resulted in increased (although not statistically significant) weight loss compared with the Mediterranean diet¹⁸⁰. A study in 37 participants who practised intermittent fasting in the context of Ramadan for 1 month showed improvement in PsA outcomes despite limited or no weight loss¹⁸¹. However, in addition to their small size, these studies lack clarity regarding intervention implementation

and adherence. Overall, evidence supports the view that weight loss, rather than the type of diet itself, probably contributes to improvement of disease.

Importantly, given the overall increased risk of cardiovascular events in psoriatic disease¹⁸², dietary weight loss interventions have also been shown to improve cardiovascular outcomes in psoriatic disease. In an RCT including 60 individuals with psoriasis and obesity, participants were given either a low-energy diet or a normal diet. In addition to losing more weight, the intervention group had reduced blood pressure, resting heart rate, cholesterol and plasma glucose¹⁸³. Improvement in lipids, glucose and need for antihypertensive treatment was maintained after a 2-year follow-up of individuals with PsA who had achieved weight loss¹⁷⁷.

Effects of bariatric surgery in psoriatic disease

Bariatric surgery is another weight loss option that improves psoriatic disease outcomes. Case reports document complete resolution of psoriasis after Roux-en-Y gastric bypass surgery^{184–186}. A case series of 33 individuals with psoriasis and a BMI of ≥40 mg/m² undergoing bariatric procedures found that, after the procedure, 30% of patients were able to reduce their psoriasis medication and 25% had improvement in psoriasis severity, for which gastric bypass conferred more improvement than non-bypass procedures¹⁸⁷. In a small monocentre study in 86 participants with psoriatic disease, self-reported outcomes for both skin and joint activity improved 1 year after bariatric surgery, and this effect was most pronounced among those with severe disease at baseline¹⁸⁸.

Bariatric surgery might decrease the incidence of psoriatic disease, as shown in two large European population-based studies. Data from a Swedish registry showed that bariatric surgery (compared with usual care) was associated with a reduced incidence of psoriasis in individuals with obesity (hazard ratio 0.65) but did not affect the risk of PsA¹⁸⁹. In a Danish cohort, the risk of incident psoriasis was reduced with gastric bypass (hazard ratio 0.44), but not with gastric banding (hazard ratio 1.23), whereas the incidence of PsA was decreased by both procedures (hazard ratios 0.29 and 0.53)¹⁹⁰. These differential impacts of gastric bypass and banding might be because of effects beyond changes in adiposity. Although both procedures lead to similar levels of weight loss, gastric bypass also elicits endocrine changes. Specifically, gastric bypass increases postprandial secretion of the gut-derived hormone GLP-1, which is not affected by gastric banding^{191,192}. Therefore, the use of GLP-1-based therapies might mimic the effect of gastric bypass without the need to undergo surgery and could thus provide a new strategy for treating obesity in psoriatic disease.

Effect of GLP-1 based therapies

Case reports, case series and small cohort studies have shown the efficacy of GLP-1-based therapies in the treatment of skin psoriasis in individuals with concomitant T2DM^{193–197}. Small interventional studies have also generally shown positive results. In one study, 25 individuals with active plaque psoriasis and T2DM were randomized to acitretin alone or acitretin plus the GLP-1 receptor agonist (RA) liraglutide. Treatment with acitretin and liraglutide led to greater improvements in both PASI and Dermatology Life Quality Index (DLQI), more weight loss and improvement in cholesterol measures than treatment with acitretin alone. Although both groups showed decreases in levels of IL-17, IL-23, and TNF in psoriatic lesions after treatment, the liraglutide group had lower expression of IL-17 and IL-23 than the acitretin-alone group¹⁹⁸. Similarly, in an RCT in 31 individuals with psoriasis and T2DM

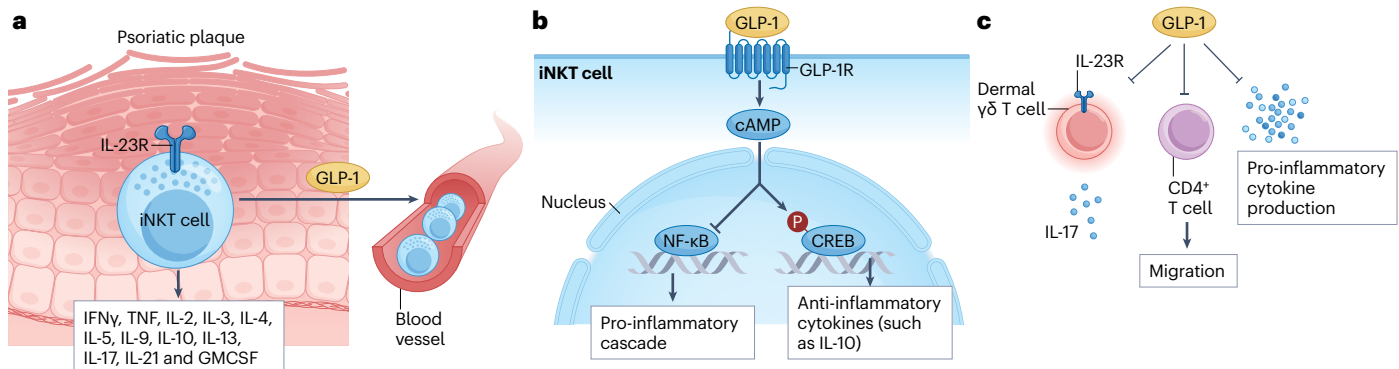


Fig. 5 | Direct anti-inflammatory and immunological effects of GLP-1-based therapies in psoriatic disease. Glucagon-like peptide-1 (GLP-1) receptor agonists might have direct anti-inflammatory effects through multiple mechanisms. **a**, Invariant natural killer T (iNKT) cells are implicated in the pathogenesis of psoriasis and have several immunoregulatory effector functions, including the rapid production of cytokines. In response to GLP-1-based therapies, iNKT cells move from psoriatic plaques to the periphery, thereby reducing the severity of psoriasis. **b**, GLP-1 receptors (GLP-1Rs) are located on iNKTs. Stimulation of GLP-1R upregulates cAMP, leading to the inhibition of nuclear factor- κ B (NF- κ B) and its subsequent pro-inflammatory cascade

and the phosphorylation of cAMP response element-binding protein (CREB), which in turn upregulates anti-inflammatory pathways. **c**, IL-23-responsive, IL-17-producing dermal $\gamma\delta$ T cells are implicated in psoriasis pathogenesis and are found in high numbers in psoriatic plaques. GLP-1 therapies decrease the percentage of dermal $\gamma\delta$ T cells and reduce IL-17 expression. In addition, GLP-1 therapies inhibit CD4⁺ T cell migration via effects on phosphoinositide 3-kinase and stromal cell-derived factor 1, and decrease overall production of pro-inflammatory cytokines including TNF, GM-CSF, granulocyte-macrophage colony-stimulating factor; IL-23R, IL-23 receptor.

that compared the GLP-1RA semaglutide with usual care, those receiving semaglutide had improved PASI scores and reduced DLQI, serum IL-6 and C-reactive protein (CRP)¹⁹⁹. Conversely, in a study in individuals with obesity, but not T2DM, no differences in PASI, DLQI or CRP were found in patients taking liraglutide compared with the placebo group, despite weight loss²⁰⁰. Clinical data regarding PsA outcomes are limited, but a case series from North America demonstrated an association between GLP-1-based therapies and clinically meaningful weight loss and improvement in CRP and pain²⁰¹.

Importantly, improvement in skin involvement can precede weight loss with the use of GLP-1-based therapies^{194,202}. Individuals with psoriasis on liraglutide show improvements in PASI, CRP and quality of life independent of weight loss²⁰³. This improvement is probably a consequence of direct anti-inflammatory and immunoregulatory effects of GLP-1-IRA on key psoriatic disease pathways, including invariant natural killer T cells, dermal $\gamma\delta$ T cells, lymphocyte migration and pro-inflammatory cytokines^{193,204–210} (Fig. 5).

GLP-1-based therapies have already shown efficacy in osteoarthritis, which can occur concomitantly with psoriasis and PsA. In the STEP 9 trial, once-weekly treatment with semaglutide was associated with considerable reduction in body weight and pain related to knee osteoarthritis²¹¹. Although the pain reduction might have been solely owing to the observed weight loss, mechanistic studies have shown the presence of GLP-1 receptors in the synovial membrane in individuals with osteoarthritis, and GLP-1-based therapies might therefore suppress degradation of the joint^{212–214}. Given the effects of both weight loss and possible direct anti-inflammatory and immunomodulating effects of GLP-1-based therapies, they might represent a unique adjuvant therapy for psoriatic disease, but evidence remains preliminary.

GLP-1-based therapies lead to consistent improvement of cardiovascular outcomes in the general population^{215,216}. Furthermore, a 2025 study using the TriNetX database showed that GLP-1-IRA use in individuals with psoriasis is associated with a statistically significant decrease in

all-cause mortality and risk of major adverse cardiac events²¹⁷. Interestingly, the risk reduction is higher in individuals with psoriasis than in those without psoriasis²¹⁷, which possibly indicates an added benefit in inflammatory disease. Decreased rates of mortality and major adverse cardiac events have similarly been shown in individuals with PsA treated with GLP-1-IRAs²¹⁸.

Other FDA-approved weight loss medications include orlistat (a lipase inhibitor), phentermine–topiramate (a combination sympathomimetic and anticonvulsant that works by blocking voltage-gated sodium channels) and naltrexone–bupropion (a combination opioid antagonist and antidepressant), but are generally less effective than GLP-1-based therapies²¹⁹. Thus far, no studies have assessed these drugs for psoriatic disease or inflammation, but small case series and case reports provide limited insights. In a case series of seven individuals with psoriasis and concomitant mood disorders, treatment with topiramate monotherapy resulted in improved psoriatic skin activity and weight loss²²⁰. Conversely, case reports have revealed that naltrexone–bupropion might even flare psoriatic skin disease^{221,222}. Furthermore, the extensive cardiovascular benefits reported with GLP-1-based therapies are not observed with the other approved weight loss drugs²²³; however, orlistat has shown mild cardiovascular benefits²²⁴, and phentermine–topiramate and naltrexone–bupropion have shown mixed results regarding their effect on cardiovascular risk and disease^{225–228}.

Unmet needs and future research

The relationship between obesity and the development, severity and therapeutic responsiveness of PsA is now well established. Obesity is not merely a comorbidity but exerts a pathogenic influence on disease phenotype. The emergence of GLP-1-based therapies has introduced a promising pharmacological strategy to address both obesity and the associated state of metabolic inflammation in PsA²²⁹. Although traditional interventions, such as hypocaloric diets and bariatric surgery, remain viable options, the former often yields only modest and usually

Box 1 | Proposed approach to obesity management in psoriatic disease

Addressing overweight and obesity needs to be part of a systematic approach to psoriatic disease management and should start with the rheumatologist. From there, an interdisciplinary team should be formed to help patients achieve their weight loss goals, including nutritionists, primary health-care providers, advanced practice providers, cardiologists, endocrinologists and/or bariatric surgeons. All individuals with overweight and obesity with a BMI of $>25\text{ mg/m}^2$ should be referred to a nutritionist for diet counselling. In addition, if an individual with obesity has a BMI of $>27\text{ mg/m}^2$ plus comorbidities or a BMI of $>30\text{ mg/m}^2$, pharmaceutical intervention should be considered. We propose that GLP-1-based therapies should be considered first if using pharmaceutical interventions, along with lifestyle changes such as diet and exercise. Bariatric surgery should be considered for individuals with a BMI of $\geq 35\text{ mg/m}^2$ plus comorbidities or a BMI of $>40\text{ mg/m}^2$ with or without pharmacological interventions.

unsustainable weight loss, whereas the latter, though more effective, is invasive and not without procedural risk.

By contrast, GLP-1-based therapies offer a non-invasive, pharmacologically effective and safe approach to achieving meaningful weight reduction in individuals with a broad spectrum of body mass indices. These agents might confer additional benefits beyond weight loss, including anti-inflammatory effects, pain reduction and cardiometabolic protection, making them particularly attractive for use in individuals with PsA^{230,231}. Although clinical evidence is limited, small studies have suggested that targeting obesity through GLP-1-based therapies in adiposity-driven conditions such as Ps, might function not only as a symptomatic intervention, but also as a disease-modifying adjunct that potentially influences disease progression and enhances therapeutic response. Additional high-quality evidence is needed to address this hypothesis. The TOGETHER-PsO (NCT06588283) and TOGETHER-PsA (NCT06588296) trials are investigating treatment with ixekizumab and tirzepatide compared with ixekizumab alone in psoriasis and PsA, respectively. The TOGETHER-AMPLIFY-PsO (NCT06857942) and TOGETHER-AMPLIFY-PsA (NCT06864026) trials are currently evaluating the efficacy of adding tirzepatide for individuals with psoriasis or PsA who are already on ixekizumab. Registries and observational cohorts are also underway.

Although the general health benefits of weight loss in obesity are well substantiated, the high cost of GLP-1-based therapies and limited access to comprehensive obesity care programmes raise important questions about the optimal timing and patient selection for weight-loss interventions within PsA management paradigms. Most individuals with overweight or obesity would probably benefit from GLP-1-based therapies, but three populations might be of particular interest: individuals with psoriasis who are at high risk of progression to synovio-entheseal disease; patients who are early in their disease course; and patients who meet the definition of treatment-refractory or difficult-to-manage PsA²³² (Fig. 2).

Despite their potential, GLP-1-based therapies are not without limitations. Gastrointestinal adverse effects, especially nausea and vomiting, are common and might affect adherence²³³. These symptoms overlap with known adverse effect profiles of commonly prescribed

DMARDs, such as methotrexate, apremilast and leflunomide, potentially complicating attribution and management^{234–236}. Concerns also exist regarding unintended loss of lean muscle mass, which might be particularly relevant in the context of pre-existing sarcopenia (a common, under-recognized issue in PsA¹⁶³). Thus, real-world data on tolerability, safety and outcomes in individuals with psoriatic disease are essential. In addition, interdisciplinary care models, including allied health professional and obesity specialists are required to optimize the implementation of GLP-1-based therapy. Close monitoring, individualized dose titration and comprehensive patient education are key to mitigating risks and enhancing outcomes.

The mechanistic basis by which weight loss improves PsA outcomes remains incompletely understood. Interventional studies offer a unique opportunity to investigate candidate pathways, including changes in adipokine signalling, metabolic inflammation, immune cell function, pain sensitization and more. Integration of translational research methodologies, such as advanced imaging, tissue biomarker profiling and multi-omics, will be necessary to delineate the relative contributions of these mechanisms.

Proposed strategies to address obesity in psoriatic disease

Despite robust data indicating the potential of weight loss to enhance therapeutic efficacy and expert panel consensus that it should be an essential part of psoriatic disease management^{237,238}, weight loss is not currently systematically incorporated into PsA treatment algorithms. Although weight is often a sensitive topic, the introduction of obesity as part of psoriatic disease management must begin with dermatologists and rheumatologists (Box 1). Primary rheumatological and/or dermatological intervention should be followed by the involvement of interdisciplinary health-care providers including family physicians, cardiologists, endocrinologists, and even bariatric surgeons and medical nutritionists, to help patients achieve optimal weight goals. For individuals considering pharmacological interventions, we propose that GLP-1 therapeutic agents should be considered before other weight loss therapies, given their proven efficacy and safety and potentially synergistic effects.

Conclusions

Obesity is a chronic, inflammatory condition that interacts inextricably with psoriatic disease. Incorporating structured obesity interventions, especially for selected high-risk individuals, represents a promising frontier in PsA management. Strategic, pragmatic trials are needed to inform the development of future clinical practice guidelines that incorporate weight management not simply as a lifestyle recommendation, but as a disease-modifying adjunctive therapy in psoriatic disease. Real-world evidence on the short-term and long-term effectiveness of these therapies, predictors of optimal treatment outcomes and cost-effective therapies are urgently needed for individuals with psoriatic disease. Finally, future research that incorporates underlying molecular and immune features is needed to understand the in-depth mechanisms of the obesity–psoriatic disease axis.

Published online: 24 November 2025

References

1. Armstrong, A. W. et al. Psoriasis prevalence in adults in the United States. *JAMA Dermatol.* **157**, 940–946 (2021).
2. Ritchlin, C. T., Colbert, R. A. & Gladman, D. D. Psoriatic arthritis. *N. Engl. J. Med.* **376**, 957–970 (2017).
3. Schett, G. et al. Psoriatic arthritis from a mechanistic perspective. *Nat. Rev. Rheumatol.* **18**, 311–325 (2022).

4. Wharton, S. et al. Obesity in adults: a clinical practice guideline. *CMAJ* **192**, E875–E891 (2020).
5. [No authors listed] Clinical guidelines on the identification, evaluation, and treatment of overweight and obesity in adults — the evidence report. National Institutes of Health. *Obes. Res.* **6**, 51s–209s (1998).
6. Neimann, A. L. et al. Prevalence of cardiovascular risk factors in patients with psoriasis. *J. Am. Acad. Dermatol.* **55**, 829–835 (2006).
7. Phan, C. et al. Metabolic comorbidities and hypertension in psoriasis patients in France. Comparisons with French national databases. *Ann. Dermatol. Venereol.* **143**, 264–274 (2016).
8. Eder, L., Abji, F., Rosen, C. F., Chandran, V. & Gladman, D. D. The association between obesity and clinical features of psoriatic arthritis: a case-control study. *J. Rheumatol.* **44**, 437–443 (2017).
9. Labitigan, M. et al. Higher rates and clustering of abnormal lipids, obesity, and diabetes mellitus in psoriatic arthritis compared with rheumatoid arthritis. *Arthritis Care Res.* **66**, 600–607 (2014).
10. Stirnimann, M. et al. Obesity in psoriatic arthritis is increasingly affecting men and appears less dependent of socioeconomic status than in the general population [abstract OP0064]. *Ann. Rheum. Dis.* **82**, 44 (2023).
11. Langan, S. M. et al. Prevalence of metabolic syndrome in patients with psoriasis: a population-based study in the United Kingdom. *J. Invest. Dermatol.* **132**, 556–562 (2012).
12. Ishchenko, A., Pazmino, S., Neerinx, B., Lories, R. & de Vlam, K. Comorbidities in early psoriatic arthritis: data from the Metabolic Disturbances in Psoriatic Arthritis Cohort Study. *Arthritis Care Res.* **76**, 231–240 (2024).
13. Scriffignano, S. et al. Cardiovascular comorbidities in psoriatic arthritis: epidemiology and risk factors in two different European populations. *Clin. Exp. Rheumatol.* **41**, 1815–1822 (2023).
14. Eder, L., Thavaneswaran, A., Chandran, V., Cook, R. J. & Gladman, D. D. Obesity is associated with a lower probability of achieving sustained minimal disease activity state among patients with psoriatic arthritis. *Ann. Rheum. Dis.* **74**, 813–817 (2015).
15. Jacobi, A., Langenbruch, A., Purwins, S., Augustin, M. & Radtke, M. A. Prevalence of obesity in patients with psoriasis: results of the national study PsoHealth3. *Dermatology* **231**, 231–238 (2015).
16. Schwarz, P. E. H., Pinter, A., Melzer, N., Bartczek, P. & Reinhardt, M. ERAPSO: revealing the high burden of obesity in German psoriasis patients. *Dermatol. Ther.* **9**, 579–587 (2019).
17. Armstrong, A. W., Harskamp, C. T. & Armstrong, E. J. The association between psoriasis and obesity: a systematic review and meta-analysis of observational studies. *Nutr. Diabetes* **2**, e54 (2012).
18. Queiro, R., Lorenzo, A., Tejón, P., Coto, P. & Pardo, E. Obesity in psoriatic arthritis: comparative prevalence and associated factors. *Medicine* **98**, e16400 (2019).
19. Toussiro, E. et al. Visceral adiposity in patients with psoriatic arthritis and psoriasis alone and its relationship with metabolic and cardiovascular risk. *Rheumatology* **60**, 2816–2825 (2020).
20. Snevik, I. et al. Obesity, Waist Circumference, weight change, and risk of incident psoriasis: prospective data from the HUNT study. *J. Invest. Dermatol.* **137**, 2484–2490 (2017).
21. Kumar, S., Han, J., Li, T. & Qureshi, A. A. Obesity, waist circumference, weight change and the risk of psoriasis in US women. *J. Eur. Acad. Dermatol. Venereol.* **27**, 1293–1298 (2013).
22. Setty, A. R., Curhan, G. & Choi, H. K. Obesity, waist circumference, weight change, and the risk of psoriasis in women: Nurses' Health Study II. *Arch. Intern. Med.* **167**, 1670–1675 (2007).
23. Danielsen, K., Wilsgaard, T., Olsen, A. O. & Furberg, A. S. Overweight and weight gain predict psoriasis development in a population-based cohort. *Acta Derm. Venereol.* **97**, 332–339 (2017).
24. Thomsen, R. S. et al. Adiposity and physical activity as risk factors for developing psoriatic arthritis: longitudinal data from a population-based study in Norway. *Arthritis Care Res.* **73**, 432–441 (2021).
25. Li, W., Han, J. & Qureshi, A. A. Obesity and risk of incident psoriatic arthritis in US women. *Ann. Rheum. Dis.* **71**, 1267–1272 (2012).
26. Lewinson, R. T. et al. Depression is associated with an increased risk of psoriatic arthritis among patients with psoriasis: a population-based study. *J. Invest. Dermatol.* **137**, 828–835 (2017).
27. Love, T. J. et al. Obesity and the risk of psoriatic arthritis: a population-based study. *Ann. Rheum. Dis.* **71**, 1273–1277 (2012).
28. Green, A. et al. Modifiable risk factors and the development of psoriatic arthritis in people with psoriasis. *Br. J. Dermatol.* **182**, 714–720 (2020).
29. Soltani-Arabshahi, R. et al. Obesity in early adulthood as a risk factor for psoriatic arthritis. *Arch. Dermatol.* **146**, 721–726 (2010).
30. Murray, M. L., Bergstresser, P. R., Adams-Huet, B. & Cohen, J. B. Relationship of psoriasis severity to obesity using same-gender siblings as controls for obesity. *Clin. Exp. Dermatol.* **34**, 140–144 (2009).
31. Duarte, G. V. et al. Association between obesity measured by different parameters and severity of psoriasis. *Int. J. Dermatol.* **52**, 177–181 (2013).
32. Huang, Y. H. et al. Relationships between obesity and the clinical severity of psoriasis in Taiwan. *J. Eur. Acad. Dermatol. Venereol.* **24**, 1035–1039 (2010).
33. Wang, H. et al. BMI matters: understanding the link between weight and severe psoriasis. *Sci. Rep.* **15**, 11158 (2025).
34. Eder, L. et al. Is the MADrid Sonographic Enthesitis Index useful for differentiating psoriatic arthritis from psoriasis alone and healthy controls? *J. Rheumatol.* **41**, 466–472 (2014).
35. Kim, K. J. et al. Serum leptin levels are associated with the presence of syndesmophytes in male patients with ankylosing spondylitis. *Clin. Rheumatol.* **31**, 1231–1238 (2012).
36. Kim, S. K., Choe, J. Y., Lee, S. S. & Shin, K. Body mass index is related with the presence of syndesmophyte in axial spondyloarthritis: data from the Korean College of Rheumatology BIOLogics (KOBIO) registry. *Mod. Rheumatol.* **27**, 855–861 (2017).
37. Maas, F. et al. Spinal radiographic progression in patients with ankylosing spondylitis treated with TNF- α blocking therapy: a prospective longitudinal observational cohort study. *PLoS ONE* **10**, e0122693 (2015).
38. Aydin, S. Z. et al. A relationship between spinal new bone formation in ankylosing spondylitis and the sonographically determined Achilles tendon enthesophytes. *Rheumatol. Int.* **36**, 397–404 (2016).
39. Leung, Y. Y. et al. Association between obesity and likelihood of remission or low disease activity status in psoriatic arthritis applying index-based and patient-based definitions of remission: a cross-sectional study. *RMD Open* **9**, e003157 (2023).
40. Walsh, J. A. et al. Measuring outcomes in psoriatic arthritis: comparing routine assessment of patient index data and psoriatic arthritis impact of disease. *J. Rheumatol.* **47**, 1496–1505 (2020).
41. Pirro, F. et al. Impact of body mass index on the efficacy of biological therapies in patients with psoriasis: a real-world study. *Clin. Drug. Investig.* **41**, 917–925 (2021).
42. Chiricozzi, A. et al. Real-life 9-year experience with adalimumab in psoriasis and psoriatic arthritis: results of a single-centre, retrospective study. *J. Eur. Acad. Dermatol. Venereol.* **31**, 304–311 (2017).
43. Demirel Ögüt, N., Koç Yıldırım, S., Erbağcı, E. & Hapa, F. A. Ixekizumab treatment in patients with moderate-to-severe plaque psoriasis in a real-world clinical setting. *J. Cosmet. Dermatol.* **21**, 6215–6224 (2022).
44. Rompoti, N. et al. Real-world data from a single Greek centre on the use of secukinumab in plaque psoriasis: effectiveness, safety, drug survival, and identification of patients that sustain optimal response. *J. Eur. Acad. Dermatol. Venereol.* **34**, 1240–1247 (2020).
45. Naldi, L. et al. Impact of body mass index and obesity on clinical response to systemic treatment for psoriasis. Evidence from the Psocare project. *Dermatology* **217**, 365–373 (2008).
46. Enos, C. W. et al. Comorbid obesity and history of diabetes are independently associated with poorer treatment response to biologics at 6 months: a prospective analysis in Corrona Psoriasis Registry. *J. Am. Acad. Dermatol.* **86**, 68–76 (2022).
47. Gkalpakiotis, S. et al. Risankizumab for the treatment of moderate-to-severe psoriasis: real-life multicenter experience from the Czech Republic. *Dermatol. Ther.* **11**, 1345–1355 (2021).
48. Narcisi, A. et al. Real-life effectiveness of tildrakizumab in chronic plaque psoriasis: a 52-week multicenter retrospective study-IL PSO (Italian landscape psoriasis). *J. Eur. Acad. Dermatol. Venereol.* **37**, 93–103 (2023).
49. Caldarella, G. et al. Risankizumab for the treatment of moderate-to-severe psoriasis: a multicenter, retrospective, 1 year real-life study. *Dermatol. Ther.* **35**, e15489 (2022).
50. Malara, G. et al. Effectiveness of apremilast in real life in patients with psoriasis: a longitudinal study. *Acta Derm. Venereol.* **101**, adv00545 (2021).
51. Langley, R. G. et al. Efficacy of apremilast in patients with moderate-to-severe plaque psoriasis across weight and BMI subgroups: analysis from ESTEEM 1 and ESTEEM 2 [abstract 43250]. *J. Am. Acad. Dermatol.* **89**, AB158 (2023).
52. Kim, H. J., Lee, E. H. & Jung, Y. Association of obesity and innate immune markers with resistance to biologic therapy in psoriasis. *JAMA Dermatol.* **161**, 629–634 (2025).
53. Assan, F. et al. First-Line biologic therapy and obesity in moderate-to-severe psoriasis: results from the prospective multicenter cohort Psobioteq. *Dermatology* **237**, 338–346 (2021).
54. Warren, R. B. et al. Differential drug survival of biologic therapies for the treatment of psoriasis: a prospective observational cohort study from the British Association of Dermatologists Biologic Interventions Register (BADBIR). *J. Invest. Dermatol.* **135**, 2632–2640 (2015).
55. Mourad, A., Straube, S., Armijo-Olivo, S. & Gniadecki, R. Factors predicting persistence of biologic drugs in psoriasis: a systematic review and meta-analysis. *Br. J. Dermatol.* **181**, 450–458 (2019).
56. di Minno, M. N. et al. Obesity and the prediction of minimal disease activity: a prospective study in psoriatic arthritis. *Arthritis Care Res.* **65**, 141–147 (2013).
57. Vallejo-Yagüe, E., Burkard, T., Micheroli, R. & Burden, A. M. Minimal disease activity and remission in patients with psoriatic arthritis with elevated body mass index: an observational cohort study in the Swiss Clinical Quality Management cohort. *BMJ Open* **12**, e061474 (2022).
58. Singh, S. et al. Obesity and response to anti-tumor necrosis factor- α agents in patients with select immune-mediated inflammatory diseases: a systematic review and meta-analysis. *PLoS ONE* **13**, e0195123 (2018).
59. Højgaard, P. et al. The influence of obesity on response to tumour necrosis factor- α inhibitors in psoriatic arthritis: results from the DANBIO and ICEBIO registries. *Rheumatology* **55**, 2191–2199 (2016).
60. Ogdie, A. et al. Predictors of achieving remission among patients with psoriatic arthritis initiating a tumor necrosis factor inhibitor. *J. Rheumatol.* **46**, 475–482 (2019).
61. Queiro, R. & Cañete, J. D. Impact of cardiovascular risk factors on the achievement of therapeutic goals in psoriatic arthritis: is there any association? *Clin. Rheumatol.* **37**, 661–666 (2018).

62. Papp, K. et al. Apremilast, an oral phosphodiesterase 4 (PDE4) inhibitor, in patients with moderate to severe plaque psoriasis: results of a phase III, randomized, controlled trial (Efficacy and Safety Trial Evaluating the Effects of Apremilast in Psoriasis [ESTEEM] 1). *J. Am. Acad. Dermatol.* **73**, 37–49 (2015).
63. Yang, Z. S., Lin, N. N., Li, L. & Li, Y. The effect of TNF inhibitors on cardiovascular events in psoriasis and psoriatic arthritis: an updated meta-analysis. *Clin. Rev. Allergy Immunol.* **51**, 240–247 (2016).
64. Gisondi, P., Cotena, C., Tessari, G. & Girolomoni, G. Anti-tumour necrosis factor- α therapy increases body weight in patients with chronic plaque psoriasis: a retrospective cohort study. *J. Eur. Acad. Dermatol. Venereol.* **22**, 341–344 (2008).
65. Patsalos, O., Dalton, B., Leppanen, J., Ibrahim, M. A. A. & Himmerich, H. Impact of TNF- α inhibitors on body weight and BMI: a systematic review and meta-analysis. *Front. Pharmacol.* **11**, 481 (2020).
66. Haberman, R. H. et al. Burden and determinants of multi-b/tsDMARD failure in psoriatic arthritis. *Arthritis Res. Ther.* **27**, 46 (2025).
67. Stober, C. et al. Prevalence and predictors of tumour necrosis factor inhibitor persistence in psoriatic arthritis. *Rheumatology* **57**, 158–163 (2018).
68. Rida, M. A., Lee, K. A., Chandran, V., Cook, R. J. & Gladman, D. D. Persistence of biologics in the treatment of psoriatic arthritis: data from a large hospital-based longitudinal cohort. *Arthritis Care Res.* **75**, 2174–2181 (2023).
69. Bassi, M. & Singh, S. Impact of obesity on response to biologic therapies in patients with inflammatory bowel diseases. *BioDrugs* **36**, 197–203 (2022).
70. Leibold, M. et al. Impact of weight on the efficacy and safety of ustekinumab in patients with moderate to severe psoriasis: rationale for dosing recommendations. *J. Am. Acad. Dermatol.* **63**, 571–579 (2010).
71. Mortato, E. et al. Predictive factors for super responder status and long-term effectiveness of guselkumab in psoriasis: a multicenter retrospective study. *Dermatol. Ther.* **15**, 1239–1250 (2025).
72. Puig, L. Obesity and psoriasis: body weight and body mass index influence the response to biological treatment. *J. Eur. Acad. Dermatol. Venereol.* **25**, 1007–1011 (2011).
73. Lee, J. E. et al. Effect of body weight on risk-benefit and dosing regimen recommendation of secukinumab for the treatment of moderate to severe plaque psoriasis. *Clin. Pharmacol. Ther.* **106**, 78–80 (2019).
74. Di Minno, M. N. et al. Weight loss and achievement of minimal disease activity in patients with psoriatic arthritis starting treatment with tumour necrosis factor α blockers. *Ann. Rheum. Dis.* **73**, 1157–1162 (2014).
75. Keizer, R. J., Huitema, A. D., Schellens, J. H. & Beijnen, J. H. Clinical pharmacokinetics of therapeutic monoclonal antibodies. *Clin. Pharmacokinet.* **49**, 493–507 (2010).
76. Zhang, S. et al. The alterations in and the role of the Th17/Treg balance in metabolic diseases. *Front. Immunol.* **12**, 678355 (2021).
77. Liu, C. & Li, X. Role of leptin and adiponectin in immune response and inflammation. *Int. Immunopharmacol.* **161**, 115082 (2025).
78. Schleh, M. W. et al. Metaflammation in obesity and its therapeutic targeting. *Sci. Transl. Med.* **15**, eadf9382 (2023).
79. Hotamisligil, G. S. Inflammation, metaflammation and immunometabolic disorders. *Nature* **542**, 177–185 (2017).
80. Pedersen, S. D. et al. Pharmacotherapy for obesity management in adults: 2025 clinical practice guideline update. *CMAJ* **197**, E797–E809 (2025).
81. Bluhner, M. Obesity: global epidemiology and pathogenesis. *Nat. Rev. Endocrinol.* **15**, 288–298 (2019).
82. El-Sayed Moustafa, J. S. & Froguel, P. From obesity genetics to the future of personalized obesity therapy. *Nat. Rev. Endocrinol.* **9**, 402–413 (2013).
83. Hagberg, C. E. & Spalding, K. L. White adipocyte dysfunction and obesity-associated pathologies in humans. *Nat. Rev. Mol. Cell Biol.* **25**, 270–289 (2024).
84. Zwick, R. K., Guerrero-Juarez, C. F., Horsley, V. & Plikus, M. V. Anatomical, physiological, and functional diversity of adipose tissue. *Cell Metab.* **27**, 68–83 (2018).
85. Gomez-Ambrosi, J., Catalan, V. & Fruhbeck, G. The evolution of the understanding of obesity over the last 100 years. *Int. J. Obes.* **49**, 168–176 (2025).
86. Endo, Y. et al. Obesity drives Th17 cell differentiation by inducing the lipid metabolic kinase, ACC1. *Cell Rep.* **12**, 1042–1055 (2015).
87. Pugh, G. H. et al. T cells dominate peripheral inflammation in a cross-sectional analysis of obesity-associated diabetes. *Obesity* **30**, 1983–1994 (2022).
88. Weisberg, S. P. et al. Obesity is associated with macrophage accumulation in adipose tissue. *J. Clin. Invest.* **112**, 1796–1808 (2003).
89. Wculek, S. K., Dunphy, G., Heras-Murillo, I., Mastrangelo, A. & Sancho, D. Metabolism of tissue macrophages in homeostasis and pathology. *Cell Mol. Immunol.* **19**, 384–408 (2022).
90. Shaikh, S. R., Beck, M. A., Alwarawrah, Y. & MacIver, N. J. Emerging mechanisms of obesity-associated immune dysfunction. *Nat. Rev. Endocrinol.* **20**, 136–148 (2024).
91. Teixeira, A., Garrido, A., Ferrer, A., Perna, C. & Djouder, N. Inhibition of the IL-17A axis in adipocytes suppresses diet-induced obesity and metabolic disorders in mice. *Nat. Metab.* **3**, 496–512 (2021).
92. Tilg, H., Ianiro, G., Gasbarrini, A. & Adolph, T. E. Adipokines: masterminds of metabolic inflammation. *Nat. Rev. Immunol.* **25**, 250–265 (2025).
93. Kiernan, K. & MacIver, N. J. The role of the adipokine leptin in immune cell function in health and disease. *Front. Immunol.* **11**, 622468 (2020).
94. Fatica, M. et al. Metabolic challenges of glucose and lipid dysregulation in psoriatic arthritis: a narrative review from pathogenesis to clinical practice. *Acta Diabetol.* <https://doi.org/10.1007/s00592-025-02565-5> (2025).
95. Ferguson, L. D. et al. Psoriatic arthritis is associated with adverse body composition predictive of greater coronary heart disease and type 2 diabetes propensity — a cross-sectional study. *Rheumatology* **60**, 1858–1862 (2021).
96. Frouhi, N. G., Sattar, N. & McKeigue, P. M. Relation of C-reactive protein to body fat distribution and features of the metabolic syndrome in Europeans and South Asians. *Int. J. Obes. Relat. Metab. Disord.* **25**, 1327–1331 (2001).
97. Park, H. S., Park, J. Y. & Yu, R. Relationship of obesity and visceral adiposity with serum concentrations of CRP, TNF- α and IL-6. *Diabetes Res. Clin. Pract.* **69**, 29–35 (2005).
98. Zhao, S. S., Bowes, J., Barton, A., Davey Smith, G. & Richardson, T. Separating the effects of childhood and adult body size on inflammatory arthritis: a Mendelian randomisation study. *RMD Open* **8**, e002321 (2022).
99. Zhao, S. S., Yiu, Z. Z. N., Barton, A. & Bowes, J. Association of lipid-lowering drugs with risk of psoriasis: a Mendelian randomization study. *JAMA Dermatol.* **159**, 275–280 (2023).
100. Zhao, S. S. & Bowes, B. A. J. Impaired glycaemic control is associated with increased risk of psoriatic arthritis: Mendelian randomisation study [abstract POS0307]. *Ann. Rheum. Dis.* **81**, 403 (2022).
101. Jin, J. Q. et al. Mendelian Randomization studies in psoriasis and psoriatic arthritis: a systematic review. *J. Invest. Dermatol.* **143**, 762–776.e3 (2023).
102. Boehncke, W. H., Boehncke, S., Tobin, A. M. & Kirby, B. The ‘psoriatic march’: a concept of how severe psoriasis may drive cardiovascular comorbidity. *Exp. Dermatol.* **20**, 303–307 (2011).
103. Nakamizo, S. et al. High fat diet exacerbates murine psoriatic dermatitis by increasing the number of IL-17-producing $\gamma\delta$ T cells. *Sci. Rep.* **7**, 14076 (2017).
104. Higashi, Y. et al. High-fat diet exacerbates imiquimod-induced psoriasis-like dermatitis in mice. *Exp. Dermatol.* **27**, 178–184 (2018).
105. Zhang, Y. et al. Epidermal fatty acid binding protein promotes skin inflammation induced by high-fat diet. *Immunity* **42**, 953–964 (2015).
106. Toussiroot, E. Mini-Review: the contribution of adipokines to joint inflammation in inflammatory rheumatic diseases. *Front. Endocrinol.* **11**, 606560 (2020).
107. Eder, L. et al. Serum adipokines in patients with psoriatic arthritis and psoriasis alone and their correlation with disease activity. *Ann. Rheum. Dis.* **72**, 1956–1961 (2013).
108. Radic, M. et al. Cardiometabolic risk in psoriatic arthritis: a hidden burden of inflammation and metabolic dysregulation. *Metabolites* **15**, 206 (2025).
109. Kyriakou, A., Patsatsi, A., Sotiriadis, D. & Goulis, D. G. Serum leptin, resistin, and adiponectin concentrations in psoriasis: a meta-analysis of observational studies. *Dermatology* **233**, 378–389 (2017).
110. Cerman, A. A. et al. Serum leptin levels, skin leptin and leptin receptor expression in psoriasis. *Br. J. Dermatol.* **159**, 820–826 (2008).
111. Toledano, E. et al. Clinical features and disease activity in psoriatic arthritis: a sex-related perspective on leptin and comorbidity. *J. Clin. Med.* **13**, 2959 (2024).
112. Xue, Y. et al. Adipokines in psoriatic arthritis patients: the correlations with osteoclast precursors and bone erosions. *PLoS ONE* **7**, e46740 (2012).
113. Busso, N. et al. Leptin signaling deficiency impairs humoral and cellular immune responses and attenuates experimental arthritis. *J. Immunol.* **168**, 875–882 (2002).
114. Collins, K. et al. Adipose tissue is a critical regulator of osteoarthritis. *Proc. Natl. Acad. Sci. USA* **118**, e2021096118 (2021).
115. Porta, S. et al. Adipokines, cardiovascular risk, and therapeutic management in obesity and psoriatic arthritis. *Front. Immunol.* **11**, 590749 (2020).
116. Eymard, F. et al. Knee and hip intra-articular adipose tissues (IAATs) compared with autologous subcutaneous adipose tissue: a specific phenotype for a central player in osteoarthritis. *Ann. Rheum. Dis.* **76**, 1142–1148 (2017).
117. Zeng, N., Yan, Z. P., Chen, X. Y. & Ni, G. X. Infrapatellar fat pad and knee osteoarthritis. *Aging Dis.* **11**, 1317–1328 (2020).
118. Araujo, E. G. & Schett, G. Entesitis in psoriatic arthritis (part 1): pathophysiology. *Rheumatology* **59**, ii0–ii4 (2020).
119. Thorarensen, S. M. et al. Physical trauma recorded in primary care is associated with the onset of psoriatic arthritis among patients with psoriasis. *Ann. Rheum. Dis.* **76**, 521–525 (2017).
120. Eder, L. et al. Association between environmental factors and onset of psoriatic arthritis in patients with psoriasis. *Arthritis Care Res.* **63**, 1091–1097 (2011).
121. Hsieh, J., Kadavath, S. & Efthimiou, P. Can traumatic injury trigger psoriatic arthritis? A review of the literature. *Clin. Rheumatol.* **33**, 601–608 (2014).
122. Gracey, E. et al. Tendon and ligament mechanical loading in the pathogenesis of inflammatory arthritis. *Nat. Rev. Rheumatol.* **16**, 193–207 (2020).
123. Crowe, L. A. N. et al. Pathways driving tendinopathy and enthesitis: siblings or distant cousins in musculoskeletal medicine? *Lancet Rheumatol.* **5**, e293–e304 (2023).
124. Bakirci, S. et al. Enteseal changes in response to age, body mass index, and physical activity: an ultrasound study in healthy people. *J. Rheumatol.* **47**, 968–972 (2020).
125. Falsetti, P., Conticini, E., Baldi, C., Acciai, C. & Frediani, B. High prevalence of ultrasound-defined enthesitis in patients with metabolic syndrome. Comment on: How normal is the enthesitis by ultrasound in healthy subjects? Di Matteo et al. *Clin. Exp. Rheumatol.* **39**, 435–436 (2021).
126. Bolam, S. M. et al. Obesity impairs enthesitis healing after rotator cuff repair in a rat model. *Am. J. Sports Med.* **49**, 3959–3969 (2021).
127. Jacques, P. et al. Proof of concept: enthesitis and new bone formation in spondyloarthritis are driven by mechanical strain and stromal cells. *Ann. Rheum. Dis.* **73**, 437–445 (2014).

128. Scher, J. U. et al. Decreased bacterial diversity characterizes the altered gut microbiota in patients with psoriatic arthritis, resembling dysbiosis in inflammatory bowel disease. *Arthritis Rheumatol.* **67**, 128–139 (2015).
129. Cani, P. D. & Van Hul, M. Gut microbiota in overweight and obesity: crosstalk with adipose tissue. *Nat. Rev. Gastroenterol. Hepatol.* **21**, 164–183 (2024).
130. Backhed, F., Manchester, J. K., Semenkovich, C. F. & Gordon, J. I. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc. Natl Acad. Sci. USA* **104**, 979–984 (2007).
131. Taurog, J. D. et al. The germfree state prevents development of gut and joint inflammatory disease in HLA-B27 transgenic rats. *J. Exp. Med.* **180**, 2359–2364 (1994).
132. Myers, B. et al. The gut microbiome in psoriasis and psoriatic arthritis. *Best. Pract. Res. Clin. Rheumatol.* **33**, 101494 (2019).
133. Scher, J. U., Nayak, R. & Clemente, J. C. Microbiome research in autoimmune and immune-mediated inflammatory diseases: lessons, advances and unmet needs. *Ann. Rheum. Dis.* **84**, 9–13 (2025).
134. Everard, A. et al. Cross-talk between *Akkermansia muciniphila* and intestinal epithelium controls diet-induced obesity. *Proc. Natl Acad. Sci. USA* **110**, 9066–9071 (2013).
135. Guo, X. et al. High fat diet alters gut microbiota and the expression of Paneth cell-antimicrobial peptides preceding changes of circulating inflammatory cytokines. *Mediators Inflamm.* **2017**, 9474896 (2017).
136. Brun, P. et al. Increased intestinal permeability in obese mice: new evidence in the pathogenesis of nonalcoholic steatohepatitis. *Am. J. Physiol. Gastrointest. Liver Physiol.* **292**, G518–G525 (2007).
137. Regnier, M., Van Hul, M., Knauf, C. & Cani, P. D. Gut microbiome, endocrine control of gut barrier function and metabolic diseases. *J. Endocrinol.* **248**, R67–R82 (2021).
138. Bonomo, M. G., D'Angelo, S., Picerno, V., Carriero, A. & Salzano, G. Recent advances in gut microbiota in psoriatic arthritis. *Nutrients* **17**, 1323 (2025).
139. Barros, G., Duran, P., Vera, I. & Bermudez, V. Exploring the links between obesity and psoriasis: a comprehensive review. *Int. J. Mol. Sci.* **23**, 7499 (2022).
140. de Vos, W. M., Tilg, H., Van Hul, M. & Cani, P. D. Gut microbiome and health: mechanistic insights. *Gut* **71**, 1020–1032 (2022).
141. Buhas, M. C. et al. Gut microbiota in psoriasis. *Nutrients* **14**, 2970 (2022).
142. Hao, Y. et al. Gut microbiota dysbiosis and altered bile acid catabolism lead to metabolic disorder in psoriasis mice. *Front. Microbiol.* **13**, 853566 (2022).
143. Li, R., Andreu-Sanchez, S., Kuipers, F. & Fu, J. Gut microbiome and bile acids in obesity-related diseases. *Best. Pract. Res. Clin. Endocrinol. Metab.* **35**, 101493 (2021).
144. Paine, A. et al. Dysregulation of bile acids, lipids, and nucleotides in psoriatic arthritis revealed by unbiased profiling of serum metabolites. *Arthritis Rheumatol.* **75**, 53–63 (2023).
145. Torres, B. et al. Use of probiotics in preventing and treating excess weight and obesity. A systematic review. *Obes. Sci. Pract.* **10**, e759 (2024).
146. Hu, D., Zhao, J., Zhang, H., Wang, G. & Gu, Z. Fecal microbiota transplantation for weight and glycemic control of obesity as well as the associated metabolic diseases: meta-analysis and comprehensive assessment. *Life* **13**, 1488 (2023).
147. Kragtsnaes, M. S. et al. Safety and efficacy of faecal microbiota transplantation for active peripheral psoriatic arthritis: an exploratory randomised placebo-controlled trial. *Ann. Rheum. Dis.* **80**, 1158–1167 (2021).
148. Sanchez, P. et al. Efficacy of probiotics in rheumatoid arthritis and spondyloarthritis: a systematic review and meta-analysis of randomized controlled trials. *Nutrients* **14**, 354 (2022).
149. Garcia, M. M. et al. Adults with excess weight or obesity, but not with overweight, report greater pain intensities than individuals with normal weight: a systematic review and meta-analysis. *Front. Endocrinol.* **15**, 1340465 (2024).
150. Currado, D. et al. The relationship between metabolic syndrome and pain catastrophizing in psoriatic arthritis. *Rheumatol. Ther.* **12**, 581–592 (2025).
151. Binvignat, M., Sellam, J., Berenbaum, F. & Felson, D. T. The role of obesity and adipose tissue dysfunction in osteoarthritis pain. *Nat. Rev. Rheumatol.* **20**, 565–584 (2024).
152. Lubbeke, A. et al. Do synovial leptin levels correlate with pain in end stage arthritis? *Int. Orthop.* **37**, 2071–2079 (2013).
153. Zapata-Linares, N., Berenbaum, F. & Houard, X. Role of joint adipose tissues in osteoarthritis. *Ann. Endocrinol.* **85**, 214–219 (2024).
154. Ballegaard, C. et al. Knee pain and inflammation in the infrapatellar fat pad estimated by conventional and dynamic contrast-enhanced magnetic resonance imaging in obese patients with osteoarthritis: a cross-sectional study. *Osteoarthr. Cartil.* **22**, 933–940 (2014).
155. Karagiannides, I. & Pothoulakis, C. Substance P, obesity, and gut inflammation. *Curr. Opin. Endocrinol. Diabetes Obes.* **16**, 47–52 (2009).
156. Wang, L. et al. Levels of neuropeptide Y in synovial fluid relate to pain in patients with knee osteoarthritis. *BMC Musculoskelet. Disord.* **15**, 319 (2014).
157. Gowler, P. R. W. et al. Peripheral brain-derived neurotrophic factor contributes to chronic osteoarthritis joint pain. *Pain* **161**, 61–73 (2020).
158. Jafarzadeh, S. R. et al. Mediating role of bone marrow lesions, synovitis, pain sensitization, and depressive symptoms on knee pain improvement following substantial weight loss. *Arthritis Rheumatol.* **72**, 420–427 (2020).
159. Heuts, E. A. F., de Jong, L. D., Hazebroek, E. J., Wagener, M. & Somford, M. P. The influence of bariatric surgery on hip and knee joint pain: a systematic review. *Surg. Obes. Relat. Dis.* **17**, 1637–1653 (2021).
160. Gudbergsen, H. et al. Weight loss is effective for symptomatic relief in obese subjects with knee osteoarthritis independently of joint damage severity assessed by high-field MRI and radiography. *Osteoarthr. Cartil.* **20**, 495–502 (2012).
161. Patel, S. & Kumthekar, A. Psoriatic arthritis: the influence of co-morbidities on drug choice. *Rheumatol. Ther.* **9**, 49–71 (2022).
162. Xu, H., Wang, Z., Xu, L. & Su, Y. Refractory psoriatic arthritis: emerging concepts in whole process management. *Clin. Rheumatol.* **44**, 583–590 (2025).
163. Blake, T., Gullick, N. J., Hutchinson, C. E. & Barber, T. M. Psoriatic disease and body composition: a systematic review and narrative synthesis. *PLoS ONE* **15**, e0237598 (2020).
164. Jensen, M. D. et al. 2013 AHA/ACC/TOS guideline for the management of overweight and obesity in adults: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines and the Obesity Society. *Circulation* **129**, S102–S138 (2014).
165. Group, L. A. R. Eight-year weight losses with an intensive lifestyle intervention: the Look AHEAD study. *Obesity* **22**, 5–13 (2014).
166. O'Brien, P. E. et al. Long-Term outcomes after bariatric surgery: a systematic review and meta-analysis of weight loss at 10 or more years for all bariatric procedures and a single-centre review of 20-year outcomes after adjustable gastric banding. *Obes. Surg.* **29**, 3–14 (2019).
167. Arterburn, D. et al. Comparative effectiveness and safety of bariatric procedures for weight loss: a PCORnet cohort study. *Ann. Intern. Med.* **169**, 741–750 (2018).
168. Upala, S. & Sanguankeo, A. Effect of lifestyle weight loss intervention on disease severity in patients with psoriasis: a systematic review and meta-analysis. *Int. J. Obes.* **39**, 1197–1202 (2015).
169. Jensen, P. et al. Effect of weight loss on the severity of psoriasis: a randomized clinical study. *JAMA Dermatol.* **149**, 795–801 (2013).
170. Al-Mutairi, N. & Nour, T. The effect of weight reduction on treatment outcomes in obese patients with psoriasis on biologic therapy: a randomized controlled prospective trial. *Expert. Opin. Biol. Ther.* **14**, 749–756 (2014).
171. Gisondi, P., Del Giglio, M., Di Francesco, V., Zamboni, M. & Girolomoni, G. Weight loss improves the response of obese patients with moderate-to-severe chronic plaque psoriasis to low-dose cyclosporine therapy: a randomized, controlled, investigator-blinded clinical trial. *Am. J. Clin. Nutr.* **88**, 1242–1247 (2008).
172. Kimball, A. B., Alavian, C., Alora-Pallii, M. & Bagel, J. Weight loss in obese patients with psoriasis can be successfully achieved during a course of phototherapy. *J. Eur. Acad. Dermatol. Venereol.* **26**, 1582–1584 (2012).
173. Del Giglio, M., Gisondi, P., Tessari, G. & Girolomoni, G. Weight reduction alone may not be sufficient to maintain disease remission in obese patients with psoriasis: a randomized, investigator-blinded study. *Dermatology* **224**, 31–37 (2012).
174. Naldi, L. et al. Diet and physical exercise in psoriasis: a randomized controlled trial. *Br. J. Dermatol.* **170**, 634–642 (2014).
175. Abou-Raya, A., Abou-Raya, S. & Helmi, M. Effect of exercise and dietary weight loss on symptoms and systemic inflammation in obese adults with psoriatic arthritis: randomized controlled trial [abstract OPO076]. *Ann. Rheum. Dis.* **73**, 89–90 (2014).
176. Klingberg, E. et al. Weight loss improves disease activity in patients with psoriatic arthritis and obesity: an interventional study. *Arthritis Res. Ther.* **21**, 17 (2019).
177. Klingberg, E., Bjorkman, S., Eliasson, B., Larsson, I. & Bilberg, A. Weight loss is associated with sustained improvement of disease activity and cardiovascular risk factors in patients with psoriatic arthritis and obesity: a prospective intervention study with two years of follow-up. *Arthritis Res. Ther.* **22**, 254 (2020).
178. Eder, L. et al. Dietary interventions in psoriatic arthritis: a randomized controlled clinical trial [abstract]. *Arthritis Rheumatol.* **77**, 2690 (2025).
179. Ramonda, R. et al. Ketogenic diet improves disease activity and cardiovascular risk in psoriatic arthritis: a proof of concept study. *PLoS ONE* **20**, e0321140 (2025).
180. Lambadiari, V. et al. The effect of a ketogenic diet versus Mediterranean diet on clinical and biochemical markers of inflammation in patients with obesity and psoriatic arthritis: a randomized crossover trial. *Int. J. Mol. Sci.* **25**, 2475 (2024).
181. Adawi, M. et al. The impact of intermittent fasting (Ramadan fasting) on psoriatic arthritis disease activity, enthesitis, and dactylitis: a multicentre study. *Nutrients* **11**, 601 (2019).
182. Polachek, A., Touma, Z., Anderson, M. & Eder, L. Risk of cardiovascular morbidity in patients with psoriatic arthritis: a meta-analysis of observational studies. *Arthritis Care Res.* **69**, 67–74 (2017).
183. Jensen, P. et al. Effect of weight loss on the cardiovascular risk profile of obese patients with psoriasis. *Acta Derm. Venereol.* **94**, 691–694 (2014).
184. Higa-Sansone, G. et al. Psoriasis remission after laparoscopic Roux-en-Y gastric bypass for morbid obesity. *Obes. Surg.* **14**, 1132–1134 (2004).
185. de Menezes Ettinger, J. E. et al. Remission of psoriasis after open gastric bypass. *Obes. Surg.* **16**, 94–97 (2006).
186. Hossler, E. W., Maroon, M. S. & Mowad, C. M. Gastric bypass surgery improves psoriasis. *J. Am. Acad. Dermatol.* **65**, 198–200 (2011).
187. Romero-Talamás, H. et al. Psoriasis improvement after bariatric surgery. *Surg. Obes. Relat. Dis.* **10**, 1155–1159 (2014).
188. Sethi, M., Ren-Fieldin, C., Caminer, A., Scher, J. & Reddy, S. Clinical improvements in psoriasis and psoriatic arthritis with surgical weight loss [abstract]. *Arthritis Rheumatol.* **67**, 688 (2015).
189. Maglio, C., Peltonen, M., Rudin, A. & Carlsson, L. M. S. Bariatric surgery and the incidence of psoriasis and psoriatic arthritis in the Swedish Obese Subjects Study. *Obesity* **25**, 2068–2073 (2017).

190. Egeberg, A., Sørensen, J. A., Gislason, G. H., Knop, F. K. & Skov, L. Incidence and prognosis of psoriasis and psoriatic arthritis in patients undergoing bariatric surgery. *JAMA Surg.* **152**, 344–349 (2017).
191. Laferrère, B. Effect of gastric bypass surgery on the incretins. *Diabetes Metab.* **35**, 513–517 (2009).
192. Usinger, L. et al. Gastric emptying of orally administered glucose solutions and incretin hormone responses are unaffected by laparoscopic adjustable gastric banding. *Obes. Surg.* **21**, 625–632 (2011).
193. Ahern, T. et al. Glucagon-like peptide-1 analogue therapy for psoriasis patients with obesity and type 2 diabetes: a prospective cohort study. *J. Eur. Acad. Dermatol. Venereol.* **27**, 1440–1443 (2013).
194. Buyschaert, M., Tennstedt, D. & Preumont, V. Improvement of psoriasis during exenatide treatment in a patient with diabetes. *Diabetes Metab.* **38**, 86–88 (2012).
195. Reid, C. T., Tobin, A. M., Ahern, T., O’Shea, D. & Kirby, B. Liraglutide in combination with acitretin for severe recalcitrant psoriasis. *Br. J. Dermatol.* **169**, 230–231 (2013).
196. Xu, X. et al. Treatment with liraglutide, a glucagon-like peptide-1 analogue, improves effectively the skin lesions of psoriasis patients with type 2 diabetes: a prospective cohort study. *Diabetes Res. Clin. Pract.* **150**, 167–173 (2019).
197. Costanzo, G., Curatolo, S., Busà, B., Belfiore, A. & Gullo, D. Two birds one stone: semaglutide is highly effective against severe psoriasis in a type 2 diabetic patient. *Endocrinol. Diabetes Metab. Case Rep.* **2021**, 21-0007 (2021).
198. Lin, L. et al. Glucagon-like peptide-1 receptor agonist liraglutide therapy for psoriasis patients with type 2 diabetes: a randomized-controlled trial. *J. Dermatol. Treat.* **33**, 1428–1434 (2022).
199. Petković-Dabić, J. et al. Effects of semaglutide treatment on psoriatic lesions in obese patients with type 2 diabetes mellitus: an open-label, randomized clinical trial. *Biomolecules* **15**, 46 (2025).
200. Faurischou, A. et al. Lack of effect of the glucagon-like peptide-1 receptor agonist liraglutide on psoriasis in glucose-tolerant patients — a randomized placebo-controlled trial. *J. Eur. Acad. Dermatol. Venereol.* **29**, 555–559 (2015).
201. Eder, L. et al. Glucagon-like peptide-1 receptor agonists therapy is associated in improvement in psoriatic arthritis-related and metabolic outcomes: a retrospective analysis of two cohorts [abstract]. *Arthritis Rheumatol.* **77**, 2687 (2025).
202. Faurischou, A. et al. Improvement in psoriasis after treatment with the glucagon-like peptide-1 receptor agonist liraglutide. *Acta Diabetol.* **51**, 147–150 (2014).
203. Nicolau, J. et al. Effects of liraglutide among patients living with psoriasis and obesity. *Med. Clin.* **161**, 293–296 (2023).
204. Brennan, P. J., Brigl, M. & Brenner, M. B. Invariant natural killer T cells: an innate activation scheme linked to diverse effector functions. *Nat. Rev. Immunol.* **13**, 101–117 (2013).
205. Peternel, S. & Kastelan, M. Immunopathogenesis of psoriasis: focus on natural killer T cells. *J. Eur. Acad. Dermatol. Venereol.* **23**, 1123–1127 (2009).
206. Hogan, A. E. et al. Glucagon-like peptide-1 (GLP-1) and the regulation of human invariant natural killer T cells: lessons from obesity, diabetes and psoriasis. *Diabetologia* **54**, 2745–2754 (2011).
207. Wen, A. Y., Sakamoto, K. M. & Miller, L. S. The role of the transcription factor CREB in immune function. *J. Immunol.* **185**, 6413–6419 (2010).
208. Cai, Y. et al. Pivotal role of dermal IL-17-producing $\gamma\delta$ T cells in skin inflammation. *Immunity* **35**, 596–610 (2011).
209. Buyschaert, M. et al. Improvement of psoriasis during glucagon-like peptide-1 analogue therapy in type 2 diabetes is associated with decreasing dermal $\gamma\delta$ T-cell number: a prospective case-series study. *Br. J. Dermatol.* **171**, 155–161 (2014).
210. Marx, N. et al. Glucagon-like peptide-1(1-37) inhibits chemokine-induced migration of human CD4-positive lymphocytes. *Cell Mol. Life Sci.* **67**, 3549–3555 (2010).
211. Bliddal, H. et al. Once-weekly semaglutide in persons with obesity and knee osteoarthritis. *N. Engl. J. Med.* **391**, 1573–1583 (2024).
212. Mei, J., Sun, J., Wu, J. & Zheng, X. Liraglutide suppresses TNF- α -induced degradation of extracellular matrix in human chondrocytes: a therapeutic implication in osteoarthritis. *Am. J. Transl. Res.* **11**, 4800–4808 (2019).
213. Meurot, C. et al. Liraglutide, a glucagon-like peptide 1 receptor agonist, exerts analgesic, anti-inflammatory and anti-degradative actions in osteoarthritis. *Sci. Rep.* **12**, 1567 (2022).
214. Cheng, J., Solomon, T., Estee, M., Cicuttini, F. M. & Lim, Y. Z. Effect of glucagon-like peptide-1 receptor agonists in osteoarthritis: a systematic review of pre-clinical and human studies. *Osteoarthritis Cartil. Open* **7**, 100567 (2025).
215. Malhotra, K. et al. GLP-1 receptor agonists in diabetes for stroke prevention: a systematic review and meta-analysis. *J. Neurol.* **267**, 2117–2122 (2020).
216. Sinha, B. & Ghosal, S. Meta-analyses of the effects of DPP-4 inhibitors, SGLT2 inhibitors and GLP1 receptor analogues on cardiovascular death, myocardial infarction, stroke and hospitalization for heart failure. *Diabetes Res. Clin. Pract.* **150**, 8–16 (2019).
217. Olbrich, H. et al. GLP-1RA and reduced mortality, cardiovascular and psychiatric risks in psoriasis: a large-scale cohort study. *Br. J. Dermatol.* <https://doi.org/10.1093/bjd/ljaf346> (2025).
218. Tsbadze, N., Tskhakaia, I. & Tan, I. Mortality and major adverse cardiac events (MACE) with GLP-1 receptor agonists in psoriatic arthritis [abstract]. *Arthritis Rheumatol.* **77**, 0849 (2025).
219. Patel, J. P. et al. Comparative effectiveness of semaglutide, liraglutide, orlistat, and phentermine for weight loss in obese individuals: a systematic review. *Cureus* **17**, e80321 (2025).
220. Ryback, R. Topiramate in the treatment of psoriasis: a pilot study. *Br. J. Dermatol.* **147**, 130–133 (2002).
221. Singh, P. A., Cassel, K. P., Moscatti, R. M. & Eckersley, D. Acute generalized erythrodermic pustular psoriasis associated with bupropion/naltrexone (Contrave®). *J. Emerg. Med.* **52**, e111–e113 (2017).
222. Eakes, J., Moore, S., Hennessy, K. & Lin, A. Naltrexone-induced acute generalized exanthematous pustulosis. *JAAD Case Rep.* **39**, 115–117 (2023).
223. Lincoff, A. M. et al. Semaglutide and cardiovascular outcomes in obesity without diabetes. *N. Engl. J. Med.* **389**, 2221–2232 (2023).
224. Ardissino, M. et al. Long-term cardiovascular outcomes after orlistat therapy in patients with obesity: a nationwide, propensity-score matched cohort study. *Eur. Heart J. Cardiovasc. Pharmacother.* **8**, 179–186 (2022).
225. Nissen, S. E. et al. Effect of naltrexone-bupropion on major adverse cardiovascular events in overweight and obese patients with cardiovascular risk factors: a randomized clinical trial. *JAMA* **315**, 990–1004 (2016).
226. Sposito, A. C. et al. Cardiovascular safety of naltrexone and bupropion therapy: systematic review and meta-analyses. *Obes. Rev.* **22**, e13224 (2021).
227. Ritchey, M. E. et al. Cardiovascular safety during and after use of phentermine and topiramate. *J. Clin. Endocrinol. Metab.* **104**, 513–522 (2019).
228. Islam, K. N. et al. Obesity medications and their impact on cardiovascular health: a narrative review. *Cureus* **16**, e71875 (2024).
229. Drucker, D. J. Efficacy and safety of GLP-1 medicines for type 2 diabetes and obesity. *Diabetes Care* **47**, 1873–1888 (2024).
230. Skrobucha, A., Pindlowski, P., Krajewska, N., Grabowski, M. & Jonik, S. Anti-inflammatory effects of glucagon-like peptide-1 (GLP-1) in coronary artery disease: a comprehensive review. *Front. Cardiovasc. Med.* **11**, 1446468 (2024).
231. Halloum, W., Dughem, Y. A., Beier, D. & Pellesi, L. Glucagon-like peptide-1 (GLP-1) receptor agonists for headache and pain disorders: a systematic review. *J. Headache Pain.* **25**, 112 (2024).
232. Marzo-Ortega, H. et al. EULAR points to consider for the definitions of difficult-to-manage and treatment-refractory psoriatic arthritis [abstract POS1269]. *Ann. Rheum. Dis.* **84**, 1319–1320 (2025).
233. Alfaris, N. et al. GLP-1 single, dual, and triple receptor agonists for treating type 2 diabetes and obesity: a narrative review. *eClinicalMedicine* **75**, 102782 (2024).
234. Sherbini, A. A., Sharma, S. D., Gwinnutt, J. M., Hyrich, K. L. & Verstappen, S. M. M. Prevalence and predictors of adverse events with methotrexate mono- and combination-therapy for rheumatoid arthritis: a systematic review. *Rheumatology* **60**, 4001–4017 (2021).
235. Hewitson, P. J., Debroe, S., McBride, A. & Milne, R. Leflunomide and rheumatoid arthritis: a systematic review of effectiveness, safety and cost implications. *J. Clin. Pharm. Ther.* **25**, 295–302 (2000).
236. Kavanaugh, A. et al. Treatment of psoriatic arthritis in a phase 3 randomised, placebo-controlled trial with apremilast, an oral phosphodiesterase 4 inhibitor. *Ann. Rheum. Dis.* **73**, 1020–1026 (2014).
237. Burshtein, J. et al. The association between obesity and efficacy of psoriasis therapies: an expert consensus panel. *J. Am. Acad. Dermatol.* **92**, 807–815 (2025).
238. Sheth, S., Inestroza, K., Merola, J. F., Weber, B. & Garshick, M. Practical recommendations on cardiovascular risk evaluation in patients with psoriasis and psoriatic arthritis for dermatologists, rheumatologists, and primary care physicians by the Psoriasis and Psoriatic Arthritis Clinics Multicenter Advancement Network. *J. Psoriasis Psoriatic Arthritis* **28**, 24755303251337020 (2025).

Author contributions

The authors contributed equally to all aspects of the article.

Competing interests

R.H.H. has served as a consultant to Johnson & Johnson and Novartis. A.O. has received grant support from AbbVie, Johnson & Johnson, Novartis and Pfizer, and to Forward database from Amgen and Bristol Myers Squibb; and has served as a consultant fees from AbbVie, Amgen, Bristol Myers Squibb, Celgene, CorEvitas, Lilly, Gilead, Happify, Janssen, Novartis, Pfizer and UCB. J.F.M. has received personal fees from AbbVie, Amgen, AstraZeneca, Boehringer Ingelheim, Bristol Myers Squibb, Dermavant, Janssen, Lilly, Moonlake, UCB and Novartis; and has received grants from Pfizer and Sun Pharma. J.U.S. has served as a consultant for AbbVie, Johnson & Johnson, Kaleido, Novartis, Pfizer, Sanofi and UCB; and has received funding for investigator-initiated studies from Johnson & Johnson, Novartis, and Pfizer. L.E. has received grants from AbbVie, Pfizer, Johnson & Johnson, UCB, Novartis and Lilly.

Additional information

Peer review information *Nature Reviews Rheumatology* thanks Maria Sole Chimenti, Rubén Queiro and Eric Toussiot for their contribution to the peer review of this work.

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

© Springer Nature Limited 2025


Endosome traffic in rheumatic diseases: mechanistic insights and therapeutic opportunities

Andras Perl  

Abstract

Endosomal traffic governs various core processes that maintain immune homeostasis and self-tolerance, including receptor signalling, antigen processing, cytokine secretion and cellular metabolism. Traffic-regulated receptors – both intracellular and on the cell surface – modulate immune sensing of infection, nutrient availability and endogenous stress signals arising from cellular or tissue injury. In rheumatic diseases, such as systemic lupus erythematosus, rheumatoid arthritis, ankylosing spondylitis, systemic sclerosis, Sjögren syndrome and osteoarthritis, mounting evidence implicates disruptions in endosomal pathways as important drivers of disease onset and progression. Dysregulated endosomal trafficking contributes to type I interferon activation via signalling through Toll-like receptors, aberrant autoantigen presentation, and altered expression of metabolite transporters in immune cells and target organs. Endosome trafficking mediates autophagosome formation, the production of exosomes and the turnover of organelles, such as mitochondria that generate oxidative stress, thereby controlling chronic inflammation and connective tissue remodelling. Therefore, understanding the molecular architecture of endosomal recycling pathways and their integration with immune cell function can provide important insight into rheumatic diseases. Restoring trafficking fidelity – through modulation of RAB GTPases, endosomal Toll-like receptor signalling, metabolic reprogramming, autophagic flux and extracellular vesicle biology – represents a promising therapeutic strategy.

Departments of Medicine, Microbiology and Immunology, and Biochemistry and Molecular Biology, State University of New York Upstate Medical University, Norton College of Medicine, Syracuse, NY, USA.

 e-mail: perla@upstate.edu

Sections

[Introduction](#)[Endosomal function](#)[Genetic and epigenetic drivers of endosomal dysfunction](#)[Endosomal pathways in immune processes](#)[Endosomal trafficking in rheumatic diseases](#)[Therapeutic implications and opportunities](#)[Conclusions](#)

Key points

- Endosomes are sorting stations that receive proteins from the Golgi or plasma membrane via endocytosis and recycle them to the cell surface, trans-Golgi network, or lysosomes for degradation.
- Disrupted endosomal traffic contributes to rheumatic diseases, including systemic lupus erythematosus, rheumatoid arthritis and systemic sclerosis, by impairing immune regulation and tissue homeostasis.
- Endosomes regulate immune homeostasis through receptor traffic and signal transduction, antigen processing, cytokine secretion, and cellular metabolism.
- Endosomal pathways initiate autophagosomes and exosome formation and remove damaged organelles, including oxidative stress-generating mitochondria that promote inflammation and connective tissue remodelling.
- Dysregulated endosomal traffic activates type I interferon via Toll-like receptors, disrupts antigen presentation and alters metabolite transporter expression in immune cells.
- Key checkpoints of endosomal traffic, including RAB GTPases, endosomal Toll-like receptors, recycled receptors, metabolite cargos, autophagy flux and exosome formation, represent promising therapeutic targets in endosomal dysfunction.

Introduction

The immune system depends on tightly coordinated membrane trafficking to regulate surface receptor expression, signal transduction, and degradation or recycling of internalized cargo¹. These trafficking decisions, primarily orchestrated through endosomes, influence critical immune processes such as antigen processing², T cell and B cell activation³, cytokine release⁴, and resolution of inflammation⁵. Defects in endosomal trafficking are increasingly recognized not merely as downstream consequences of inflammation but as causal contributors in the pathogenesis of autoimmune and inflammatory disorders.

Endosomal trafficking senses and regulates virtually every aspect of cellular function⁶. In rheumatic diseases, where sustained activation of innate and adaptive immune responses drives chronic tissue injury, endosome dysfunction can amplify immunogenicity and perturb immune tolerance³. Endosomal traffic has a central role in regulating the recycling and expression of nutrient-sensing surface receptors, the activity of nucleic acid-sensing intracellular receptors, and the biogenesis and degradation of organelles through lysosomal pathways. Endosome traffic is guided by RAB GTPases along a network of microtubules sustained by the actin cytoskeleton⁷. In humans, over 60 RAB GTPases function as master regulators of intracellular membrane traffic^{8,9}. Distinct RAB GTPases control different phases of endosome traffic to ensure that membrane-bound cargoes are transported to their correct destinations within the cell¹⁰.

In this Review, we examine recent advances in the biogenesis and trafficking of endosomes, with an emphasis on Toll-like receptor (TLR)-dependent interferon signalling in the innate immune system, the recycling of metabolite-sensing receptors in adaptive immune

responses and the regulation of end-organ resistance. These insights reveal broader implications for the pathogenesis and treatment of rheumatic diseases, including in the development and progression of degenerative joint disorders.

Endosomal function

Endosomes are dynamic cellular compartments that function as sorting stations for proteins and lipids, orchestrating membrane-enclosed traffic and intracellular communication. Endocytic recycling pathways regulate the trafficking of receptors and organelles, shaping cell membrane composition and cellular responsiveness¹¹. Endosomes receive cargo from both the extracellular environment and biosynthetic pathways, directing this cargo to various destinations such as the plasma membrane, Golgi or lysosomes for degradation. Essentially, endosomes are responsible for regulating the flow of cellular components, influencing cell signalling and maintaining cellular homeostasis. Defects in endosome function can lead to various diseases¹², including metabolic disorders, neurodegenerative diseases, cancer and rheumatic diseases, the latter of which is the focus of this Review.

Endosomal biogenesis and maturation

Like all cell membranes within the endomembrane system, endosome membranes are composed of a phospholipid bilayer⁷. This bilayer features hydrophilic head groups that are oriented outwards, facing the aqueous environment, and hydrophobic tails that point inwards, forming the core of the membrane. Endosomal biogenesis is a highly coordinated process essential for controlling membrane protein turnover, receptor signalling and antigen presentation. Endosomes are formed by endocytosis, where the cell membrane invaginates and pinches off to form a vesicle. During biogenesis, the endosome membrane is structurally similar to other cellular membranes; however, endosomes also contain unique proteins and lipids that define their specific functions and maturation stages.

Endosome maturation begins with the formation of early endosomes at the cell periphery, following the internalization of cargo via endocytosis¹³. These early endosomes serve as dynamic sorting hubs, directing internalized material towards recycling or degradation pathways (Fig. 1). Endocytosis proceeds through two main pathways: clathrin-mediated endocytosis and clathrin-independent endocytosis^{14,15}. Clathrin-mediated endocytosis involves the assembly of clathrin and adaptor proteins, such as AP2, at the plasma membrane, forming a clathrin-coated pit. This pit invaginates and pinches off to form a clathrin-coated vesicle¹⁶. Clathrin-mediated endocytosis accounts for 95% of endocytic vesicles¹⁷.

Early endosomes undergo maturation processes that involve morphological changes, spatial movement and acidification. The mature late endosome fuses with a lysosome, forming a transient organelle known as an endolysosome, which subsequently becomes a classical dense lysosome¹³. RAB GTPases, functioning as molecular switches, orchestrate each stage of this process¹⁸ (Fig. 1). RAB5, enriched on early endosomes, regulates internalization and homotypic fusion as well as cargo sorting via effectors such as early endosomal antigen 1 (EEA1) and the class III phosphoinositide 3-kinase (PI3K) complex¹⁹. Cargo destined for rapid recycling to the plasma membrane engages RAB4A, which mediates fast recycling via direct sorting tubules. This pathway is critical for rapid restoration of immune receptors such as CD98 (also known as SLC3A2) and CD71 during lymphocyte activation³. These receptors are internalized following phosphorylation by protein kinase C (PKC). By contrast, RAB11 regulates slower recycling from perinuclear recycling endosomes²⁰, maintaining surface expression of immune receptors, such as CD4

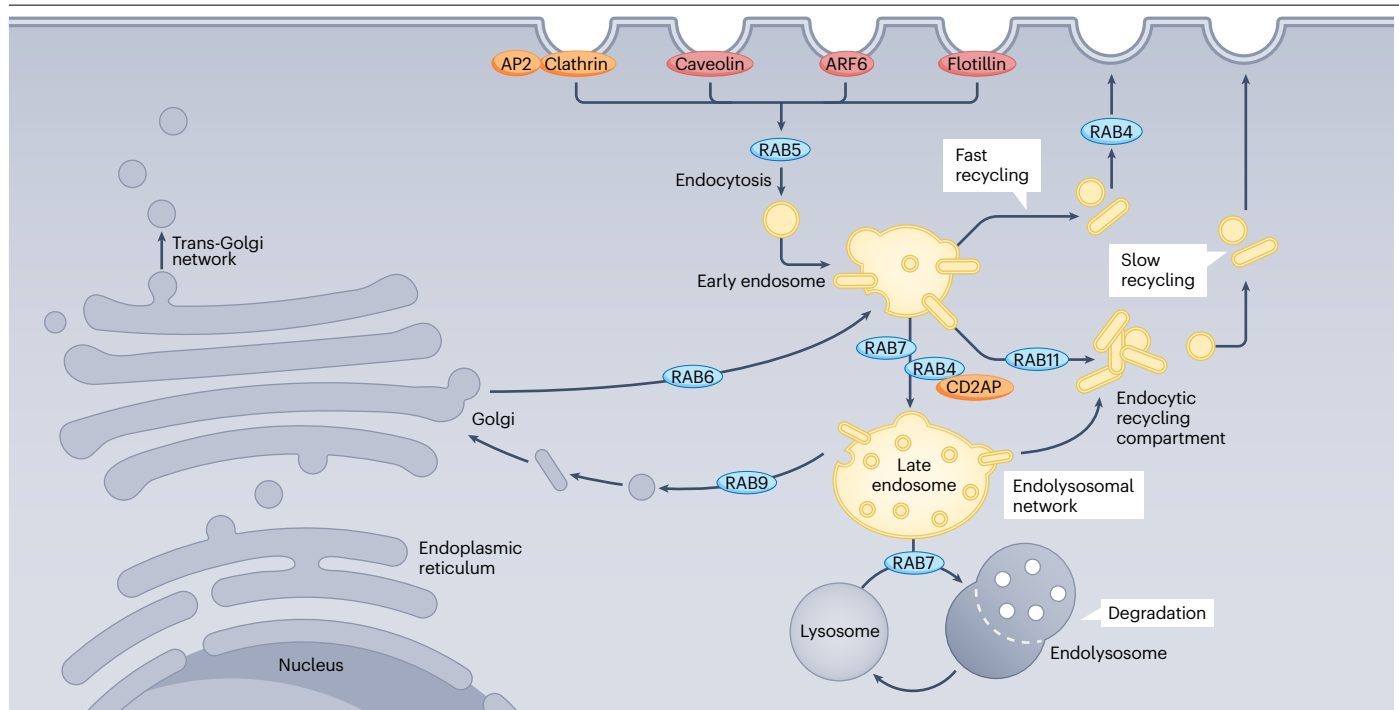


Fig. 1 | Endosomal pathways: biogenesis, recycling, exosome formation and lysosomal delivery. Endosome biogenesis relies on clathrin²⁷⁴, caveolin²²⁷, ARF6 (ref. 275) or flotillin¹⁴⁹ for vesicle formation. Clathrin is associated with AP2. RAB5A is a key regulator of the formation and internalization of early endosomes, which are the initial sorting stations for internalized cargo. RAB4 collaborates with RAB11 in recycling processes: RAB4 controls the fast-recycling pathway, directly returning cargo from early endosomes to the plasma membrane or to the lysosome; whereas RAB11 is located in perinuclear recycling endosomes and regulates the slow-recycling pathway, which involves the transport of cargo from

the recycling endosomes to the cell surface. RAB7 has a role in the maturation of early endosomes to late endosomes (facilitating the RAB5 to RAB7 switch) and in the subsequent fusion of late endosomes with lysosomes, essential for lysosomal degradation. RAB9 isoforms mediate the traffic from late endosomes to the trans-Golgi network²⁴, whereas RAB6 oppositely regulates transport between the Golgi and endosomes²⁵. CD2-associated protein (CD2AP) interacts with RAB4 and helps to regulate the sorting and recycling of cargo between early and late endosomes²⁸. CD2AP also promotes the transition from RAB5⁺ early endosomes to RAB7⁺ late endosomes, targeting cargo for lysosomal degradation²⁹.

(ref. 21), and integrins during sustained responses²². Maturation towards degradation involves RAB conversion, whereby RAB5 is replaced by RAB7 (ref. 23) through the action of the MON1-CCZ1 exchange complex, promoting cargo transport to late endosomes and lysosomes¹¹.

RAB9A and RAB9B are primarily involved in the traffic from late endosomes to the trans-Golgi network²⁴ (Fig. 1). RAB6 regulates bidirectional transport between the Golgi and endosomes²⁵. RAB7-mediated trafficking controls antigen processing in lysosome-related compartments for major histocompatibility complex (MHC) antigen loading. The spatiotemporal regulation of RAB activation by guanine nucleotide exchange factors and GTPase-activating proteins ensures precise coordination of endosomal dynamics²⁶, linking receptor fate decisions to innate and adaptive immune function²⁷. CD2-associated protein (CD2AP) interacts with RAB4A to regulate cargo sorting and recycling between early and late endosomes²⁸. CD2AP also promotes the transition from RAB5⁺ early endosomes to RAB7⁺ late endosomes, targeting cargo for lysosomal degradation²⁹. Dysregulation of these pathways has been implicated in aberrant immune activation and impaired tolerance, contributing to the pathogenesis of autoimmune rheumatic disease^{30,31}.

Endosomal trafficking pathways

After internalization, endosomes travel through an interconnected tubular network that controls the transfer of cargoes between organelles.

As already discussed, early endosomes (marked by RAB5) initiate sorting, directing cargo either for recycling via RAB4⁺ or RAB11⁺ compartments or for degradation through late endosomes and lysosomes¹. Endosomes can also be returned to the cell surface and become exosomes^{32,33}. Thus, upon fusion with the plasma membrane, endosomes can become exosomes that allow sharing of intracellular materials between cells in close proximity and throughout the body³⁴.

The acidic environment of the early endosome forces ligands to be released from the receptors such as transferrin and iron from CD71 (Fig. 2a). Most ligands are sorted into recycling endosomes and directed to lysosomes for degradation. The receptors themselves can also be degraded within lysosomes or recycled back to the cell surface for reuse¹⁰. As an example of endosomal trafficking, the transferrin receptor CD71 is internalized via clathrin-mediated endocytosis³⁵, a process regulated by RAB5, which also facilitates the uptake of its cargo, transferrin and iron¹⁵. Once internalized, CD71 is recycled to the plasma membrane via RAB4A^{36–38} or RAB11 (ref. 39). In turn, RAB7⁺ vesicles transport newly synthesized CD71 from the Golgi to the plasma membrane⁴⁰. The intracellular protein dynamin-related protein 1 (DRP1) also traffics via clathrin-coated vesicles⁴¹. DRP1 is critical for the initiation of mitochondrial fission, a process required for mitochondrial turnover through mitochondrial autophagy, known as mitophagy⁴². When DRP1 is depleted via lysosomal trafficking,

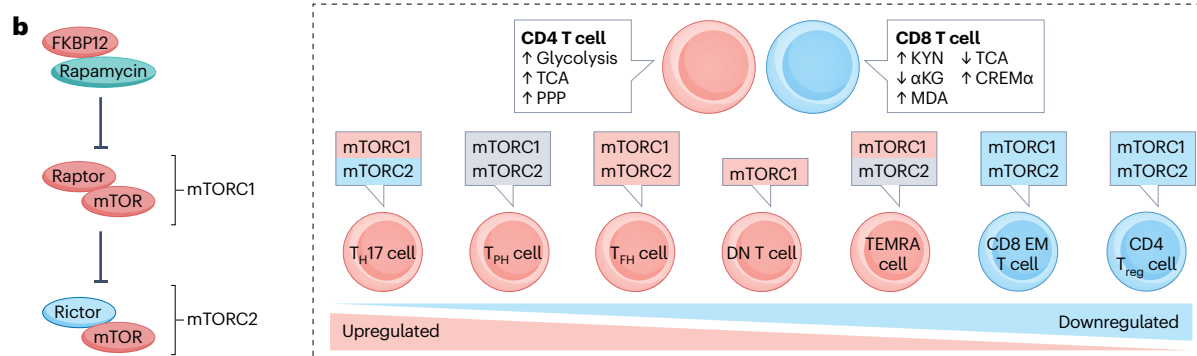
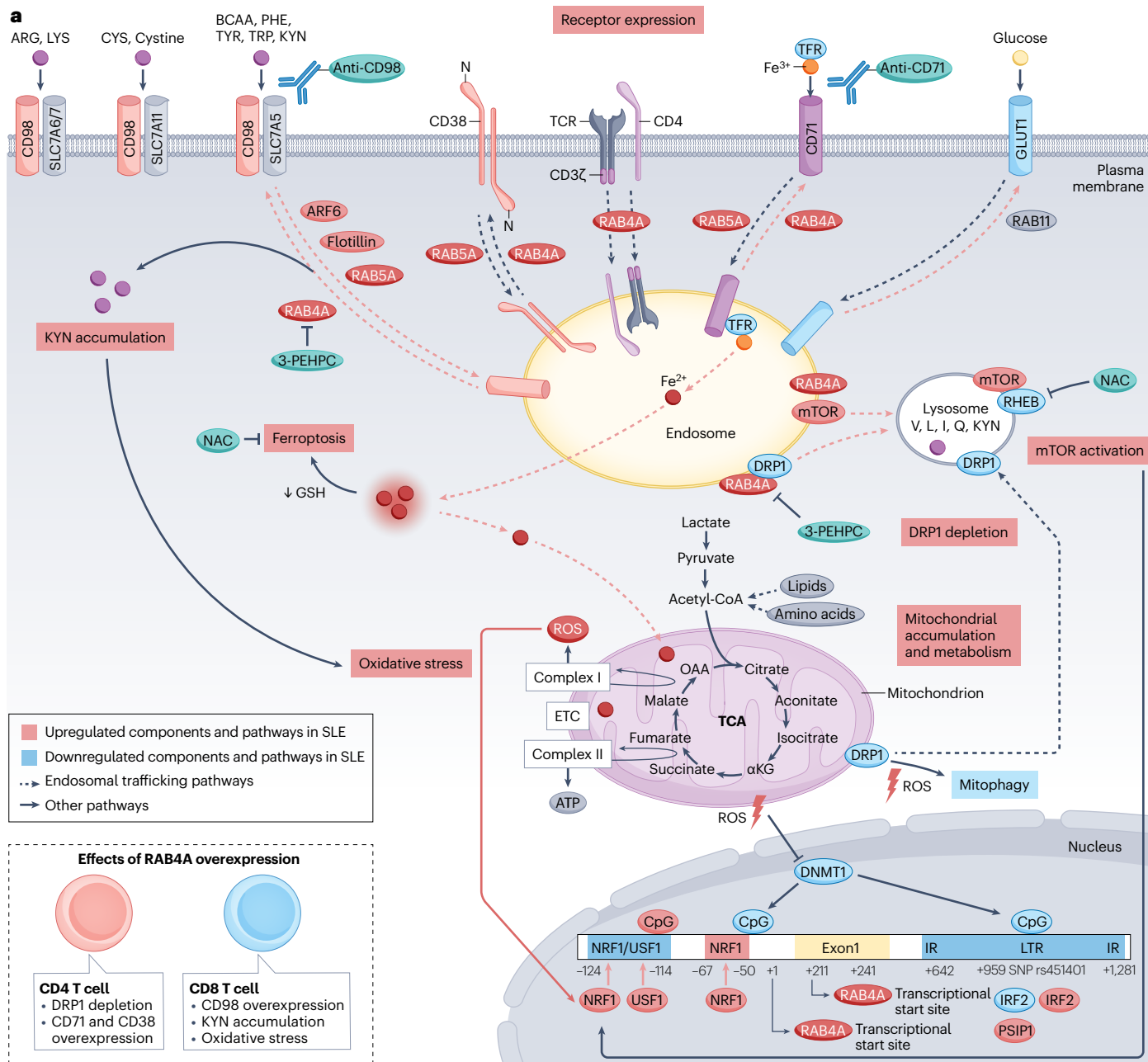


Fig. 2 | Complexity of endosome traffic-mediated T cell lineage skewing in SLE. Endosomal trafficking has an important role in the pathogenesis of systemic lupus erythematosus (SLE). Multiple mechanistically interconnected checkpoints that regulate endosomal trafficking and mechanistic target of rapamycin complex (mTOR) signalling, including CREM α , Raptor, Rictor, RAB4A, RAB5A and CD98, are dysregulated in SLE. These components can lead to the upregulation and downregulation of various processes (depicted in red and blue boxes, respectively), in a T cell-specific fashion, leading to skewed T cell lineage commitment. **a**, RAB4A engages in cell-type-specific positive feedback loops with mTOR signalling, driving metabolic and immunological reprogramming. In CD4⁺ T cells, RAB4A promotes dynamin-related protein 1 (DRP1) depletion, upregulates CD71 and CD38, and enhances mitochondrial activity and pentose phosphate pathway (PPP) activity. In CD8⁺ T cells, increased CD98 recycling leads to kynurenine (KYN) accumulation, oxidative stress and cell loss. RAB4A also facilitates lysosomal degradation of CD4 and CD3 ζ and impaired T cell receptor (TCR) signalling. Increased iron uptake via CD71, combined with glutathione (GSH) depletion, elicits ferroptosis²⁷⁶. RAB4A expression is regulated via its 5' promoter by redox-sensitive transcription factors NRF1 and USF1 and its polymorphic LTR-enhancer by interferon-regulated IRF2 (ref. 25), linking oxidative stress to RAB4A induction. Reactive oxygen species (ROS) further inhibit DNA methyltransferase 1 (DNMT1), causing demethylation of the RAB4A promoter and its overexpression. Additionally, mTOR amplifies this axis by

upregulating NRF1 and RAB4A expression. **b**, RAB4A overexpression enhances mTORC1 activity while suppressing mTORC2 activity in human primary CD4⁺ T cells¹⁶⁸. Consistent with this pattern, T cells from patients with SLE exhibit increased mTORC1 and reduced mTORC2 signalling²⁷⁷; however, the degree of dysregulation is lineage specific across SLE T cell subsets^{181,278,279}. Stimulatory (red), inhibitory (blue) or undefined (grey) roles of mTORC1 and mTORC2 are shown for T cell development (bottom right box). For both panels, molecules shown in red are upregulated in SLE, and those shown in blue are downregulated in SLE. Red arrows indicate molecular pathways that are accelerated in SLE, and grey arrows indicate molecular pathways that have no defined directional abnormality in SLE; dashed arrows indicate endosome trafficking, and solid arrows represent other pathways. For RAB molecules, those shown in red are upregulated in SLE, and those shown in grey are involved in the trafficking of indicated proteins without evidence of altered expression in SLE. In both figure parts, pharmacological interventions with 3-PHPC, N-acetylcysteine (NAC) and rapamycin as well as with CD71 and CD98 blocking antibodies are shown in green. α KG, α -ketoglutarate; BCAA, branched-chain amino acids; DN, double negative; EM, effector memory; ETC, electron transport chain; GLUT1, glucose transporter 1; MDA, malondialdehyde; OAA, oxaloacetate; TCA, tricarboxylic acid; TEMRA, terminally differentiated effector memory T cells re-expressing CD45RA; TFH, T follicular helper; TFR, transferrin; T_H17, T helper 17; T_{PH}, T peripheral helper; T_{reg}, regulatory T.

mitophagy is disrupted, resulting in the accumulation of mitochondria. These mitochondria can contribute membrane components for accelerated autophagosome formation^{43,44}.

Clathrin-independent endocytosis encompasses various pathways that do not rely on clathrin for vesicle formation⁴⁵. These pathways include caveolin-mediated endocytosis, which involves the caveolin proteins 1, 2 and 3, and the formation of caveolae (flask-shaped invaginations of the plasma membrane)⁴⁶; ARF6-dependent endocytosis, which relies on the small GTPase ARF6 and its associated proteins; and flotillin-dependent endocytosis, which utilizes flotillin proteins and lipid rafts⁴⁵ (Fig. 1). As an example, the amino acid transporter CD98 might be internalized via clathrin-independent endocytosis⁴⁵ involving flotillin⁴⁷, ARF6 (ref. 48), or RAB5A⁴⁷ and recycled back to the plasma membrane via RAB4A-regulated endosomes³ (Fig. 2a).

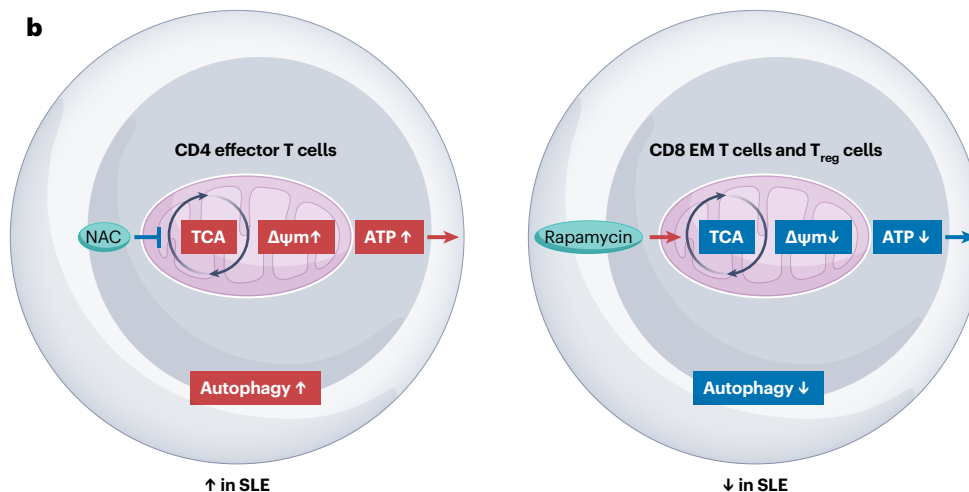
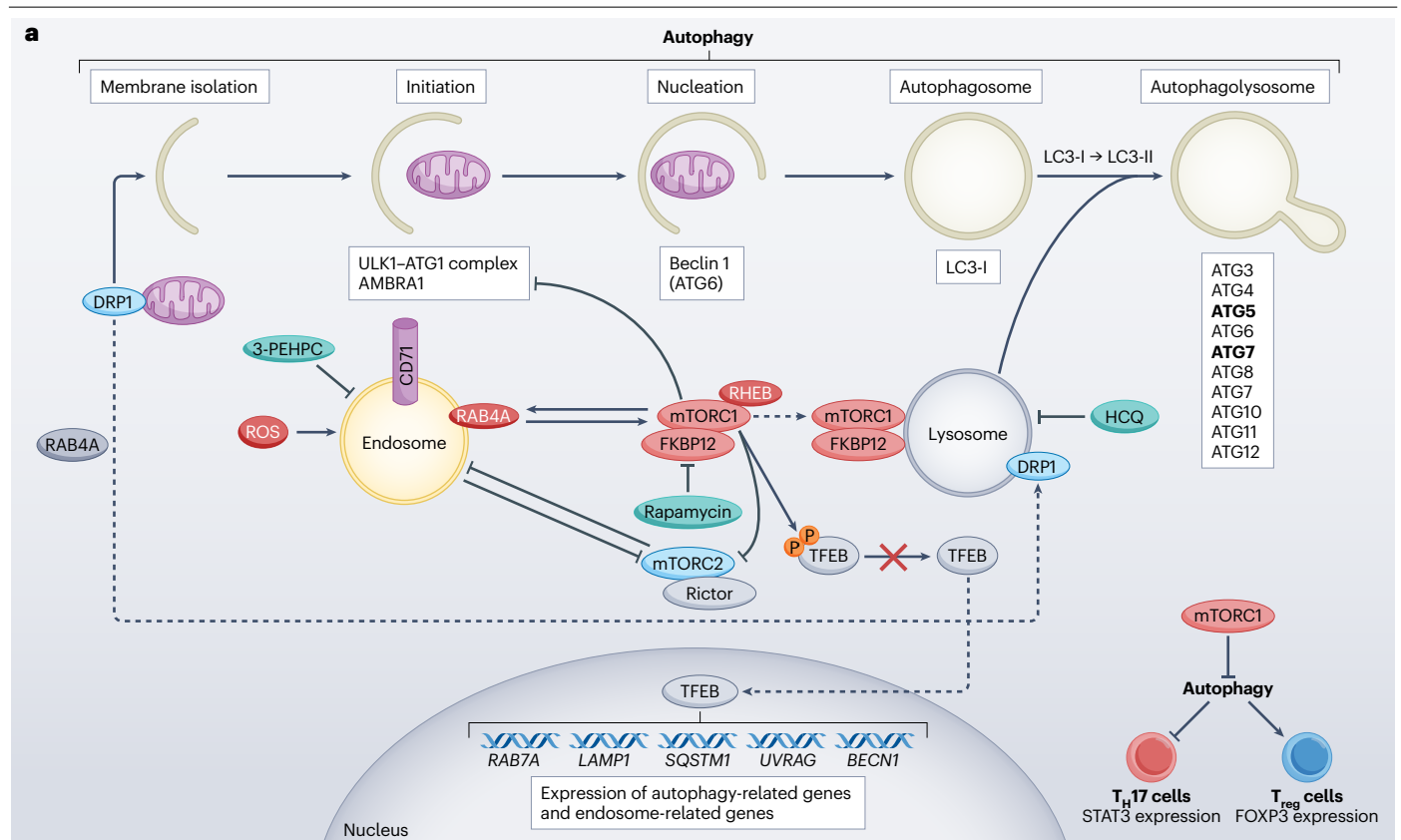
Exosome formation

Extracellular vesicles constitute a broad array of cell-derived, lipid-bound particles⁴⁹. Exosomes represent a subset of extracellular vesicles that originate from endosomes and are released when multivesicular bodies fuse with the plasma membrane, whereas other extracellular vesicles, such as microvesicles, are formed by direct budding from the plasma membrane. Exosomes are generally smaller in size (30–150 nm) than microvesicles (100–1,000 nm) and are characterized by specific markers, including tetraspanins such as lysosome-associated membrane protein 3 (LAMP3; also known as CD63)⁴⁹. The accumulation of endosomes stimulates the formation and secretion of exosomes⁵⁰, facilitating the intercellular spread of inflammatory signals and autoantigens^{51,52}. Among the RAB GTPases, RAB4A has emerged as a prominent regulator of exosome biogenesis^{32,33,53} and colocalizes with LAMP3 (refs. 54,55). The LAMP3 interactome also includes several other endosomal GTPases such as RAB5A, RAB11A, RAB7A, RAB27A, RAB27B and ARL8B⁵⁶. Notably, exosome production is enhanced^{57,58}, along with the expression of RAB4A, in both patients and mice with systemic lupus erythematosus (SLE)-like disease^{31,59,60}. Extracellular vesicles are also implicated in other rheumatic diseases, including rheumatoid arthritis (discussed in a later section).

Endosomal control of autophagy

Endocytosis and autophagy are both cellular processes that deliver materials to lysosomes for degradation, yet these processes differ fundamentally in their origin, cargo and mechanism. Endocytosis mediates the internalization of extracellular substances and the redistribution of membrane-bound components. By contrast, autophagy is a catabolic process that begins within the cell with the formation of an autophagosome, and involves the removal and recycling of damaged or unnecessary intracellular components, macromolecules and organelles. The two pathways exhibit extensive 'crosstalk' and can influence each other⁶¹. Transcription factor EB (TFEB) is a central regulator of endosomal and lysosomal biogenesis, essential for the recycling of organelles and intracellular debris during autophagy⁶² (Fig. 3a). For example, under nutrient-rich conditions, mechanistic target of rapamycin complex 1 (mTORC1) phosphorylates TFEB on serine residues 142 and 211, retaining TFEB in the cytoplasm in an inactive state^{63,64}. Upon dephosphorylation, when nutrient levels are low, TFEB translocates to the nucleus and regulates the expression of autophagy-related genes within the CLEAR (coordinated lysosomal expression and regulation) network, including genes encoding ATG5 and ATG8 (also known as LC3), and of RAB7 (ref. 65). In addition to controlling autophagy-related genes, TFEB regulates genes involved in endosomal trafficking and maturation, such as genes encoding RAB5A, RAB7A and caveolin 2, and an overall large subset of 623 endocytosis-related genes, thereby linking autophagy with endocytic pathways⁶². mTORC1 directly inhibits autophagy by blocking the function of the activating molecule in Beclin1-regulated autophagy (AMBRA1, Fig. 3).

RAB4A has a pivotal role in autophagosome formation⁴⁴ and endosomal trafficking^{31,60}. RAB4A directs cargo, such as CD3 ζ ⁶⁰ and DRP1 (ref. 31), to lysosomes for degradation, and facilitates the recycling of CD71 and CD98 to the plasma membrane in a cell-type-specific manner³. Functional studies have shown that increased RAB4A expression contributes to these trafficking outcomes, as small interfering RNA-mediated knockdown in primary human T cells⁶⁰ or CD4Cre-driven deletion in mouse T cells reverses these effects³. RAB4A-mediated downregulation of the autophagy regulator DRP1



has been implicated as a pathogenic mechanism in SLE, as discussed in a later section.

Genetic and epigenetic drivers of endosomal dysfunction

Various genetic and epigenetic factors are linked to the disruption of endosomal pathways, contributing to the pathogenesis of rheumatic diseases. For example, the *UNC93B1* protein facilitates the trafficking of nucleotide-sensing TLRs from the endoplasmic reticulum to endolysosomes⁶⁶, where they initiate immune responses. Genome-wide association studies in patients with SLE have identified risk variants in

UNC93B1 (refs. 67–69), highlighting the role of intracellular trafficking in autoimmune pathogenesis. Epigenetic regulation also intersects with trafficking pathways: DNA methylation-dependent associations involving DNA methyltransferase 1 (DNMT1)⁷⁰ (Fig. 2a) and DNMT3 have been reported between RAB4A and SLE²¹, and between RAB4B and chronic obstructive pulmonary disease⁷¹, implicating RAB GTPases in immune dysfunction. Notably, the retroviral features of the *HRES1/RAB4* locus³⁸ and the high level of sequence homology between RAB4A and RAB4B (Supplementary Fig. 1) suggest a gene duplication event via retroposition⁷². Regulatory non-coding RNAs further modulate trafficking pathways: microRNA-155 controls exosome secretion⁷³,

Fig. 3 | Endosome–autophagy interactions shape T cell metabolic reprogramming in SLE. Multiple regulatory nodes integrate endosomal trafficking, autophagy and mitochondrial metabolism to shape T cell dysfunction in systemic lupus erythematosus (SLE). **a**, Autophagy occurs through a series of steps under the regulation of various molecules (shown in white boxes). Autophagosome membrane formation might be initiated by sourcing membranes from mitochondria²⁸⁰, a process that is accelerated by RAB4A⁴⁴. Endosome biogenesis is regulated by transcription factor EB (TFEB)-induced gene expression and post-translational modifications mediated by mechanistic target of rapamycin complex 1 (mTORC1). TFEB also regulates the expression of various autophagy-related genes (including Beclin 1, encoded by *BECN1*). Phosphorylation of serine residues on TFEB by mTORC1 retains TFEB in the cytosol in an inactive form⁶³. mTORC1 also promotes the activity of RAB4A and inhibits the activity of activating molecule in Beclin1-regulated autophagy (AMBRA1), a key regulator of autophagy. RAB4A targets dynamin-related

protein 1 (DRP1) for lysosomal degradation and supports the accumulation of oxidative stress-generating mitochondria, particularly in T helper 17 (T_H17) cells. Under physiological conditions, autophagy supports FOXP3 expression and regulatory T (T_{reg}) cell development and restrains STAT3 expression and T_H17 cell development. In SLE, various components of these pathways are upregulated (shown in red) or downregulated (shown in blue). Notably, in this setting, crosstalk between RAB4A⁺CD71⁺ recycling endosomes and lysosomes activates mTOR, establishing a positive feedback loop that sustains mTOR activation and RAB4A overexpression. **b**, In SLE, mitochondrial metabolic flux, ATP production and autophagy are enhanced in CD4⁺ T cells but reduced in CD8⁺ effector memory (EM) and T_{reg} cells²⁷⁸. mTOR activation blocks autophagy in T_{reg} cells (not shown). For both panels, pharmacological interventions with 3-PEHPC, N-acetylcysteine (NAC), hydroxychloroquine (HCQ) and rapamycin are shown in green. $\Delta\Psi_m$, mitochondrial transmembrane potential; ROS, reactive oxygen species; TCA, tricarboxylic acid.

whereas microRNA-574 is transferred intracellularly between cells via exosomes and activates TLR7, promoting type I interferon production by plasmacytoid dendritic cells (pDCs)⁵⁷. Together, these findings point to a convergence of genetic susceptibility and epigenetic mechanisms in endosome-regulated and exosome-regulated immune signalling.

Nucleotide-binding domain (NOD)-like receptors (NLRs) are intracellular proteins that detect pathogens and activate the inflammasome. The subcellular localization and interaction of these receptors with endosomal compartments vary by receptor subtype. Although NOD1 and NOD2 sense cytosolic bacterial products, NLRP3 is also recruited to endosomes in response to the build-up of lipid phosphatidylinositol 4-phosphate (PI4P), a process driven by endocytic disruption^{74,75}. Notably, genetic polymorphisms of *NLRP3* have been linked to rheumatoid arthritis and Sjögren syndrome⁷⁶.

C-type lectin receptors (CLRs) are endocytic receptors that internalize bound ligands, including both microbial components and host-derived molecules, through pathways such as clathrin-mediated endocytosis⁷⁷. Following internalization, these CLRs and their ligands are delivered to early and late endosomes, where their fate – either recycling to the cell surface or degradation in lysosomes or phagolysosomes – depends on the specific CLR and its ligand. CLRs are crucial for innate immunity and antigen presentation. One CLR of particular interest in this context is C-type lectin-like domain family 16 member A (CLEC16A). Variants in the gene encoding CLEC16A have been linked to a wide spectrum of autoimmune disorders, including juvenile idiopathic arthritis⁷⁸, anti-cyclic citrullinated peptide-negative rheumatoid arthritis⁷⁸, type 1 diabetes⁷⁹, Crohn disease⁸⁰ and multiple sclerosis⁸¹. Although the precise functional relevance of this link is not yet fully understood, CLEC16A is expressed in multiple immune cell types, where it regulates both autophagic processes and receptor expression. Notably, the autoimmune risk genotype is associated with higher expression of CLEC16A in CD4⁺ T cells⁸². In B cell lines in vitro, CLEC16A expression and its disease-associated locus, rs17673553, promote starvation-induced autophagy by inhibiting mTOR⁸³. However, in primary human lymphocytes, increased CLEC16A expression instead activates mTOR and confers resistance to autophagy, suggesting a cell lineage-specific effect⁸⁴.

Functionally, CLEC16A encodes an endosomal membrane protein that supports mitophagy⁸⁵ and localizes to RAB4A⁺ recycling endosomes in CD4⁺ T cells⁸¹. In CLEC16A-deficient mice, mitophagy is disrupted in both T cells and B cells, but this defect can be reversed by pharmacological inhibition of PI3K and mitogen-activated protein

kinase kinase (MEK)⁸⁶. These findings further implicate CLEC16A as a key mediator linking endosome trafficking to the pathogenesis of autoimmune diseases.

The cGAS–STING pathway also engages endosomal compartments at multiple stages of its activation and resolution⁷¹. Both cGAS and STING translocate to endosomes as part of their activation pathway, and the presence of STING in endosomal membranes is essential for interferon induction (Supplementary Fig. 2). Binding of cGAS to the endosomal membrane activates the enzyme to synthesize cGAMP, which in turn helps STING translocate from the endoplasmic reticulum to endosomes. Subsequently, RAB7-driven endosomal traffic moves STING to lysosomes for degradation, which counters excessive inflammation^{71,87}. Activating genetic mutations in STING predispose to STING-associated vasculopathy with onset in infancy⁸⁸ and SLE⁸⁹.

Endosomal pathways in immune processes

Endosomal pathways have a central role in shaping immune responses by regulating receptor signalling, antigen processing, cellular metabolism and lineage specification. Their roles are particularly evident in TLR signalling and in the pathogenesis of autoimmune diseases such as SLE and rheumatoid arthritis.

Endosomal pathways in TLR signalling

Endosomal TLRs, including TLR3, TLR7, TLR8, TLR9 and TLR13, recognize pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs), initiating innate immune responses⁹⁰. Although TLR13 is restricted to mice⁹¹, the remaining TLRs are critical sensors in human immunity. PAMPs and DAMPs trigger immune system activation through binding to the leucine-rich repeat segment of TLRs⁹². PAMPs, such as bacterial CpG DNA and single-stranded RNA, are derived from pathogens such as bacteria and viruses⁹³, whereas DAMPs, such as double-stranded RNA, are released from damaged or dying host cells⁹⁴. Endosome-trafficked TLR3 senses double-stranded RNA and induces the production of pro-inflammatory cytokines, including type I interferons⁹⁵, which are implicated in the pathogenesis of several rheumatic diseases^{96,97} (Fig. 4).

Systemic lupus erythematosus. In SLE, altered endosomal trafficking has a central role in dysregulated nucleic acid sensing by B cells and the innate immune system. The transmembrane protein UNC93B1 regulates the localization of TLR7 to endosomes⁹⁸ (Fig. 4). TLR7 and TLR9, normally restricted to endolysosomal compartments, become

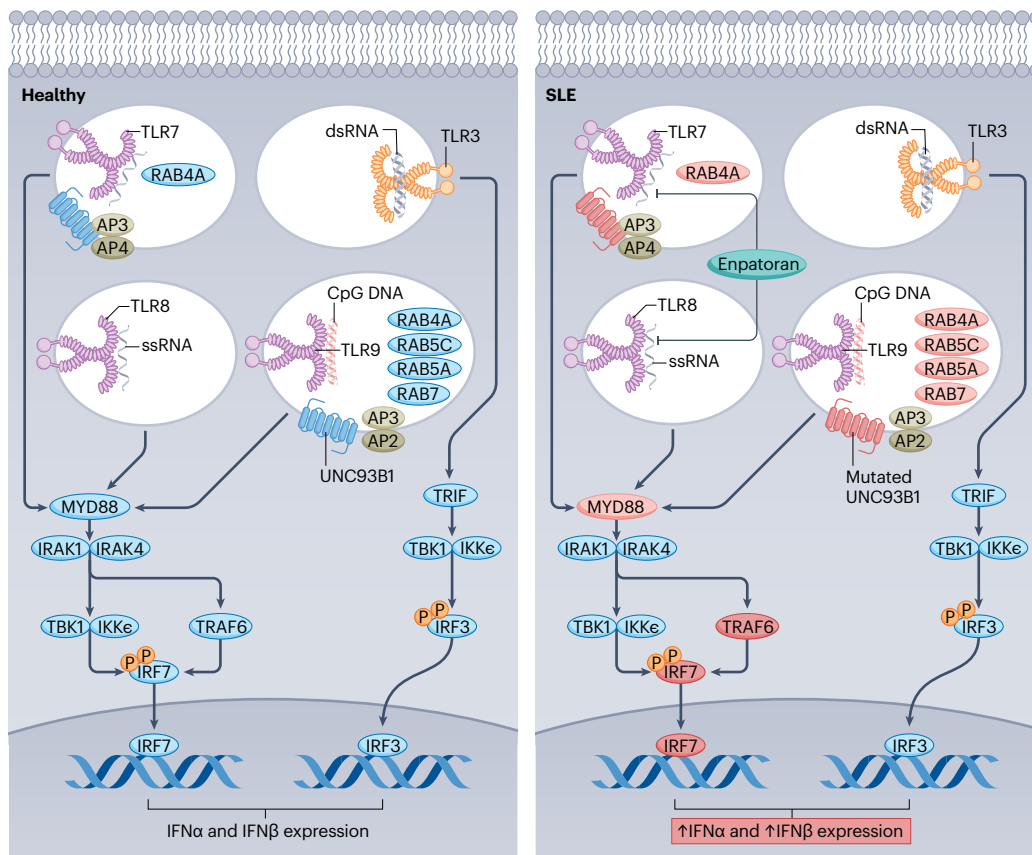


Fig. 4 | Regulation of TLR-mediated interferon production via endosome traffic. Toll-like receptor (TLR)-mediated interferon production is altered in systemic lupus erythematosus (SLE) (right). In healthy conditions (left), single-stranded RNA (ssRNA), double-stranded RNA (dsRNA) and CpG DNA from viruses and bacteria activate TLR3, TLR7, TLR8 and TLR9, initiating MYD88-dependent and TRIF-dependent signalling. TLR7 and TLR9 require endosomal trafficking through RAB5A⁺, RAB4A⁺ and RAB7⁺ endosomes¹⁰⁸. RAB4A directly binds CD2-associated protein (CD2AP)^{60,204} and interacts with AP1 (ref. 26), AP2 (ref. 281) and AP3 adaptor proteins²⁶ (not shown). Deletion of CD2AP abrogates dendritic cell

migration²⁸². Both TLR7 and TLR9 signalling depend on AP3 (ref. 283). UNC93B1 (shown in blue, left) regulates TLR7 and TLR9 endosomal trafficking, using adaptor proteins AP2 and AP4 for TLR9 and TLR7 transport, respectively⁶⁶. In SLE (right), mutated forms of UNC93B1 (shown in red, right) skew trafficking towards TLR7, amplifying MYD88-mediated and TRAF6-mediated expression of IFNα and IFNβ¹⁰³. Enpatoran, a dual TLR7 and TLR8 inhibitor (shown in green), has shown preliminary evidence for safety and efficacy in SLE¹⁰⁴. AP, accessory protein; dsDNA, double-stranded DNA.

hyperactivated in SLE owing to overexpression, defective trafficking (for example, via UNC93B1), and/or persistent engagement with self-RNA or DNA^{99,100}. UNC93B1 controls homeostatic TLR7 activation by balancing TLR9 to TLR7 trafficking; pro-inflammatory mutations, such as D34A, skew this balance, promoting TLR7-mediated B cell-dependent CD4 T cell activation in mouse models¹⁰¹.

UNC93B1 mutations can cause monogenic SLE or chilblain lupus owing to differentially enhanced TLR7 and TLR8 signalling over TLR9 signalling^{67,68}. Similarly, missense mutations in UNC93B1 have been associated with either autosomal dominant or autosomal recessive forms of SLE in multiple families⁶⁷. G325C and I317M amino acid substitutions in UNC93B1 enhance TLR7 signalling and type I interferon production in patients with SLE and chilblain lupus⁶⁷. These mutations activate TLR7 signalling both in monocytes and pDCs and elicit excessive type I interferon (IFNα and IFNβ) production, hallmarks of SLE^{67,68,102}. Mechanistically, SLE-associated UNC93B1 mutations promote MyD88-mediated and TRAF6-mediated expression of IFNα and IFNβ via preferential TLR7 signalling¹⁰³. Enpatoran, a dual TLR7

and TLR8 inhibitor, has shown preliminary evidence of safety and efficacy in SLE¹⁰⁴.

Viral infections are well-established triggers of disease flares in patients with SLE and other rheumatic diseases^{105,106}. Viral proteins can directly interact with Rab GTPases¹⁰⁷. Additionally, viral RNA can activate TLR7 and TLR9, a process that is tightly regulated by endosomal trafficking involving RAB4A⁺ and RAB7A⁺ endosomes¹⁰⁸ (Fig. 4).

Endosomal TLR signalling is also critical for xenobiotic-induced autoimmunity, as demonstrated by the requirement of TLR trafficking from the endoplasmic reticulum to endolysosomal and phagosomal compartments for HgCl₂-induced hypergammaglobulinaemia and autoantibody production, independent of type I interferon signalling¹⁰⁰. In SLE, patients carrying E92G and R336L variants in UNC93B1 exhibit hypergammaglobulinaemia and expansion of memory-switched B cells, consistent with enhanced B cell hyperresponsiveness driven by TLR7 hyperactivity¹⁰². These findings indicate that type I interferon production by pDCs and autoantibody generation by B cells could be independently triggered through TLR signalling in SLE.

Rheumatoid arthritis. In rheumatoid arthritis, synovial fibroblasts exhibit increased endocytic recycling involving RAB44 (ref. 109) and TLRs that enhance invasiveness and cytokine responsiveness¹¹⁰. These processes involve the overexpression of endosome-bound TLRs (such as TLR7, TLR8 and TLR9) on rheumatoid arthritis synoviocytes¹¹¹. TLR7 expression is also upregulated on CD8 T cells in rheumatoid arthritis¹¹². Endogenous ligands, such as U11snRNA¹¹³, miR-let-7b¹¹⁴ and extracellular miR-574-5p, activate TLR7 and TLR8 signalling, driving osteoclast differentiation and contributing to joint destruction^{115,116}. Neutrophil extracellular traps, which release nucleic acids into the extracellular space, serve as additional sources of TLR ligands in rheumatoid arthritis^{111,117}. Neutrophil extracellular traps are internalized by macrophages in rheumatoid arthritis via RAB5A-trafficked endosomes, triggering the production of pro-inflammatory cytokines such as IL-6 and TNF¹¹⁸. Similarly, TLR4 activation by lipopolysaccharides (LPS) in macrophages depends on RAB5A-mediated endocytosis in rheumatoid arthritis¹¹⁹. Given the involvement of endosome traffic in rheumatoid arthritis pathogenesis, pro-inflammatory RAB5A alone or in combination with TLRs might serve as novel targets for therapeutic interventions.

Antigen processing

Endosomes serve as primary compartments for the internalization and processing of exogenous antigens, such as bacteria or viruses, by antigen-presenting cells (APCs) through processes such as phagocytosis and endocytosis^{120,121}. Endosomes guide phagocytosed cargo for presentation via MHC class I and class II pathways¹²². The type of receptor used for internalization can determine the initial routing and subsequent processing of the antigen. Specific sorting signals on the cytoplasmic tails of receptors guide them, along with their internalized cargo, into various endosomal pathways¹²³. For instance, some receptors target antigens to non-degradative endosomes for cross-presentation, whereas others lead to rapid degradation in lysosomes¹²⁴. In particular, RAB4-directed^{125,126} and endophilin A2-directed endosome traffic promotes antigen presentation by B cells¹²⁷.

HLA-B27, a MHC class I molecule, is strongly associated with susceptibility to spondyloarthropathies, particularly ankylosing spondylitis¹²⁸. Within APCs, endosome-derived vesicular traffic facilitates HLA-B27-mediated antigen presentation at the immunological synapse, promoting activation of CD8 T cells in ankylosing spondylitis¹²⁹. Notably, β 2-microglobulin-free HLA-B27 heavy chains, including disulfide-linked homodimers (B27₂), can be expressed at the cell surface in ankylosing spondylitis following endosomal recycling of conventional β 2-microglobulin-associated HLA-B27 heterotrimers. These free heavy-chain forms, including B27₂, engage innate immune receptors on T cells, natural killer cells and myeloid cells, thereby promoting pro-inflammatory signalling¹³⁰. Although direct mechanistic evidence is lacking, genes involved in endosomal trafficking, such as *RAB5C*, *SYNJ1* and *RNF19B*, are overexpressed in synovial tissue-infiltrating T cells and monocytes from patients with ankylosing spondylitis¹³¹. *SYNJ1* encodes synaptojanin 1, a phosphoinositide phosphatase critical for synaptic vesicle recycling and membrane trafficking¹³², whereas *RNF19B* encodes a ubiquitin ligase involved in autophagy¹³³. Thus, endosome trafficking pathways might contribute to enhanced MHC class I-dependent antigen presentation in ankylosing spondylitis¹³⁴.

Immune cell metabolism

Endosome traffic influences immune cell metabolism by regulating the cell-surface expression of receptors that mediate nutrient uptake.

These receptors include glucose transporters (glucose transporter 1 (GLUT1) and GLUT4)¹³⁵, CD71 (the transferrin receptor that mediates iron uptake)¹³⁶ and the CD98–SLC7A5 heterodimer, which mediates the transport of branched-chain amino acids (including leucine, isoleucine and valine) and aromatic amino acids (including tryptophan, kynurenine, tyrosine and phenylalanine)³ (Fig. 2a). Beyond SLC7A5, CD98 also forms heterodimers with SLC7A6 or SLC7A7, which transports arginine and lysine, and SLC7A11, which transports cysteine and cystine.

Endosomal pathways are also involved in mitochondrial metabolic pathways. RAB4A controls endosomal trafficking of DRP1 to the lysosome, and RAB4A-driven DRP1 depletion promotes the accumulation of oxidative stress-generating mitochondria³¹. In parallel, nutrient uptake supports distinct metabolic programmes across immune cell subsets. Glucose fuels glycolytic flux¹³⁷, which predominates in CD4 effector T cells¹³⁷, whereas iron supports mitochondrial biogenesis¹³⁶. By contrast, CD8 T cells and regulatory T (T_{reg}) cells rely on mitochondrial respiration for survival¹³⁸. Overexpression of phosphoenolpyruvate carboxykinase 1 (PCK1) can enhance glycolytic activity and the effector functions of tumour-infiltrating CD4 and CD8 T cells¹³⁹. Additionally, effector memory CD8 T cells might shift towards glycolysis upon viral infection¹⁴⁰.

During enhanced metabolic activity, the integrity of a cell depends on the maintenance of a reducing environment, the availability of reduced glutathione and the production of NADPH by the pentose phosphate pathway¹⁴¹. Notably, glutathione depletion activates mTORC1 in T cells from patients with SLE, an effect that can be reversed by treating patients with its amino acid precursor N-acetylcysteine¹⁴². Diminished expression of SLC7A11 and the depletion of glutathione predispose renal tubular epithelial cells to ferroptosis in lupus nephritis¹⁴³ (Fig. 2a). mTOR has an important role in sensing amino acid sufficiency¹⁴⁴, a process that depends on dynamin-mediated endocytosis, which delivers amino acids to the lysosome and facilitates the recruitment and activation of mTORC1 and mTORC2 onto the lysosomal surface^{3,145}. Distortions of these endosomal pathways lead to metabolic dysregulation and contribute to autoimmune rheumatic disorders, such as SLE, as reviewed elsewhere¹⁴⁶.

T cell lineage specification

Endosomal trafficking regulates the surface expression of receptors that are critical for signal transduction in adaptive immune cells, including both T cells^{3,38,60} and B cells^{30,125,147}. In T cells, recycling endosomes deliver T cell receptors (TCRs) to the plasma membrane, supporting immunological synapse formation and sustained signalling¹⁴⁸. TCR internalization occurs via distinct endocytic pathways depending on ligand engagement: ligand-bound TCRs are internalized together with trogocytosed MHC from APCs through a clathrin-independent process¹⁴⁹, whereas non-engaged TCRs are internalized by RAB5 via clathrin-dependent endocytosis and recycled to the plasma membrane through RAB4⁺ recycling endosomes^{150,151}.

TCR signalling also depends on the formation of supramolecular activation clusters at the T cell side of the immunological synapse¹⁵². RAB4A regulates the recycling of CD71 (ref. 153), which is also recruited to the supramolecular activation cluster in CD4 T cells¹⁵⁴. RAB4A promotes surface expression of CD71 in CD4 but not in CD8 T cells, whereas RAB4A selectively promotes the recycling and expression of CD98 in CD8 T cells³. Thus, RAB4A-mediated endosome traffic initiates distinct, cell-type-specific metabolic reprogramming via these nutrient-sensing transporters in CD4 and CD8 T cells³ (Fig. 2b). In T_{reg} cells, CTLA4 surface expression and recycling are also regulated by endosomal trafficking pathways. The expression of FOXP3 and the immunosuppressive

function of T_{reg} cells critically depend on CTLA4, the deficiency of which results in fatal autoimmunity¹⁵⁵. Endosomal trafficking of CTLA4 is controlled by the LPS-responsive beige-like anchor protein (LRBA) and RAB11 (ref. 156). RAB4B has also been associated with T cell dysfunction and is implicated in disrupting the balance between T helper 17 cells and T_{reg} cells, a dysregulation linked to the development of insulin resistance¹⁵⁷.

Finally, endosomal trafficking facilitates mTOR activation, glycolysis and mitochondrial metabolism via GLUT1-mediated glucose transport¹³⁷ and CD71-mediated iron uptake, as discussed in the previous section. These metabolic processes support the development of effector memory T cells re-expressing CD45RA and contribute to the metabolic regulation of T helper 17 cell expansion relative to T_{reg} cells¹³⁶.

Endosomal trafficking in rheumatic diseases

Disruptions in endosome trafficking can lead to both immune dysregulation and end-organ vulnerability in rheumatic diseases (such as the compromised resilience of renal podocytes observed in SLE^{158,159}), increasing susceptibility to inflammatory and tissue damage. Within the immune system, endosome traffic affects the surface expression of antigen receptors, such as the TCR, signal transduction via the TCR and other receptors, immune synapse formation, phagocytosis, and cytokine and antibody production. Beyond immune cells, aberrant endosomal trafficking affects cellular metabolism, autophagy and apoptotic signalling, contributing to end-organ injury. For example, pathological changes can occur in chondrocytes¹⁶⁰, fibroblasts¹⁶¹ and renal podocytes¹⁶², which are central to the progression of joint, connective tissue and kidney involvement in rheumatic diseases.

Systemic lupus erythematosus

RAB4A has emerged as a key molecular link between genetic susceptibility and immune dysregulation in SLE. Polymorphic alleles of *RAB4A* have been associated with susceptibility to SLE^{163,164} and multiple sclerosis^{165–167}. Overexpression and hyperactivation of RAB4A disrupt T cell development and contribute to SLE pathogenesis^{3,31}. RAB4A expression is transcriptionally regulated via its 5' promoter by redox-sensitive transcription factors NRF1 and USF1, and through its polymorphic LTR-enhancer by interferon-regulated IRF2 (ref. 168) (Fig. 2a). Thus, oxidative stress and activation of the interferon pathway contribute to RAB4A overexpression¹⁶⁸.

The gene encoding RAB4A originates from the *HRES1/RAB4* human endogenous retroviral element³⁸, which might function as a sensor of viral infection and potential mediator of autoimmunity^{169,170}. Polymorphic alleles of *HRES1/RAB4* have been linked to autoantibody production and susceptibility to autoimmune diseases, including SLE^{163,164} and multiple sclerosis^{165,166}. Specific *HRES1* haplotypes influence autoantibody profiles and organ involvement, such as glomerulonephritis, in patients with SLE¹⁶³. *HRES1* polymorphisms impact the expression RAB4A. Importantly, RAB4A is overexpressed in patients with SLE carrying rs451401CC alleles in the retroviral long terminal repeat of *HRES1*, which elicits mTORC1 activation²⁵.

In SLE, RAB4A-mediated downregulation of the autophagy regulator DRP1 impairs mitochondrial clearance, leading to the accumulation of damaged mitochondria and elevated oxidative stress^{31,171}. Autophagy is generally diminished in SLE owing to cell-type-specific mTOR activation and underlying genetic defects^{172,173}. Diminished autophagic flux leads to the accumulation of immunogenic debris and enhancement of TLR-dependent inflammation¹⁷⁴. Lysosomal acidification is also impaired in lupus-prone models, altering endosomal maturation and

antigen degradation^{175,176}. Although RAB4A activates mTOR¹⁶⁸, the underlying molecular mechanisms are incompletely understood³.

RAB4A directly facilitates mTOR trafficking to lysosomes, where mTOR senses amino acid sufficiency, including branched-chain amino acids, tryptophan and kynurenine, which are accumulated in lupus-prone mice^{177,178} and patients with SLE¹⁷⁹. The two mTOR protein complexes (mTORC1 and mTORC2) have crucial roles in lineage development both within the innate and adaptive immune systems^{180,181}. Notably, mTOR enhances the expression of redox-sensitive transcription factor NRF1 (ref. 182) that binds to the *RAB4A* promoter¹⁶⁸. Thus, RAB4A and mTOR form a positive feedback loop (Fig. 2a). mTOR itself localizes to endosomes¹⁸³ alongside small GTPases such as RAB4A⁶⁰, RAB7 (ref. 183) and RAB5 (ref. 184). Both RAB4A and RAB5 are overexpressed in T cells from patients with SLE⁶⁰ and lupus-prone mice³¹. However, only RAB4A overexpression, and not RAB5 overexpression, precedes mTOR activation, anti-nuclear antibody (ANA) production and the onset of nephritis in lupus-prone mice³¹.

RAB4A promotes pro-inflammatory T cell lineage specification via mTOR activation at multiple levels, as demonstrated in lupus-prone B6 SLE1.2.3 triple-congenic (B6.TC) mice. In this model, constitutively active RAB4^{AQ72L} drives mTOR activation and reveals cell-type-specific roles for RAB4A in promoting inflammatory T cell lineage development³. In CD4 T cells, RAB4A enhances iron uptake via CD71, supporting mitochondrial metabolism by increasing mitochondrial accumulation, facilitating recycling and expression of CD71 (which provides iron for electron transport¹³⁶), and increasing mitochondrial ATP production³. This metabolic reprogramming supports elevated flux through the tricarboxylic acid cycle and pentose phosphate pathway, thereby sustaining cell proliferation. In CD8 T cells, RAB4A accelerates the recycling and surface expression of CD98 (Fig. 2a), which serves as a receptor for branched-chain amino acids, phenylalanine, tyrosine and tryptophan metabolites such as kynurenine. Kynurenine accumulates in serum and CD8 T cells via CD98 uptake in SLE^{19,31}, contributing to oxidative stress and CD8 T cell depletion. This imbalance amplifies inflammation by favouring CD4 T cell expansion over CD8 T cells. Activation of RAB4A-mediated mTOR pathways augments ANA and antiphospholipid autoantibody production and promotes glomerulonephritis in mice³. By contrast, targeted inactivation of RAB4A in T cells, or pharmacological blockade of mTOR, abrogates mTOR hyperactivation, prevents lineage skewing and attenuates autoimmune pathology³. Notably, elevated CD98 expression by T cells predicts the therapeutic response to mTOR blockade in patients with SLE^{3,185}.

RAB4A also promotes the endocytic recycling and surface expression of CD38 (ref. 186), a transmembrane glycoprotein expressed by multiple immune cell types that has an important role in NAD⁺ metabolism. Elevated expression of CD38 in SLE consequently drives NAD⁺ depletion, activates mTORC1 and suppresses IL-2 production in CD4 T cells. Mechanistically, CD38 activity depletes NAD⁺ and leads to the build-up of nicotinamide and ADP-ribose and a secondary reduction in cyclic ADP-ribose levels. CD38 is internalized by RAB5A¹⁸⁷ and is recycled by RAB4A¹⁸⁶. Notably, RAB4A-dependent upregulation of CD38 and associated IL-2 suppression occur independently of mTOR signalling. Instead, enhanced activity of CD38 elevates STAT3 expression and acetylation as well as FOXO1 expression, collectively contributing to IL-2 repression in CD4 T cells¹⁸⁶. Together, these findings identify a previously unrecognized RAB4A–CD38 signalling axis that links receptor trafficking to pro-inflammatory metabolic reprogramming, highlighting potential therapeutic targets in SLE¹⁸⁸ (Fig. 5).

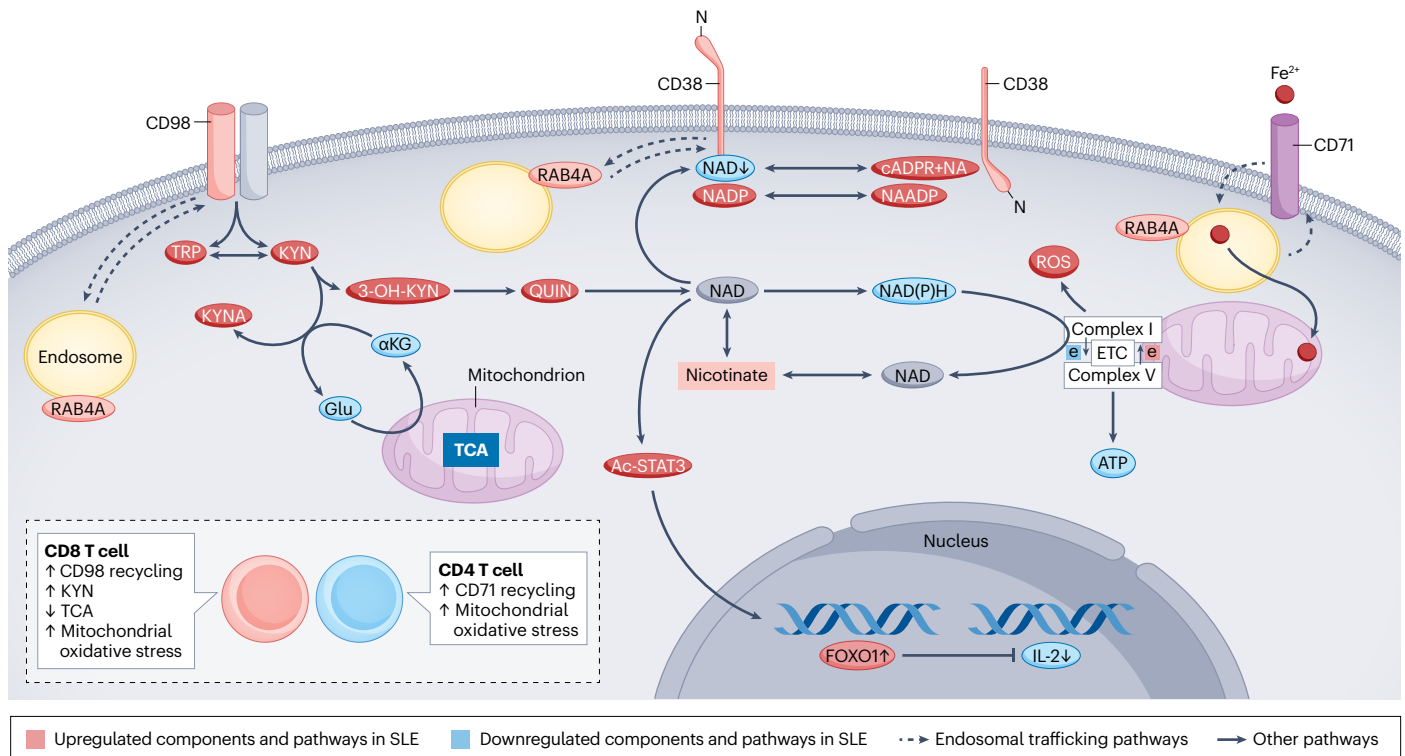


Fig. 5 | RAB4A-recycled receptors synergistically promote mitochondrial oxidative stress in T cells in SLE. RAB4A promotes the recycling of CD71 and CD98 in CD4⁺ and CD8⁺ T cells, respectively³, as well as CD38. CD98 facilitates the transport of kynurenine (KYN), a precursor for de novo NAD biosynthesis, yet KYN metabolism depletes α -ketoglutarate (α KG), a key tricarboxylic acid (TCA) cycle intermediate. Notably, immune cells can make NAD from KYN²⁸⁴. Concurrently, CD38 – a bidirectional ectoenzyme – reduces NAD levels both intracellularly and extracellularly²⁸⁵, impairing electron transport chain (ETC) activity and triggering reactive oxygen species (ROS) generation at complex I²⁸⁶.

Increased activity of CD38 also promotes STAT3 expression and acetylation, and subsequent FOXO1 expression, and suppresses IL-2 expression in CD4 T cells. In parallel, CD71-mediated iron uptake exacerbates mitochondrial oxidative stress and promotes ferroptosis²⁸⁷. Together, these processes converge to amplify metabolic stress in systemic lupus erythematosus (SLE) T cells, with molecules shown in blue downregulated and those in red upregulated in SLE. 3-OH-KYN, 3-hydroxykynurenine; Ac-STAT3, acetylated STAT3; cADPR, cyclic ADP-ribose; NA, nicotinic acid; NAADP, nicotinic acid adenine dinucleotide phosphate; QUIN, quinolinic acid.

Although RAB4A interacts with CD4, CD71 and CD2AP, it does not interact with the CD8 antigen⁶⁰. Intracellular CD8 expression has been observed in monocytes¹⁸⁹; however, the potential association of CD8 with endosomal compartments, its recycling dynamics or targeting for lysosomal degradation remain undefined. In terms of other RAB GTPases³¹, palmitoylated CD8 associates with Ick in lipid rafts¹⁹⁰, a configuration that might facilitate interaction with RAB11⁺ endosomes¹⁹¹. Within CD8 T cells, RAB27A regulates the release of cytotoxic granules¹⁹² and is notably overexpressed in lymphocytes from patients with SLE¹⁹³. Finally, in the B cell compartment, RAB7 controls immunoglobulin class switching and plasma cell survival via CD40 trafficking and NF- κ B signalling in murine models of SLE, directly linking endosomal dynamics to humoral autoimmunity³⁰.

Lupus nephritis

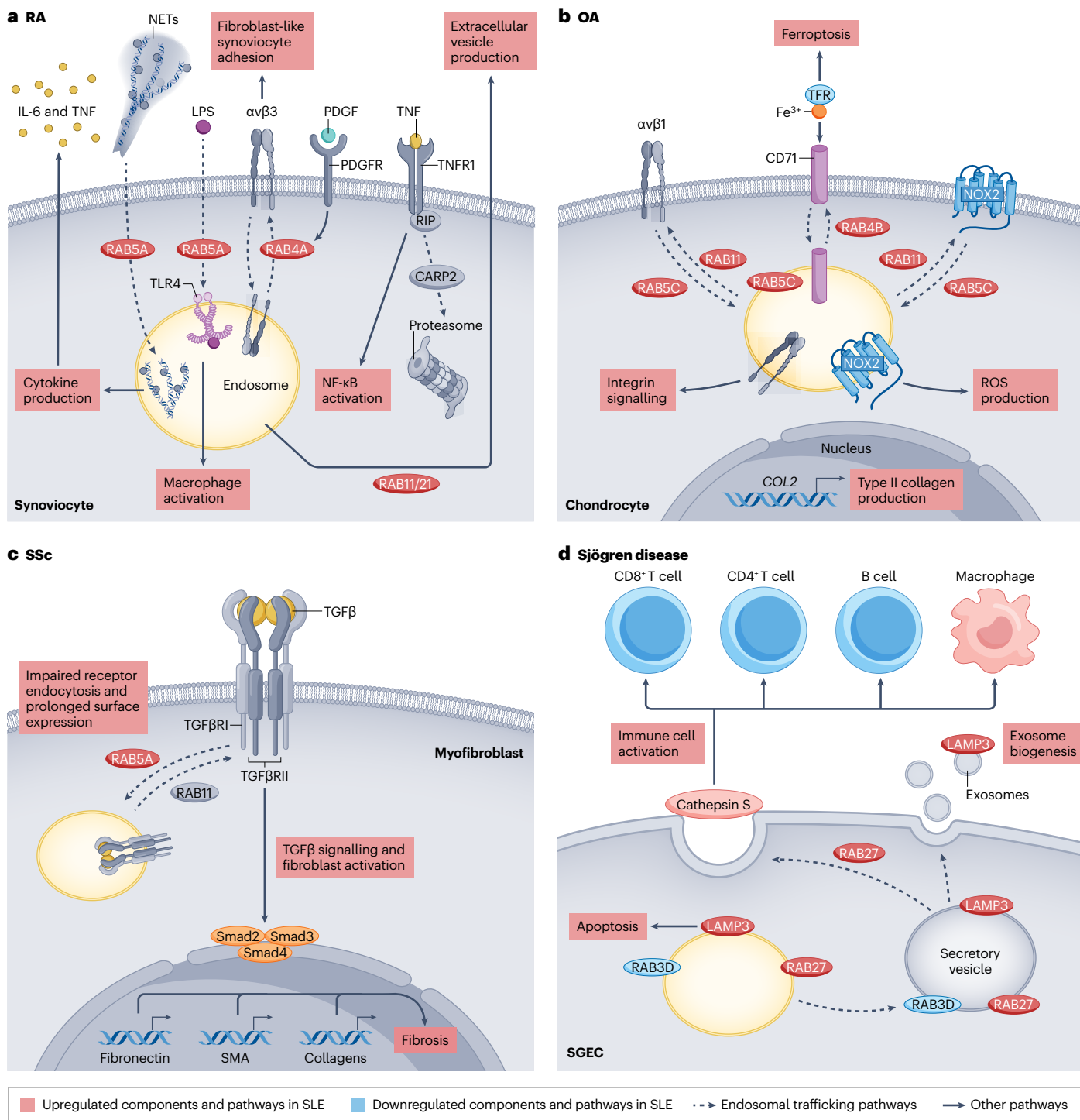
Lupus nephritis, a severe manifestation of SLE, arises from infiltration of the kidney by cells of the innate and adaptive immune system¹⁹⁴, deposition of immune complexes¹⁹⁵ and intrinsic defects in renal cell homeostasis, potentially involving dysregulated endosomal traffic¹⁹⁶. Disruption of endosomal protein sorting impairs podocyte integrity and contributes to glomerular injury in humans and mouse models

of nephritis^{162,197}. Deletion of key endocytic traffic regulators, including class III phosphatidylinositol 3-kinase vacuolar protein sorting 34 (Vps34), dynamin and synaptojanin, results in aberrant endosomal membrane morphology in podocytes, accompanied by foot process effacement and proteinuria in mouse models^{198,199}. Endosomal uptake of IgG via the natural Fc receptor (FcRn) is required for immune complex-induced autoimmunity²⁰⁰ and glomerulonephritis²⁰¹. Deletion of CD2AP, an endosomal protein that is critical for stabilizing contacts between T cells and APCs, leads to congenital nephritis syndrome²⁰². Notably, both FcRn²⁰³ and CD2AP interact with RAB4A in T cells⁷ and podocytes²⁰⁴. Whether activation of RAB4A directly affects podocyte function in SLE remains to be determined³.

Rheumatoid arthritis

In rheumatoid arthritis, endosomal trafficking regulates integrin recycling, which contributes to persistent fibroblast migration and cartilage destruction. The α v β 3 integrin, expressed on activated macrophages, osteoclasts and endothelial cells²⁰⁵, mediates adhesion of fibroblast-like synoviocytes (FLSs) in rheumatoid arthritis joints²⁰⁶. Platelet-derived growth factor (PDGF) activates FLSs in rheumatoid arthritis joints^{207,208}, and is involved in RAB4-dependent recycling of α v β 3 integrins (Fig. 6a).

Review article



RAB4 mediates the recycling of $\alpha v \beta 3$ integrin from early endosomes to the plasma membrane, supporting cell migration^{209,210}. Further downstream, the RAB4 effector protein Rabip4 mediates cell migration, as shown in NIH 3T3 fibroblasts²¹⁰. Furthermore, deficiency of RAB44, another regulator of endocytic recycling, blocks the development of collagen antibody-induced arthritis in a mouse model of rheumatoid arthritis¹⁰⁹.

Dysregulation of TNF receptor 1 (TNFR1) recycling sustains NF- κ B activation in rheumatoid arthritis. Upon TNF stimulation, receptor-interacting protein kinase (RIP) – a death-domain-containing serine–threonine kinase – is heavily ubiquitinated at early cell-surface TNFR1 complexes. The ubiquitin ligase CARP2, which contains a phospholipid-binding domain, localizes to endocytic vesicles and is recruited to internalized early TNFR1 complexes, where CARP2

Fig. 6 | Endosome trafficking in other rheumatic diseases. Involvement of endosome trafficking in rheumatoid arthritis (RA), osteoarthritis (OA), systemic sclerosis (SSc) and Sjögren disease. **a**, In RA synoviocytes (such as macrophages and fibroblast-like synoviocytes), endosomal trafficking promotes various pathogenic processes. For example, neutrophil extracellular traps (NETs) are internalized in macrophages via RAB5A-trafficked endosomes, inducing the production of pro-inflammatory cytokines such as IL-6 and TNF. Lipopolysaccharide (LPS)-induced Toll-like receptor 4 (TLR4)-mediated macrophage activation also depends on RAB5A-mediated endocytic internalization. Integrin trafficking sustains fibroblast migration and cartilage destruction; α v β 3 integrin, expressed on activated macrophages, osteoclasts and endothelial cells²⁰⁵, mediates fibroblast-like synoviocyte adhesion²⁰⁶ and is recycled by RAB4 in response to platelet-derived growth factor (PDGF) signalling^{209,210}, promoting migration. Dysregulation of TNF receptor 1 (TNFR1) recycling sustains NF- κ B activation in RA. CARP2, a ubiquitin protein ligase on endocytic vesicles, limits TNF-induced NF- κ B activation by targeting internalized TNFR1 for proteasome-mediated degradation²¹¹. Extracellular vesicles from arthritic mice express glucose transporter 1 (GLUT1)²¹², which is trafficked via endosomal pathways involving RAB11 (ref. 215) and RAB21 (ref. 216). **b**, OA chondrocytes exhibit RAB5C-dependent reactive oxygen species (ROS) production¹⁶⁰; NADPH oxidase 2 (NOX2) is a major source of ROS and is regulated via endosomal traffic by RAB5C and RAB11. ROS-producing endosomes (called redoxosomes) contain NOX2 and mediate α v β 1 integrin signalling. Type II collagen expression is also linked to RAB5A; chondrogenic progenitor cells that

lack RAB5C overexpress multiple chondrogenic markers and exhibit increased synthesis and deposition of collagen type II (COL2)²¹⁹. Additionally, RAB4B is overexpressed in OA synovial tissue; however, its protective or permissive role in pathogenesis needs clarification²²⁰. Chondrocytes express CD71 (ref. 221), which might contribute to their loss in OA via ferroptosis²²². **c**, SSc is characterized by excessive collagen synthesis resulting in fibrosis across multiple organ systems²²³. Under normal conditions, transforming growth factor- β (TGF β) receptors (TGF β R) are internalized through clathrin-mediated endocytosis by RAB5A²²⁵, and then returned to the plasma membrane via a RAB11-dependent mechanism²²⁶. In SSc, impaired endocytosis of TGF β prolongs surface TGF β signalling, promoting fibroblast activation and fibrosis²²⁴. **d**, Sjögren disease is characterized by increased exosome biogenesis. LAMP3 (also known as CD63) is a component of both early and late endosomes and exosomes, and is overexpressed in the salivary gland epithelial cells (SGECs) of people with Sjögren disease, which predisposes these cells to apoptosis²³¹. Concurrently, loss of RAB3D from secretory vesicles and increased RAB27-mediated release of cathepsin S in tears amplify local inflammation²³³. Cathepsin S functions as a potent activator of T cells, B cells and other immune cells, fuelling immune responses and perpetuating glandular damage²³⁴. For all the panels, molecules shown in red are upregulated in disease, those shown in blue are downregulated in disease; dashed arrows indicate endosome trafficking and solid arrows represent other pathways. RIP, receptor-interacting protein kinase; TFR, transferrin; TNFR1, TNF receptor 1.

targets RIP for proteasome-mediated degradation, thereby limiting TNF-induced NF- κ B activation²¹¹ (Fig. 6a). CARP2 might thus function as a negative regulator of NF- κ B-driven inflammation in rheumatoid arthritis by modulating endosomal trafficking²¹¹.

Extracellular vesicles are also implicated in rheumatoid arthritis. Extracellular vesicles from arthritic mice express the joint-homing receptor α v β 3 integrin²¹². Extracellular vesicles from both arthritic and healthy mice preferentially migrate to the inflamed synovia and avoid healthy joints²¹². Similarly, plasma extracellular vesicles from patients with rheumatoid arthritis overexpress α V integrin, and these vesicles are efficiently internalized by LPS-activated or TNF-activated human synovial cells in vitro. Extracellular vesicles accumulate in both the plasma and synovial fluid of patients with rheumatoid arthritis²¹³. Extracellular vesicles contribute to cell-to-cell communication and have a profound effect on immune responses, affecting the production of TNF, IL-6 and IL-1 β by monocytes and macrophages²¹⁴. Importantly, extracellular vesicles from arthritic mice express GLUT1 (ref. 212), which is trafficked via endosomal pathways involving RAB11 (ref. 215) and RAB21 (ref. 216). Taken together, these findings indicate that these extracellular vesicles might spread inflammation through the bloodstream of patients with rheumatoid arthritis.

Osteoarthritis

Metabolic dysfunction, including oxidative stress, has long been implicated in osteoarthritis²¹⁷. Reactive oxygen species (ROS)-producing endosomes (called redoxosomes) contain NADPH oxidase 2 (NOX2), which mediates α v β 1 integrin signalling and induces matrix metalloproteinase production in chondrocytes in osteoarthritis¹⁶⁰. Integrins connect and embed chondrocytes into the extracellular cartilage matrix²¹⁸. RAB5C expression is elevated in ROS-producing chondrocytes in osteoarthritis, suggesting a role for receptor internalization via endosome trafficking¹⁶⁰. Chondrogenic progenitor cells, present in osteoarthritis cartilage, contribute to fibrocartilaginous extracellular matrix production. Notably, chondrogenic progenitor cells

lacking RAB5C overexpress multiple chondrogenic markers and exhibit increased synthesis and deposition of type II collagen²¹⁹. These findings identify RAB5C as a promising therapeutic target to retain chondrocyte integrity and boost type II collagen production in osteoarthritis (Fig. 6b). RAB4B is also overexpressed in osteoarthritis synovial tissue; however, its role in osteoarthritis – whether protective or pathogenic – remains to be clarified²²⁰. Chondrocytes also express CD71 (ref. 221), which might contribute to the loss of chondrocytes via ferroptosis in osteoarthritis²²². Thus, targeting CD71-mediated iron uptake could represent a novel therapeutic strategy for osteoarthritis (Table 1).

Systemic sclerosis

Systemic sclerosis (SSc; also referred to as scleroderma) is characterized by excessive collagen synthesis and widespread fibrosis across multiple organ systems²²³. A key driver of this fibrotic process is impaired endocytosis of transforming growth factor- β (TGF β) receptors²²⁴, which prolongs surface signalling and sustains fibroblast activation²²⁴. TGF β receptors are internalized through clathrin-coated pits in a RAB5A-dependent manner²²⁵, and recycled back to the plasma membrane through RAB11-mediated traffic²²⁶ (Fig. 6c). Additionally, TGF β receptor I might also undergo caveolin-mediated internalization²²⁷. Importantly, caveolin 1 deficiency disrupts this pathway, resulting in increased receptor retention at the cell surface and prolonged TGF β signalling²²⁸, thereby promoting excessive extracellular matrix deposition²²⁹. These findings highlight that both major endocytic pathways (clathrin-mediated endocytosis and caveolin-mediated endocytosis) can independently mediate the internalization of TGF β receptors. Moreover, clathrin-mediated and caveolin-mediated endocytic pathways can converge during TGF β receptor endocytic trafficking²²⁷. Beyond TGF β signalling, endosomal pathways also influence immune responses in SSc. For example, the expression of endosome-bound sensor TLR9 is increased in patients with SSc and correlates with enhanced type I interferon signalling²³⁰.

Table 1 | Endosome traffic-directed therapeutic targets in rheumatic diseases

Drug	Target	Mechanism	Disease context	Refs.
Targeting RAB GTPases				
3-PEHPC	GGT-II and RAB4A	Endosomal traffic modulation	SLE	31
Neoandrographolide	RAB5A	Inhibition of osteoclast proliferation	OA	254
Targeting TLRs				
Enpatoran (M5049)	TLR7 and TLR8	TLR inhibition and IGS suppression	SLE	104
E6742	TLR7 and TLR8	TLR inhibition and IGS suppression	SLE	247
MHV370	TLR7 and TLR8	TLR inhibition and IGS suppression	SLE	248,249
Modulation of autophagy				
Hydroxychloroquine	Lysosomal v-ATPases	Autophagy inhibition	RA and SLE	268
Rapamycin	mTORC1 (via FKBP12)	mTOR inhibition	SLE	185,261,262, 269–272
Eltrombopag	TFEB	Suppression of autophagy through transcriptional inhibition of TFEB	SLE, ITP and APS	265–267
Modulation of metabolism				
Daratumumab	CD38	Disruption of NAD and calcium metabolism	SLE	188
PPMX-TO03	CD71	Disruption of iron metabolism	SLE and PCV	136,251
IGN523	CD98	Inhibition of tryptophan, kynurenine and BCAA transport	SLE and AML	3,273

AML, acute myeloid leukaemia; APS, antiphospholipid syndrome; BCAA, branched-chain amino acids; GGT-II, geranyl-geranyl transferase type II; IGS, interferon gene signature; ITP, immune thrombocytopenia; mTORC1, mechanistic target of rapamycin complex 1; OA, osteoarthritis; PCV, polycythemia vera; RA, rheumatoid arthritis; SLE, systemic lupus erythematosus; TFEB, transcription factor EB; TLR, Toll-like receptor.

Sjögren syndrome

Sjögren syndrome is an autoimmune sialadenitis of unknown aetiology, characterized by inflammation and dysfunction of the salivary glands. As previously discussed, LAMP3 has a central role in exosome biogenesis. This tetraspanin protein is a component of both early and late endosomes. Within lysosomes, LAMP3 contributes to degradation processes. LAMP3 is also present in exosomes and is used as a reliable marker for these extracellular vesicles⁵⁵ (Fig. 6d). In patients with Sjögren syndrome, LAMP3 is overexpressed in salivary gland epithelial cells (SGECs), increasing the susceptibility of the cells to apoptosis²³¹. Ectopic, adeno-associated virus-mediated overexpression of LAMP3 enhances SGEC apoptosis and reduces the expression of proteins involved in saliva secretion. Damaged SGECs release DAMPs, which activate immune cells via TLRs. These findings highlight the pathogenic role of distorted endosome trafficking in cells of target organs²³². Disrupted endosomal and lysosomal trafficking, along with impaired autophagy and increased apoptosis, promotes exosomal release of DAMPs into the extracellular space. These DAMPs activate immune cells, specifically monocytes and macrophages, via TLRs, that amplify the inflammatory cascade in the salivary glands, attracting more immune cells characteristic of Sjögren syndrome²³². In parallel, loss of RAB3D from secretory vesicles coincides with enhanced RAB27-mediated release of pro-inflammatory cathepsin S in the tears of patients with Sjögren syndrome²³³ (Fig. 6d). Cathepsins are potent activators of T cells and B cells, driving inflammation²³⁴. However, cathepsin S inhibition failed to demonstrate clinical efficacy in a controlled clinical trial²³⁵.

Macrophage activation syndrome

Macrophage activation syndrome (MAS) is a severe, life-threatening hyperinflammatory condition that arises as a form of secondary haemophagocytic lymphohistiocytosis (HLH), typically in the context

of rheumatic diseases or haematological malignancies. Mutations of *RAB27A* cause Griscelli syndrome type 2, a genetic form of HLH²³⁶. *RAB27A* is crucial for exocytosis of lytic granules by cytotoxic T cells and natural killer cells²³⁶, directing late endosomal perforin-containing and granzyme-containing vesicles to the plasma membrane for targeted delivery to virus-infected or malignant cells via exosome secretion²³⁷. Loss-of-function mutations in *RAB27A* impair this cytotoxic mechanism, resulting in defective clearance of activated cells and persistent immune stimulation. In addition to impaired cytotoxicity, *RAB27A* deficiency promotes dysregulated phagocytosis by macrophages²³⁸, prolonging the presence of phagocytosed targets in macrophages and eliciting persistent and uncontrolled activation of these cells. This process promotes haemophagocytosis and heightened production of pro-inflammatory cytokines, creating a 'cytokine storm' characteristic of both HLH and MAS²³⁹. Heterozygous *RAB27A* mutations might present with delayed-onset HLH or MAS in adulthood²⁴⁰. Indeed, secondary HLH, including MAS, is about tenfold more common than the primary form and typically presents in patients with uncontrolled rheumatic disease extending into adulthood^{239,241}.

Therapeutic implications and opportunities

Several emerging therapeutic strategies aim to restore endosomal homeostasis or target trafficking checkpoints to dampen inflammation in rheumatic diseases (Table 1). Hydroxychloroquine, a cationic amphiphilic compound and mainstay treatment of SLE, rheumatoid arthritis and Sjögren disease, exerts its immunomodulatory effects by increasing endosomal pH²⁴², thereby inhibiting autophagy and TLR7 activation²⁴³. Further downstream, this drug can thus suppress type I interferon production in primary pDCs from both healthy individuals and patients with SLE²⁴⁴.

Small-molecule inhibitors of endosome trafficking are under investigation. These inhibitors include compounds directed against endosome-embedded proteins, such as UNC93B1 (ref. 245) and RAB GTPases³¹, as well as upstream regulators, including mTOR and TFEB. For example, the RAB geranyl-geranyl transferase inhibitor 3-PEHPC disrupts RAB4A and RAB5A trafficking in vitro and attenuates ANA production, proteinuria and glomerulonephritis in MRL/lpr lupus-prone mice in vivo³¹. Similarly, the competitive RAB7 inhibitor CID-1067700, which interferes with GTP binding, reduces secretion of several inflammatory cytokines, including IL-1 α , IL-8, IL-17A and IL-32, in adipocyte-derived mesenchymal stem cells (ASC52tel) in vitro²⁴⁶, and suppresses B cell development and autoantibody production in vivo in MRL/lpr mice³⁰.

A series of dual inhibitors targeting endosome-bound TLR7 and TLR8 has shown preliminary clinical efficacy in SLE. In a double-blind placebo-controlled clinical trial, E6742 was found to be safe and was well tolerated, showing suppression of the interferon gene signature and signals of therapeutic efficacy signals²⁴⁷. MHV370, a selective oral inhibitor of TLR7 and TLR8, also restrained the interferon gene signature in mice and patients with SLE in vitro, and was well tolerated in healthy adults^{248,249}. Enpatoran, formerly called M5049, exhibited therapeutic efficacy in lupus-prone mice²⁵⁰ and preliminary evidence of safety and efficacy in human trials¹⁰⁴.

Various inhibitors that target endosomal mediators involved in cell metabolism are also under evaluation. For example, daratumumab, an antibody targeting endosome-trafficked CD38 (ref. 186) that is currently used in the treatment of myeloma, showed clinical benefit in six patients with refractory lupus nephritis¹⁸⁸. Monoclonal antibodies directed against CD71 (ref. 251) and CD98 (ref. 249) – both trafficked via RAB4A³ – are currently being evaluated in phase I clinical trials.

Natural compound screening has identified neoandrographolide (NAP), derived from the plant *Andrographis paniculate*, as a potential modulator of endosome traffic. Among 7,459 natural compounds screened, NAP was found to competitively bind the GDP–GTP groove on the surface of RAB5, thereby reducing its GTPase activity and inhibiting cell proliferation in vitro²⁵². In addition to its effects on RAB5, NAP suppresses the production of nitric oxide and TNF in activated macrophages²⁵³ and inhibits the formation of mature osteoclasts²⁵⁴, both processes relevant to inflammation and tissue destruction in rheumatic diseases. RAB5A is overexpressed in T cells both in patients with SLE⁶⁰ and lupus-prone mice³¹, providing further rationale for assessing the therapeutic potential of NAP in autoimmunity. Moreover, RAB5C expression is increased in ROS-producing chondrocytes in osteoarthritis¹⁶⁰. Given that the amino acid sequences of RAB5A and RAB5C overlap by 86%, with 100% identity within the GTP-binding domain (Supplementary Fig. 1C), NAP could have therapeutic benefit for a broad range of rheumatic diseases.

Pharmacological inhibition of exosome release and uptake has shown protective effects in osteoarthritis²⁵⁵ but potentially enhances inflammatory signalling in rheumatoid arthritis²⁵⁶. For example, GW4869, a widely used small-molecule inhibitor of neutral sphingomyelinase (a critical enzyme for exosome formation and release²⁵⁵), has demonstrated therapeutic benefit in mouse models of osteoarthritis²⁵⁷. Treatment with GW4869 increases type II collagen expression and reduces MMP3 expression in cartilage tissues of mice²⁵⁷. Moreover, GW4869 blocks the secretion of osteoclast-derived exosomal miR-212-3p, which otherwise suppresses anabolic activity and promotes catabolism in chondrocytes from mice and patients with osteoarthritis²⁵⁸. Beyond osteoarthritis, GW4869 also blocks the production of pro-inflammatory cytokines by macrophages and protects

against myocarditis in the setting of sepsis²⁵⁹. In mice with myosin heavy-chain-directed experimental autoimmune myocarditis, serum exosomes carrying miRNA-142 induce immunometabolic dysfunction in CD4 T cells, contributing to disease pathogenesis. Pharmacological inhibition of exosome secretion with GW4869 mitigates disease severity, underscoring the indispensable role of exosomes in disease development⁵⁸. By contrast, FLSs in rheumatoid arthritis secrete circular RNAs that promote anti-inflammatory polarization of macrophages²⁵⁶. Suppressing exosome release in this context might therefore inadvertently disrupt protective immune regulation. These findings highlight the importance of tailoring exosome-targeted therapies to the specific immunopathological context of each rheumatic disease.

TFEB is required for endosome formation during autophagy⁶² (Fig. 3a). T_{reg} cells depend on autophagy for survival and, consistent with the role of autophagy in T_{reg} cell development, TFEB responds to T_{reg} cell-related cues in an mTORC1-dependent manner²⁶⁰. In SLE, autophagy is generally restrained in T_{reg} cells but increased in effector CD4 T cells¹⁷². Notably, rapamycin, which inhibits the mTOR pathway and thereby promotes autophagy, has been shown to enhance autophagic activity in T_{reg} cells in SLE¹⁷². This effect is associated with clinical improvement^{185,261–263}. Eltrombopag, an FDA-approved thrombopoietin receptor agonist, has emerged as a direct modulator of TFEB. Eltrombopag binds to the basic helix–loop–helix leucine zipper domain of TFEB, disrupting its DNA-binding activity both in vitro and in vivo²⁶⁴. This interaction selectively inhibits the transcriptional activity of TFEB on a genomic scale and blocks autophagy in a dose-dependent manner, opening up new possibilities for cell-type-specific modulation of autophagy¹⁷². Several case series and retrospective studies have reported potential therapeutic benefit of eltrombopag in SLE²⁶⁵, immune thrombocytopenia²⁶⁶ and antiphospholipid syndrome²⁶⁷, which warrant further investigation in prospective confirmatory studies. However, given that autophagy dysfunction is specific to cell type in SLE¹⁷², systemic blockade of TFEB might be counterproductive in a subset of patients.

Conclusions

Endosomal trafficking has emerged as a central regulator of immune function and tissue homeostasis in rheumatic diseases. Beyond its canonical roles in intracellular sorting and exosome biogenesis, endosomal processing orchestrates antigen presentation, receptor signalling, cytokine secretion and cellular metabolism, processes that collectively shape the inflammatory landscape. Dysregulation of these pathways can lead to abnormal immune responses, inflammation and tissue damage. Continued elucidation of endosomal trafficking regulators across specific immune and end-organ cell subsets – using tools such as imaging flow cytometry, confocal microscopy and single-cell multi-omics – will deepen mechanistic understanding of rheumatic disease pathogenesis and guide the development of precision therapies. A more complete understanding of endosomal pathways in immune and stromal compartments holds promise for restoring immune tolerance and enhancing end-organ resistance, offering new avenues for durable disease control in rheumatic diseases.

Published online: 8 December 2025

References

1. Park, J. S. & Perl, A. Endosome traffic modulates pro-inflammatory signal transduction in CD4⁺ T cells – implications for the pathogenesis of systemic lupus erythematosus. *Int. J. Mol. Sci.* **24**, 10749 (2023).
2. Belabed, M. et al. Kinesin-1 regulates antigen cross-presentation through the scission of tubulations from early endosomes in dendritic cells. *Nat. Commun.* **11**, 1817 (2020).

3. Huang, N. et al. Rab4A-directed endosome traffic shapes pro-inflammatory mitochondrial metabolism in T cells via mitophagy, CD98 expression, and kynurenine-sensitive mTOR activation. *Nat. Commun.* **15**, 2598 (2024).
4. Manderson, A. P., Kay, J. G., Hammond, L. A., Brown, D. L. & Stow, J. L. Subcompartments of the macrophage recycling endosome direct the differential secretion of IL-6 and TNF α . *J. Cell Biol.* **178**, 57–69 (2007).
5. Stanley, A. C. et al. Recycling endosome-dependent and -independent mechanisms for IL-10 secretion in LPS-activated macrophages. *J. Leukoc. Biol.* **92**, 1227–1239 (2012).
6. Pfeffer, S. R. Membrane domains in the secretory and endocytic pathways. *Cell* **112**, 507–517 (2003).
7. Klumperman, J. & Raposo, G. The complex ultrastructure of the endolysosomal system. *Cold Spring Harb. Perspect. Biol.* **6**, a016857 (2014).
8. Conte-Zerial, P. et al. Membrane identity and GTPase cascades regulated by toggle and cut-out switches. *Mol. Syst. Biol.* **4**, 206 (2008).
9. Pfeffer, S. R. Rab GTPases: master regulators that establish the secretory and endocytic pathways. *Mol. Biol. Cell* **28**, 712–715 (2017).
10. Wandinger-Ness, A. & Zerial, M. Rab proteins and the compartmentalization of the endosomal system. *Cold Spring Harb. Perspect. Biol.* **6**, a022616 (2014).
11. Borchers, A. C., Langemeyer, L. & Ungermann, C. Who is in control? Principles of Rab GTPase activation in endolysosomal membrane trafficking and beyond. *J. Cell. Biol.* **220**, e202105120 (2021).
12. Maxfield, F. R. Role of endosomes and lysosomes in human disease. *Cold Spring Harb. Perspect. Biol.* **6**, a016931 (2014).
13. Huotari, J. & Helenius, A. Endosome maturation. *EMBO J.* **30**, 3481–3500 (2011).
14. Grant, B. D. & Donaldson, J. G. Pathways and mechanisms of endocytic recycling. *Nat. Rev. Mol. Cell Biol.* **10**, 597–608 (2009).
15. Eyster, C. A. et al. Discovery of new cargo proteins that enter cells through clathrin-independent endocytosis 14. *Traffic* **10**, 590–599 (2009).
16. Prichard, K. L., O'Brien, N. S., Murcia, S. R., Baker, J. R. & McCluskey, A. Role of clathrin and dynamin in clathrin mediated endocytosis/synaptic vesicle recycling and implications in neurological diseases. *Front. Cell Neurosci.* **15**, 754110 (2021).
17. Bitsikas, V., Correa, I. R. Jr. & Nichols, B. J. Clathrin-independent pathways do not contribute significantly to endocytic flux. *elife* **3**, e03970 (2014).
18. Langemeyer, L., Fröhlich, F. & Ungermann, C. Rab GTPase function in endosome and lysosome biogenesis. *Trends Cell Biol.* **28**, 957–970 (2018).
19. Fratti, R. A., Backer, J. M., Gruenberg, J., Corvera, S. & Deretic, V. Role of phosphatidylinositol 3-kinase and Rab5 effectors in phagosomal biogenesis and mycobacterial phagosome maturation arrest. *J. Cell Biol.* **154**, 631–644 (2001).
20. Hunyady, L. et al. Differential PI 3-kinase dependence of early and late phases of recycling of the internalized AT1 angiotensin receptor. *J. Cell Biol.* **157**, 1211–1222 (2002).
21. Shi, J. et al. HIV-1 Nef antagonizes SERINC5 restriction by downregulation of serinc5 via the endosome/lysosome system. *J. Virol.* **92**, e00196-18 (2018).
22. Powelka, A. M. et al. Stimulation-dependent recycling of integrin β 1 regulated by ARF6 and Rab11. *Traffic* **5**, 20–36 (2004).
23. Tanevski, J., Todorovski, L., Kalaidzidis, Y. & Džeroski, S. Domain-specific model selection for structural identification of the Rab5-Rab7 dynamics in endocytosis. *BMC Syst. Biol.* **9**, 31 (2015).
24. Hutagalung, A. H. & Novick, P. J. Role of Rab GTPases in membrane traffic and cell physiology. *Physiol. Rev.* **91**, 119–149 (2011).
25. Agola, J. O., Jim, P. A., Ward, H. H., Basuray, S. & Wandinger-Ness, A. Rab GTPases as regulators of endocytosis, targets of disease and therapeutic opportunities. *Clin. Genet.* **80**, 305–318 (2011).
26. D'Souza, R. S. et al. Rab4 orchestrates a small GTPase cascade for recruitment of adaptor proteins to early endosomes. *Curr. Biol.* **24**, 1187–1198 (2014).
27. Shibolet, O. et al. AKAP13, a RhoA GTPase-specific guanine exchange factor, is a novel regulator of TLR2 signaling. *J. Biol. Chem.* **282**, 35308–35317 (2007).
28. Monzo, P. et al. In *Methods in Enzymology GTPases Regulating Membrane Targeting and Fusion* vol. 403 (eds Balch, W. E., Der, C. J. & Hall, A.) 107–118 (Academic Press, 2005).
29. Furusawa, K. et al. CD2-associated protein (CD2AP) overexpression accelerates amyloid precursor protein (APP) transfer from early endosomes to the lysosomal degradation pathway. *J. Biol. Chem.* **294**, 10886–10899 (2019).
30. Lam, T. et al. Small molecule inhibition of Rab7 impairs B cell class switching and plasma cell survival to dampen the autoantibody response in murine lupus. *J. Immunol.* **197**, 3792–3805 (2016).
31. Caza, T. N. et al. HRES-1/RAB4-mediated depletion of DRP1 impairs mitochondrial homeostasis and represents a target for treatment in SLE. *Ann. Rheum. Dis.* **73**, 1887–1897 (2014).
32. Nag, S. et al. Rab4A organizes endosomal domains for sorting cargo to lysosome-related organelles. *J. Cell Sci.* **131**, jcs216226 (2018).
33. Vidal, M. J. & Stahl, P. D. The small GTP-binding proteins Rab4 and ARF are associated with released exosomes during reticulocyte maturation. *Eur. J. Cell. Biol.* **60**, 261–267 (1993).
34. Mathieu, M. et al. Specificities of exosome versus small ectosome secretion revealed by live intracellular tracking of CD63 and CD9. *Nat. Commun.* **12**, 4389 (2021).
35. Rossatti, P. et al. Rapid increase in transferrin receptor recycling promotes T cell activation. *BMC Biol.* **20**, 189 (2022).
36. Daro, E., van der, S. P., Galli, T. & Mellman, I. Rab4 and cellubrevin define different early endosome populations on the pathway of transferrin receptor recycling. *Proc. Natl Acad. Sci. USA* **93**, 9559–9564 (1996).
37. van der Sluijs, P. et al. The small GTP-binding protein rab4 controls an early sorting event on the endocytic pathway. *Cell* **70**, 729–740 (1992).
38. Nagy, G. et al. Regulation of CD4 expression via recycling by HRES-1/RAB4 controls susceptibility to HIV infection. *J. Biol. Chem.* **281**, 34574–34591 (2006).
39. Ren, M. et al. Hydrolysis of GTP on rab11 is required for the direct delivery of transferrin from the pericentriolar recycling compartment to the cell surface but not from sorting endosomes. *Proc. Natl Acad. Sci. USA* **95**, 6187–6192 (1998).
40. Deffieu, M. S. et al. Rab7-harboring vesicles are carriers of the transferrin receptor through the biosynthetic secretory pathway. *Sci. Adv.* **7**, eaba7803 (2021).
41. Li, H. et al. A Bcl-xL-Drp1 complex regulates synaptic vesicle membrane dynamics during endocytosis. *Nat. Cell. Biol.* **15**, 773–785 (2013).
42. Gomes, L. C., Benedetto, G. D. & Scorrano, L. During autophagy mitochondria elongate, are spared from degradation and sustain cell viability. *Nat. Cell Biol.* **13**, 589–598 (2011).
43. Rambold, A. S., Kostecky, B., Elia, N. & Lippincott-Schwartz, J. Tubular network formation protects mitochondria from autophagosomal degradation during nutrient starvation. *Proc. Natl Acad. Sci. USA* **108**, 10190–10195 (2011).
44. Talaber, G. et al. HRES-1/Rab4 promotes the formation of LC3⁺ autophagosomes and the accumulation of mitochondria during autophagy. *PLoS One* **9**, e84392 (2014).
45. Goldmann, O. & Medina, E. Revisiting pathogen exploitation of clathrin-independent endocytosis: mechanisms and implications. *Cells* **14**, 731 (2025).
46. Cohen, A. W., Hnasko, R., Schubert, W. & Lisanti, M. P. Role of caveolae and caveolins in health and disease. *Physiol. Rev.* **84**, 1341–1379 (2004).
47. Wu, B. et al. Basigin-mediated redistribution of CD98 promotes cell spreading and tumorigenicity in hepatocellular carcinoma. *J. Exp. Clin. Cancer Res.* **34**, 110 (2015).
48. Eyster, C. A. et al. Discovery of new cargo proteins that enter cells through clathrin-independent endocytosis. *Traffic* **10**, 590–599 (2009).
49. Doyle, L. M. & Wang, M. Z. Overview of extracellular vesicles, their origin, composition, purpose, and methods for exosome isolation and analysis. *Cells* **8**, 727 (2019).
50. Wang, Y., Xiao, T., Zhao, C. & Li, G. The regulation of exosome generation and function in physiological and pathological processes. *Int. J. Mol. Sci.* **25**, 255 (2023).
51. Fortin, P. R. et al. Distinct subtypes of microparticle-containing immune complexes are associated with disease activity, damage, and carotid intima-media thickness in systemic lupus erythematosus. *J. Rheumatol.* **43**, 2019–2025 (2016).
52. Zhao, Y., Wei, W. & Liu, M.-L. Extracellular vesicles and lupus nephritis — new insights into pathophysiology and clinical implications. *J. Autoimmun.* **115**, 102540 (2020).
53. Zhang, R. et al. An optimized exosome production strategy for enhanced yield while without sacrificing cargo loading efficiency. *J. Nanobiotechnol.* **20**, 463 (2022).
54. Schulze, U. et al. Tetraspanin CD63 controls basolateral sorting of organic cation transporter 2 in renal proximal tubules. *FASEB J.* **31**, 1421–1433 (2017).
55. Kobayashi, T. et al. The tetraspanin CD63/lamp3 cycles between endocytic and secretory compartments in human endothelial cells. *Mol. Biol. Cell* **11**, 1829–1843 (2000).
56. Verweij, F. J. et al. ER membrane contact sites support endosomal small GTPase conversion for exosome secretion. *J. Cell Biol.* **221**, e202112032 (2022).
57. Salvi, V. et al. Exosome-delivered microRNAs promote IFN- α secretion by human plasmacytoid DCs via TLR7. *JCI Insight* **3**, e98204 (2018).
58. Sun, P. et al. Circulating exosomes control CD4⁺ T cell immunometabolic functions via the transfer of miR-142 as a novel mediator in myocarditis. *Mol. Ther.* **28**, 2605–2620 (2020).
59. Mobarrez, F. et al. Microparticles in the blood of patients with SLE: size, content of mitochondria and role in circulating immune complexes. *J. Autoimmun.* **102**, 142–149 (2019).
60. Fernandez, D. R. et al. Activation of mTOR controls the loss of TCR in lupus T cells through HRES-1/Rab4-regulated lysosomal degradation. *J. Immunol.* **182**, 2063–2073 (2009).
61. Birgisdottir, Å. B. & Johansen, T. Autophagy and endocytosis - interconnections and interdependencies. *J. Cell Sci.* **133**, jcs228114 (2020).
62. Nnah, I. C. et al. TFEB-driven endocytosis coordinates MTORC1 signaling and autophagy. *Autophagy* **15**, 151–164 (2019).
63. Martina, J. A., Chen, Y., Gucek, M. & Puertollano, R. MTORC1 functions as a transcriptional regulator of autophagy by preventing nuclear transport of TFEB. *Autophagy* **8**, 903–914 (2012).
64. Settembre, C. et al. A lysosome-to-nucleus signalling mechanism senses and regulates the lysosome via mTOR and TFEB. *EMBO J.* **31**, 1095–1108 (2012).
65. Palmieri, M. et al. Characterization of the CLEAR network reveals an integrated control of cellular clearance pathways. *Hum. Mol. Genet.* **20**, 3852–3866 (2011).
66. Lee, B. L. et al. UNC93B1 mediates differential trafficking of endosomal TLRs. *eLife* **2**, e00291 (2013).
67. David, C. et al. Gain-of-function human UNC93B1 variants cause systemic lupus erythematosus and chilblain lupus. *J. Exp. Med.* **221**, e20232066 (2024).
68. Al-Azab, M. et al. Genetic variants in UNC93B1 predispose to childhood-onset systemic lupus erythematosus. *Nat. Immunol.* **25**, 969–980 (2024).
69. Leeb, T. et al. A missense variant affecting the C-terminal tail of UNC93B1 in dogs with exfoliative cutaneous lupus erythematosus (ECL). *Genes* **11**, 159 (2020).
70. Nedeljkovic, I. et al. COPD GWAS variant at 19q13.2 in relation with DNA methylation and gene expression. *Hum. Mol. Genet.* **27**, 396–405 (2018).
71. Wang, C., Sharma, N., Kessler, P. M. & Sen, G. C. Interferon induction by STING requires its translocation to the late endosomes. *Traffic* **24**, 576–586 (2023).

72. Hurles, M. Gene duplication: the genomic trade in spare parts. *PLoS Biol.* **2**, E206 (2004).
73. Mikamori, M. et al. MicroRNA-155 controls exosome synthesis and promotes gemcitabine resistance in pancreatic ductal adenocarcinoma. *Sci. Rep.* **7**, 42339 (2017).
74. Zhang, Z. et al. Distinct changes in endosomal composition promote NLRP3 inflammasome activation. *Nat. Immunol.* **24**, 30–41 (2023).
75. Lee, B. et al. Disruptions in endocytic traffic contribute to the activation of the NLRP3 inflammasome. *Sci. Signal.* **16**, eabm7134 (2023).
76. Li, R.-N. et al. NLRP3 gene polymorphisms in rheumatoid arthritis and primary Sjogren's syndrome patients. *Diagnostics* **13**, 206 (2023).
77. Cummings, R. D., Chiffolleau, E., van Kooyk, Y. & McEver, R. P. C-type lectins. In *Essentials of Glycobiology* (eds Varki, A. et al.) (Cold Spring Harbor Laboratory Press, 2022).
78. Skinningsrud, B. et al. A CLEC16A variant confers risk for juvenile idiopathic arthritis and anti-cyclic citrullinated peptide antibody negative rheumatoid arthritis. *Ann. Rheum. Dis.* **69**, 1471–1474 (2010).
79. Zoledziewska, M. et al. Variation within the CLEC16A gene shows consistent disease association with both multiple sclerosis and type 1 diabetes in Sardinia. *Genes Immun.* **10**, 15–17 (2009).
80. Márquez, A. et al. Specific association of a CLEC16A/KIAA0350 polymorphism with NOD2/CARD15 Crohn's disease patients. *Eur. J. Hum. Genet.* **17**, 1304–1308 (2009).
81. Eriksson, A. M. et al. Exploring the role of the multiple sclerosis susceptibility gene CLEC16A in T cells. *Scand. J. Immunol.* **94**, e13050 (2021).
82. Leikfoss, I. S. et al. Multiple sclerosis risk allele in CLEC16A acts as an expression quantitative trait locus for CLEC16A and SOCS1 in CD4⁺ T Cells. *PLoS One* **10**, e0132957 (2015).
83. Rallabandi, H. R., Singh, M. K., Looger, L. L. & Nath, S. K. Defining mechanistic links between the non-coding variant rs17673553 in CLEC16A and lupus susceptibility. *Int. J. Mol. Sci.* **26**, 314 (2025).
84. Tam, R. C. Y. et al. Human CLEC16A regulates autophagy through modulating mTOR activity. *Exp. Cell Res.* **352**, 304–312 (2017).
85. Soleimanpour, S. A. et al. The diabetes susceptibility gene Clec16a regulates mitophagy. *Cell* **157**, 1577–1590 (2014).
86. Pandey, R. et al. CLEC16A regulates splenocyte and NK cell function in part through MEK signaling. *PLoS One* **13**, e0203952 (2018).
87. Gonugunta, V. K. et al. Trafficking-mediated STING degradation requires sorting to acidified endolysosomes and can be targeted to enhance anti-tumor response. *Cell Rep.* **21**, 3234–3242 (2017).
88. Frémond, M.-L. et al. Overview of STING-associated vasculopathy with onset in infancy (SAVI) among 21 patients. *J. Allergy Clin. Immunol. Pract.* **9**, 803–818.e11 (2021).
89. Li, T. et al. A novel STING1-activating mutation is identified in a patient with childhood-onset systemic lupus erythematosus. *Clin. Immunol.* **276**, 110493 (2025).
90. Trivedi, S. & Greidinger, E. L. Endosomal Toll-like receptors in autoimmunity: mechanisms for clinical diversity. *Therapy* **6**, 433–442 (2009).
91. Bordon, Y. Innate immunity: TLR13, unlucky, but just for some. *Nat. Rev. Immunol.* **12**, 618–619 (2012).
92. Luchner, M., Reinke, S. & Milicic, A. TLR agonists as vaccine adjuvants targeting cancer and infectious diseases. *Pharmaceutics* **13**, 142 (2021).
93. Hemmi, H. et al. A Toll-like receptor recognizes bacterial DNA. *Nature* **408**, 740–745 (2000).
94. Sadeq, S., Al-Hashimi, S., Cusack, C. M. & Werner, A. Endogenous double-stranded RNA. *Noncoding RNA* **7**, 15 (2021).
95. Sakaniwa, K. et al. TLR3 forms a laterally aligned multimeric complex along double-stranded RNA for efficient signal transduction. *Nat. Commun.* **14**, 164 (2023).
96. Ivashkiv, L. B. & Donlin, L. T. Regulation of type I interferon responses. *Nat. Rev. Immunol.* **14**, 36–49 (2014).
97. Muskardin, T. L. W. & Niewold, T. B. Type I interferon in rheumatic diseases. *Nat. Rev. Rheumatol.* **14**, 214–228 (2018).
98. Ishida, H. et al. Cryo-EM structures of Toll-like receptors in complex with UNC93B1. *Nat. Struct. Mol. Biol.* **28**, 173–180 (2021).
99. Marshak-Rothstein, A. Toll-like receptors in systemic autoimmune disease. *Nat. Rev. Immunol.* **6**, 823–835 (2006).
100. Pollard, K. M. et al. Induction of systemic autoimmunity by a xenobiotic requires endosomal TLR trafficking and signaling from the late endosome and endolysosome but not type I IFN. *J. Immunol.* <https://doi.org/10.4049/jimmunol.1700332> (2017).
101. Fukui, R. et al. Unc93B1 restricts systemic lethal inflammation by orchestrating Toll-like receptor 7 and 9 trafficking. *Immunity* **35**, 69–81 (2011).
102. Wolf, C. et al. UNC93B1 variants underlie TLR7-dependent autoimmunity. *Sci. Immunol.* **9**, eadi9769 (2024).
103. Kawai, T. et al. Interferon- α induction through Toll-like receptors involves a direct interaction of IRF7 with MyD88 and TRAF6. *Nat. Immunol.* **5**, 1061–1068 (2004).
104. Port, A. et al. Phase 1 study in healthy participants of the safety, pharmacokinetics, and pharmacodynamics of enpatoran (M5049), a dual antagonist of toll-like receptors 7 and 8. *Pharmacol. Res. Perspect.* **9**, e00842 (2021).
105. Ramirez, G. A. et al. Infection-associated flares in systemic lupus erythematosus. *Pathogens* **13**, 934 (2024).
106. Perl, A. Mechanisms of viral pathogenesis in rheumatic diseases. *Ann. Rheum. Dis.* **58**, 454–461 (1999).
107. Spearman, P. Viral interactions with host cell Rab GTPases. *Small GTPases* **9**, 192–201 (2018).
108. Banerjee, M. et al. Platelets endocytose viral particles and are activated via TLR (Toll-Like Receptor) signaling. *Arterioscler. Thromb. Vasc. Biol.* **40**, 1635–1650 (2020).
109. Yamaguchi, Y. et al. Impaired development of collagen antibody-induced arthritis in Rab44-deficient mice. *Biomedicines* **12**, 2504 (2024).
110. Hu, F. et al. Toll-like receptors expressed by synovial fibroblasts perpetuate Th1 and Th17 cell responses in rheumatoid arthritis. *PLoS One* **9**, e100266 (2014).
111. Abdelwahab, A. et al. Differential synovial tissue expression of TLRs in seropositive and seronegative rheumatoid arthritis: a preliminary report. *Autoimmunity* **54**, 23–34 (2021).
112. Swain, N., Tripathy, A., Padhan, P., Raghav, S. K. & Gupta, B. Toll-like receptor-7 activation in CD8⁺ T cells modulates inflammatory mediators in patients with rheumatoid arthritis. *Rheumatol. Int.* **42**, 1235–1245 (2022).
113. Negishi, H. et al. Identification of U11snRNA as an endogenous agonist of TLR7-mediated immune pathogenesis. *Proc. Natl Acad. Sci. USA* **116**, 23653–23661 (2019).
114. Kim, S. et al. Identification of a novel toll-like receptor 7 endogenous ligand in rheumatoid arthritis synovial fluid that can provoke arthritic joint inflammation. *Arthritis Rheumatol.* **68**, 1099–1110 (2016).
115. Hegewald, A. B. et al. Extracellular miR-574-5p induces osteoclast differentiation via TLR 7/8 in rheumatoid arthritis. *Front. Immunol.* **11**, 585282 (2020).
116. Kim, K.-W. et al. Toll-like receptor 7 regulates osteoclastogenesis in rheumatoid arthritis. *J. Biochem.* **166**, 259–270 (2019).
117. Schneider, A. H. et al. Neutrophil extracellular traps mediate joint hyperalgesia induced by immune inflammation. *Rheumatology* **60**, 3461–3473 (2021).
118. Ye, H. et al. Internalisation of neutrophils extracellular traps by macrophages aggravate rheumatoid arthritis via Rab5a. *RMD Open* **10**, e003847 (2024).
119. Chen, F. et al. Xiaowugui decoction alleviates experimental rheumatoid arthritis by suppressing Rab5a-mediated TLR4 internalization in macrophages. *Phytomedicine* **132**, 155762 (2024).
120. Grommè, M. et al. Recycling MHC class I molecules and endosomal peptide loading. *Proc. Natl Acad. Sci. USA* **96**, 10326–10331 (1999).
121. Cohn, L. et al. Antigen delivery to early endosomes eliminates the superiority of human blood BDCA3⁺ dendritic cells at cross presentation. *J. Exp. Med.* **210**, 1049–1063 (2013).
122. Schmid, D., Pypaert, M. & Münz, C. Antigen-loading compartments for major histocompatibility complex class II molecules continuously receive input from autophagosomes. *Immunity* **26**, 79–92 (2007).
123. Pandey, K. N. Small peptide recognition sequence for intracellular sorting. *Curr. Opin. Biotechnol.* **21**, 611–620 (2010).
124. Chatterjee, B. et al. Internalization and endosomal degradation of receptor-bound antigens regulate the efficiency of cross presentation by human dendritic cells. *Blood* **120**, 2011–2020 (2012).
125. Lazzarino, D. A., Blier, P. & Mellman, I. The monomeric guanosine triphosphatase rab4 controls an essential step on the pathway of receptor-mediated antigen processing in B cells. *J. Exp. Med.* **188**, 1769–1774 (1998).
126. Krawczyk, M. et al. Expression of RAB4B, a protein governing endocytic recycling, is co-regulated with MHC class II genes. *Nucl. Acids Res.* **35**, 595–605 (2006).
127. Malinova, D. et al. Endophilin A2 regulates B-cell endocytosis and is required for germinal center and humoral responses. *EMBO Rep.* **22**, e51328 (2021).
128. Navid, F., Chen, L., Bowness, P. & Colbert, R. A. HLA-B27 and spondyloarthritis: at the crossroads of innate and adaptive immunity. *Nat. Rev. Rheumatol.* **21**, 77–87 (2025).
129. Tavassolian, F., Pastrello, C., Ahmed, Z., Jurisica, I. & Inman, R. D. Vesicular traffic-mediated cell-to-cell signaling at the immune synapse in ankylosing spondylitis. *Front. Immunol.* **13**, 1102405 (2022).
130. Bowness, P. HLA-B27. *Annu. Rev. Immunol.* **33**, 29–48 (2015).
131. Zhang, D. et al. RAB5C, SYNJ1, and RNF19B promote male ankylosing spondylitis by regulating immune cell infiltration. *Ann. Transl. Med.* **9**, 1011 (2021).
132. Fasano, D. et al. Alteration of endosomal trafficking is associated with early-onset parkinsonism caused by SYNJ1 mutations. *Cell Death Dis.* **9**, 385 (2018).
133. Lawrence, D. W., Shornick, L. P. & Kornbluth, J. Mice deficient in NKLAM have attenuated inflammatory cytokine production in a Sendai virus pneumonia model. *PLoS One* **14**, e0222802 (2019).
134. Tanaka, T. et al. Heat shock protein 90 targets a chaperone peptide to the static early endosome for efficient cross-presentation by human dendritic cells. *Cancer Sci.* **106**, 18–24 (2015).
135. Cormont, M., Gautier, N., Ilc, K. & Marchand-Brustel, Y. Expression of a prenylation-deficient Rab4 inhibits the GLUT4 translocation induced by active phosphatidylinositol 3-kinase and protein kinase B. *Biochem. J.* **356**, 143–149 (2001).
136. Voss, K. et al. Elevated transferrin receptor impairs T cell metabolism and function in systemic lupus erythematosus. *Sci. Immunol.* **8**, eabq0178 (2023).
137. Macintyre, A. N. et al. The glucose transporter Glut1 is selectively essential for CD4 T cell activation and effector function. *Cell Metab.* **20**, 61–72 (2014).
138. van der Windt, G. J. W. et al. Mitochondrial respiratory capacity is a critical regulator of CD8⁺ T cell memory development. *Immunity* **36**, 68–78 (2012).
139. Ho, P. C. et al. Phosphoenolpyruvate is a metabolic checkpoint of anti-tumor T cell responses. *Cell* **162**, 1217–1228 (2015).
140. Phan, A. T. et al. Constitutive glycolytic metabolism supports CD8⁺ T cell effector memory differentiation during viral infection. *Immunity* **45**, 1024–1037 (2016).

141. Banki, K. & Perl, A. Cell type-specific regulation of the pentose phosphate pathway during development and metabolic stress-driven autoimmune diseases: relevance for inflammatory liver, renal, endocrine, cardiovascular and neurobehavioral comorbidities, carcinogenesis, and aging. *Autoimmun. Rev.* **24**, 103781 (2025).
142. Lai, Z.-W. et al. N-acetylcysteine reduces disease activity by blocking mTOR in T cells of lupus patients. *Arthritis Rheum.* **64**, 2937–2946 (2012).
143. Alli, A. A. et al. Kidney tubular epithelial cell ferroptosis links glomerular injury to tubulointerstitial pathology in lupus nephritis. *Clin. Immunol.* **248**, 109213 (2023).
144. Condon, K. J. & Sabatini, D. M. Nutrient regulation of mTORC1 at a glance. *J. Cell Sci.* **132**, jcs222570 (2019).
145. Shibutani, S., Okazaki, H. & Iwata, H. Dynamin-dependent amino acid endocytosis activates mechanistic target of rapamycin complex 1 (mTORC1). *J. Biol. Chem.* **292**, 18052–18061 (2017).
146. Patiño-Martínez, E. & Kaplan, M. J. Immunometabolism in systemic lupus erythematosus. *Nat. Rev. Rheumatol.* **21**, 377–395 (2025).
147. Yan, H. et al. B cell endosomal RAB7 promotes TRAF6 K63 polyubiquitination and NF-κB activation for antibody class-switching. *J. Immunol.* **204**, 1146–1157 (2020).
148. Finetti, F. et al. Specific recycling receptors are targeted to the immune synapse by the intralagellar transport system. *J. Cell. Sci.* **127**, 1924–1937 (2014).
149. Compeer, E. B. et al. A mobile endocytic network connects clathrin-independent receptor endocytosis to recycling and promotes T cell activation. *Nat. Commun.* **9**, 1597 (2018).
150. Iseka, F. M. et al. Role of the EHD family of endocytic recycling regulators for TCR recycling and T cell function. *J. Immunol.* **200**, 483–499 (2018).
151. Onnis, A. & Baldari, C. T. Orchestration of immunological synapse assembly by vesicular trafficking. *Front. Cell Dev. Biol.* **7**, 110 (2019).
152. Alarcón, B., Mestre, D. & Martínez-Martín, N. The immunological synapse: a cause or consequence of T-cell receptor triggering? *Immunology* **133**, 420–425 (2011).
153. van Der Sluijs, P. et al. The small GTP-binding protein rab4 is associated with early endosomes. *Proc. Natl Acad. Sci. USA* **88**, 6313–6317 (1991).
154. Batista, A., Millan, J., Mittelbrunn, M., Sanchez-Madrid, F. & Alonso, M. A. Recruitment of transferrin receptor to immunological synapse in response to TCR engagement. *J. Immunol.* **172**, 6709–6714 (2004).
155. Wing, K. et al. CTLA-4 control over Foxp3⁺ regulatory T cell function. *Science* **322**, 271–275 (2008).
156. Janman, D. et al. Regulation of CTLA-4 recycling by LRBA and Rab11. *Immunology* **164**, 106–119 (2021).
157. Gilleron, J. et al. Rab4b deficiency in T cells promotes adipose Treg/Th17 imbalance, adipose tissue dysfunction, and insulin resistance. *Cell Rep.* **25**, 3329–3341.e5 (2018).
158. Fu, R. et al. Interleukin-23 receptor deficiency in podocytes averts the development of lupus nephritis. *Arthritis Rheumatol.* <https://doi.org/10.1002/art.43395> (2025).
159. Bagavant, H. & Fu, S. M. Pathogenesis of kidney disease in systemic lupus erythematosus. *Curr. Opin. Rheumatol.* **21**, 489–494 (2009).
160. Miao, M. Z. et al. Redox-active endosomes mediate α5β1 integrin signaling and promote chondrocyte matrix metalloproteinase production in osteoarthritis. *Sci. Signal.* **16**, eadf8299 (2023).
161. Chen, Y.-G. Endocytic regulation of TGF-β signaling. *Cell Res.* **19**, 58–70 (2009).
162. Tian, X., Bunda, P. & Ishibe, S. Podocyte endocytosis in regulating the glomerular filtration barrier. *Front. Med.* **9**, 801837 (2022).
163. Pullmann, R. Jr., Bonilla, E., Phillips, P. E., Middleton, F. A. & Perl, A. Haplotypes of the HRES-1 endogenous retrovirus are associated with development and disease manifestations of systemic lupus erythematosus. *Arthritis Rheum.* **58**, 532–540 (2008).
164. Magistrelli, C. et al. Polymorphic genotypes of the HRES-1 human endogenous retrovirus locus correlate with systemic lupus erythematosus and autoreactivity. *Immunogenetics* **49**, 829–834 (1999).
165. Rasmussen, H. B. et al. Expression of endogenous retroviruses in blood mononuclear cells and brain tissue from multiple sclerosis patients. *Mult. Scler.* **1**, 82–87 (1995).
166. Rasmussen, H. B., Kelly, M. A., Francis, D. A. & Clausen, J. Association between the endogenous retrovirus HRES-1 and multiple sclerosis in the United Kingdom—evidence of genetically different disease subsets? *Dis. Markers* **16**, 101–104 (2000).
167. Rasmussen, H. B., Kelly, M. A., Francis, D. A. & Clausen, J. Haplotypes of the endogenous retrovirus HRES-1 in multiple sclerosis patients and healthy control subjects of Shanghai Chinese origin. *Dis. Markers* **13**, 251–255 (1998).
168. Godavarthy, A. et al. Lupus-associated endogenous retroviral LTR polymorphism and epigenetic imprinting promote HRES-1/Rab4 expression and mTOR activation. *JCI Insight* **5**, e134010 (2020).
169. Perl, A. & Banki, K. Human endogenous retroviral elements and autoimmunity: data and concepts. *Trends Microbiol.* **1**, 153–156 (1993).
170. Perl, A. Editorial: LINEing up to boost interferon production: activation of endogenous retroviral DNA in autoimmunity. *Arthritis Rheumatol.* **68**, 2568–2570 (2016).
171. Chen, P.-M. et al. CD38 reduces mitochondrial fitness and cytotoxic T cell response against viral infection in lupus patients by suppressing mitophagy. *Sci. Adv.* **8**, eabo4271 (2022).
172. Kato, H. & Perl, A. The IL-21-mTOR axis blocks Treg differentiation and function by suppression of autophagy in patients with systemic lupus erythematosus. *Arthritis Rheumatol.* **70**, 427–438 (2018).
173. Zhou, X. et al. Genetic association of PRDM1-ATG5 intergenic region and autophagy with systemic lupus erythematosus in a Chinese population. *Ann. Rheum. Dis.* **70**, 1330–1337 (2011).
174. Cadwell, K. Crosstalk between autophagy and inflammatory signalling pathways: balancing defence and homeostasis. *Nat. Rev. Immunol.* **16**, 661–675 (2016).
175. Monteith, A. J. et al. Defects in lysosomal maturation facilitate the activation of innate sensors in systemic lupus erythematosus. *Proc. Natl Acad. Sci. USA* **113**, E2142–E2151 (2016).
176. Monteith, A. J. et al. mTORC2 activity disrupts lysosome acidification in systemic lupus erythematosus by impairing caspase-1 cleavage of Rab39a. *J. Immunol.* **201**, 371–382 (2018).
177. Brown, J. et al. Microbiota-mediated skewing of tryptophan catabolism modulates CD4⁺ T cells in lupus-prone mice. *iScience* **25**, 104241 (2022).
178. Choi, S. C. et al. Gut microbiota dysbiosis and altered tryptophan catabolism contribute to autoimmunity in lupus-susceptible mice. *Sci. Transl. Med.* **12**, eaax2220 (2020).
179. Perl, A. et al. Comprehensive metabolome analyses reveal N-acetylcysteine-responsive accumulation of kynurenine in systemic lupus erythematosus: implications for activation of the mechanistic target of rapamycin. *Metabolomics* **11**, 1157–1174 (2015).
180. Huang, N. & Perl, A. Metabolism as a target for modulation in autoimmune diseases. *Trends Immunol.* **39**, 562–576 (2018).
181. Perl, A. Mechanistic target of rapamycin pathway activation in rheumatic diseases. *Nat. Rev. Rheumatol.* **12**, 169–182 (2016).
182. Zhang, Y. & Manning, B. D. mTORC1 signaling activates NRF1 to increase cellular proteasome levels. *Cell Cycle* **14**, 2011–2017 (2015).
183. Sancak, Y. et al. The Rag GTPases bind raptor and mediate amino acid signaling to mTORC1. *Science* **320**, 1496–1501 (2008).
184. Bridges, D. et al. Rab5 proteins regulate activation and localization of target of rapamycin complex 1. *J. Biol. Chem.* **287**, 20913–20921 (2012).
185. Lai, Z. et al. Sirolimus in patients with clinically active systemic lupus erythematosus resistant to, or intolerant of, conventional medications: a single-arm, open-label, phase 1/2 trial. *Lancet* **391**, 1186–1196 (2018).
186. Park, J. S. et al. RAB4A drives proinflammatory CD4⁺ T cell signaling via CD38-dependent NAD⁺ metabolism. Preprint at Res. Sq. <https://doi.org/10.21203/rs.3.rs-6787101/v1> (2025).
187. Fang, C. et al. CD38 produces nicotinic acid adenosine dinucleotide phosphate in the lysosome. *J. Biol. Chem.* **293**, 8151–8160 (2018).
188. Roccatello, D. et al. Daratumumab monotherapy for refractory lupus nephritis. *Nat. Med.* **29**, 2041–2047 (2023).
189. Gibbins, D. J., Marcet-Palacios, M., Sekar, Y., Ng, M. C. Y. & Befus, A. D. CD8α is expressed by human monocytes and enhances FcγR-dependent responses. *BMC Immunol.* **8**, 12 (2007).
190. Arcaro, A. et al. Essential role of CD8 palmitoylation in CD8 coreceptor function. *J. Immunol.* **165**, 2068–2076 (2000).
191. Gorska, M. M., Liang, Q., Karim, Z. & Alam, R. Uncoordinated 119 protein controls trafficking of Lck via the Rab11 endosome and is critical for immunological synapse formation. *J. Immunol.* **183**, 1675–1684 (2009).
192. Franciszkiwicz, K. et al. Synaptic release of CCL5 storage vesicles triggers CXCR4 surface expression promoting CTL migration in response to CXCL12. *J. Immunol.* **193**, 4952–4961 (2014).
193. Yu, H. et al. The chromatin accessibility landscape of peripheral blood mononuclear cells in patients with systemic lupus erythematosus at single-cell resolution. *Front. Immunol.* **12**, 641886 (2021).
194. Crispin, J. C. et al. Expanded double negative T cells in patients with systemic lupus erythematosus produce IL-17 and infiltrate the kidneys. *J. Immunol.* **181**, 8761–8766 (2008).
195. Berthier, C. C., Kretzler, M. & Davidson, A. A systems approach to renal inflammation in SLE. *Clin. Immunol.* **185**, 109–118 (2017).
196. Umbarino, D. Lupus nephritis: NLRP3 inflammasome ignites podocyte dysfunction. *Nat. Rev. Rheumatol.* **13**, 451 (2017).
197. Caza, T., Wijewardena, C., Al-Rabadi, L. & Perl, A. Cell type-specific mechanistic target of rapamycin-dependent distortion of autophagy pathways in lupus nephritis. *Transl. Res.* **245**, 55–81 (2022).
198. Soda, K. et al. Role of dynamin, synaptotagmin, and endophilin in podocyte foot processes. *J. Clin. Invest.* **122**, 4401–4411 (2012).
199. Bechtel, W. et al. Vps34 deficiency reveals the importance of endocytosis for podocyte homeostasis. *J. Am. Soc. Nephrol.* **24**, 727–743 (2013).
200. Hubbard, J. J. et al. FcRn is a CD32a coreceptor that determines susceptibility to IgG immune complex-driven autoimmunity. *J. Exp. Med.* **217**, e20200359 (2020).
201. Olaru, F. et al. Neonatal Fc receptor promotes immune complex-mediated glomerular disease. *J. Am. Soc. Nephrol.* **25**, 918–925 (2014).
202. Shih, N. Y. et al. Congenital nephrotic syndrome in mice lacking CD2-associated protein. *Science* **286**, 312–315 (1999).
203. Ward, E. S. et al. From sorting endosomes to exocytosis: association of Rab4 and Rab11 GTPases with the Fc receptor, FcRn, during recycling. *Mol. Biol. Cell* **16**, 2028–2038 (2005).
204. Welsch, T. et al. Association of CD2AP with dynamic actin on vesicles in podocytes. *Am. J. Physiol. Ren. Physiol.* **289**, F1134–F1143 (2005).
205. Wilder, R. L. Integrin αVβ3 as a target for treatment of rheumatoid arthritis and related rheumatic diseases. *Ann. Rheum. Dis.* **61**, ii96–ii99 (2002).
206. Nam, E. J. et al. Up-regulated transforming growth factor β-inducible gene h3 in rheumatoid arthritis mediates adhesion and migration of synovial cells through αvβ3 integrin: Regulation by cytokines. *Arthritis Rheum.* **54**, 2734–2744 (2006).

207. Charbonneau, M. et al. Platelet-derived growth factor receptor activation promotes the prodestructive invadosome-forming phenotype of synoviocytes from patients with rheumatoid arthritis. *J. Immunol.* **196**, 3264–3275 (2016).
208. Matsumura, T. et al. Phosphorylated platelet-derived growth factor receptor-positive cells with anti-apoptotic properties accumulate in the synovium of patients with rheumatoid arthritis. *Front. Immunol.* **10**, 241 (2019).
209. Roberts, M., Barry, S., Woods, A., van der Sluijs, P. & Norman, J. PDGF-regulated rab4-dependent recycling of alphavbeta3 integrin from early endosomes is necessary for cell adhesion and spreading. *Curr. Biol.* **11**, 1392–1402 (2001).
210. Vukmirica, J., Monzo, P., Le Marchand-Brustel, Y. & Cormont, M. The Rab4A effector protein Rabip4 is involved in migration of NIH 3T3 fibroblasts. *J. Biol. Chem.* **281**, 36360–36368 (2006).
211. Liao, W. et al. CARP-2 is an endosome-associated ubiquitin ligase for RIP and regulates TNF-induced NF- κ B activation. *Curr. Biol.* **18**, 641–649 (2008).
212. Moskovitch, O. et al. The potential of autologous patient-derived circulating extracellular vesicles to improve drug delivery in rheumatoid arthritis. *Clin. Exp. Immunol.* **219**, uxae101 (2025).
213. Berckmans, R. J. et al. Cell-derived microparticles in synovial fluid from inflamed arthritic joints support coagulation exclusively via a factor VII-dependent mechanism. *Arthritis Rheum.* **46**, 2857–2866 (2002).
214. Malda, J., Boere, J., van de Lest, C. H. A., van Weeren, P. R. & Wauben, M. H. M. Extracellular vesicles — a new tool for joint repair and regeneration. *Nat. Rev. Rheumatol.* **12**, 243–249 (2016).
215. Wieman, H. L., Wofford, J. A. & Rathmell, J. C. Cytokine stimulation promotes glucose uptake via phosphatidylinositol-3 Kinase/Akt regulation of GLUT1 activity and trafficking. *Mol. Biol. Cell* **18**, 1437–1446 (2007).
216. Pei, Y. et al. RAB21 controls autophagy and cellular energy homeostasis by regulating retromer-mediated recycling of SLC2A1/GLUT1. *Autophagy* **19**, 1070–1086 (2023).
217. Mobasheri, A. et al. The role of metabolism in the pathogenesis of osteoarthritis. *Nat. Rev. Rheumatol.* **13**, 302–311 (2017).
218. Schwartz, M. A. Integrins and extracellular matrix in mechanotransduction. *Cold Spring Harb. Perspect. Biol.* **2**, a005066 (2010).
219. Janssen, J. N. et al. Enhancing the chondrogenic potential of chondrogenic progenitor cells by deleting RAB5C. *iScience* **24**, 102464 (2021).
220. Huang, H. et al. Identification of pathways and genes associated with synovitis in osteoarthritis using bioinformatics analyses. *Sci. Rep.* **8**, 10050 (2018).
221. Diaz-Romero, J. et al. Immunophenotypic analysis of human articular chondrocytes: changes in surface markers associated with cell expansion in monolayer culture. *J. Cell Physiol.* **202**, 731–742 (2005).
222. Wang, W. et al. TFR1 mediated iron metabolism dysfunction as a potential therapeutic target for osteoarthritis. *Arthritis Res. Ther.* **26**, 71 (2024).
223. Bale, S., Verma, P., Varga, J. & Bhattacharyya, S. Extracellular matrix-derived damage-associated molecular patterns (DAMP): implications in systemic sclerosis and fibrosis. *J. Investig. Dermatol.* **143**, 1877–1885 (2023).
224. Di Guglielmo, G. M., Le Roy, C., Goodfellow, A. F. & Wrana, J. L. Distinct endocytic pathways regulate TGF- β receptor signalling and turnover. *Nat. Cell Biol.* **5**, 410–421 (2003).
225. Yin, X., Murphy, S. J., Wilkes, M. C., Ji, Y. & Leof, E. B. Retromer maintains basolateral distribution of the type II TGF- β receptor via the recycling endosome. *Mol. Biol. Cell* **24**, 2285–2298 (2013).
226. Mitchell, H., Choudhury, A., Pagano, R. E. & Leof, E. B. Ligand-dependent and -independent transforming growth factor- β receptor recycling regulated by clathrin-mediated endocytosis and Rab11. *Mol. Biol. Cell* **15**, 4166–4178 (2004).
227. He, K. et al. Internalization of the TGF- β type I receptor into caveolin-1 and EEA1 double-positive early endosomes. *Cell Res.* **25**, 738–752 (2015).
228. Strippoli, R. et al. Caveolin-1 deficiency induces a MEK-ERK1/2-Snail-1-dependent epithelial-mesenchymal transition and fibrosis during peritoneal dialysis. *EMBO Mol. Med.* **7**, 102–123 (2015).
229. Reese, C. et al. Caveolin-1 deficiency may predispose African Americans to systemic sclerosis-related interstitial lung disease. *Arthritis Rheumatol.* **66**, 1909–1919 (2014).
230. Paradowska-Gorycka, A. et al. Variety of endosomal TLRs and Interferons (IFN- α , IFN- β , IFN- γ) expression profiles in patients with SLE, SSC and MCTD. *Clin. Exp. Immunol.* **204**, 49–63 (2021).
231. Tanaka, T. et al. LAMP3 induces apoptosis and autoantigen release in Sjögren's syndrome patients. *Sci. Rep.* **10**, 15169 (2020).
232. Nakamura, H. et al. Lysosome-associated membrane protein 3 misexpression in salivary glands induces a Sjögren's syndrome-like phenotype in mice. *Ann. Rheum. Dis.* **80**, 1031–1039 (2021).
233. Meng, Z. et al. Imbalanced Rab3D versus Rab27 increases cathepsin S secretion from lacrimal acini in a mouse model of Sjögren's syndrome. *Am. J. Physiol. Cell Physiol.* **310**, C942–C954 (2016).
234. Zhao, K., Sun, Y., Zhong, S. & Luo, J.-L. The multifaceted roles of cathepsins in immune and inflammatory responses: implications for cancer therapy, autoimmune diseases, and infectious diseases. *Biomark. Res.* **12**, 165 (2024).
235. Bentley, D., Fisher, B. A., Barone, F., Kolb, F. A. & Attley, G. A randomized, double-blind, placebo-controlled, parallel group study on the effects of a cathepsin S inhibitor in primary Sjögren's syndrome. *Rheumatology* **62**, 3644–3653 (2023).
236. Ménasché, G. et al. Mutations in RAB27A cause Griscelli syndrome associated with haemophagocytic syndrome. *Nat. Genet.* **25**, 173–176 (2000).
237. Wang, T.-T., Li, Z.-Z., Cai, Y., Ren, J.-G. & Zhao, J.-H. Emerging roles of Rab27 proteins: from normal to cancer stem cells. *Biochem. Biophys. Res. Commun.* **775**, 152109 (2025).
238. Yokoyama, K. et al. Rab27a negatively regulates phagocytosis by prolongation of the actin-coating stage around phagosomes. *J. Biol. Chem.* **286**, 5375–5382 (2011).
239. Schuler, G. S. & Cron, R. Q. The genetics of macrophage activation syndrome. *Genes Immun.* **21**, 169–181 (2020).
240. Zhang, M. et al. A Heterozygous RAB27A mutation associated with delayed cytolytic granule polarization and hemophagocytic lymphohistiocytosis. *J. Immunol.* **196**, 2492–2503 (2016).
241. Carter, S. J., Tattersall, R. S. & Ramanan, A. V. Macrophage activation syndrome in adults: recent advances in pathophysiology, diagnosis and treatment. *Rheumatology* **58**, 5–17 (2019).
242. Rennick, J. J., Nowell, C. J., Pouton, C. W. & Johnston, A. P. R. Resolving subcellular pH with a quantitative fluorescent lifetime biosensor. *Nat. Commun.* **13**, 6023 (2022).
243. Cenac, C., Ducatez, M. F. & Guéry, J.-C. Hydroxychloroquine inhibits proteolytic processing of endogenous TLR7 protein in human primary plasmacytoid dendritic cells. *Eur. J. Immunol.* **52**, 54–61 (2022).
244. Sacre, K., Criswell, L. A. & McCune, J. M. Hydroxychloroquine is associated with impaired interferon-alpha and tumor necrosis factor-alpha production by plasmacytoid dendritic cells in systemic lupus erythematosus. *Arthritis Res. Ther.* **14**, R155 (2012).
245. Webster, H. UNC93B1 variants promote SLE via TLR activation. *Nat. Rev. Rheumatol.* **20**, 528 (2024).
246. Alghazali, R., Tabebi, M., Elmasry, M. & El-Serafi, M. A. Rab7 inhibitor enhances stem cell differentiation into keratinocyte-like cells with anti-inflammatory properties. *Front. Immunol.* **16**, 1503007 (2025).
247. Tanaka, Y. et al. Safety, pharmacokinetics, biomarker response and efficacy of E6742: a dual antagonist of Toll-like receptors 7 and 8, in a first in patient, randomised, double-blind, phase I/II study in systemic lupus erythematosus. *RMD Open* **10**, e004701 (2024).
248. Hawtin, S. et al. Preclinical characterization of the Toll-like receptor 7/8 antagonist MHV370 for lupus therapy. *Cell Rep. Med.* **4**, 101036 (2023).
249. Shisha, T. et al. First-in-human study of the safety, pharmacokinetics, and pharmacodynamics of MHV370, a dual inhibitor of Toll-like receptors 7 and 8, in healthy adults. *Eur. J. Drug Metab. Pharmacokinet.* **48**, 553–566 (2023).
250. Vlach, J. et al. Discovery of M5049: a novel selective toll-like receptor 7/8 inhibitor for treatment of autoimmunity. *J. Pharmacol. Exp. Ther.* **376**, 397–409 (2021).
251. Ogama, Y. et al. Phase 1 clinical trial of PPMX-T003, a novel human monoclonal antibody specific for transferrin receptor 1, to evaluate its safety, Pharmacokinetics, and Pharmacodynamics. *Clin. Pharmacol. Drug Dev.* **12**, 579–587 (2023).
252. Zhang, J. et al. Structure-based discovery of neoandrographolide as a novel inhibitor of Rab5 to suppress cancer growth. *Comput. Struct. Biotechnol. J.* **18**, 3936–3946 (2020).
253. Liu, J., Wang, Z.-T., Ji, L.-L. & Ge, B.-X. Inhibitory effects of neoandrographolide on nitric oxide and prostaglandin E2 production in LPS-stimulated murine macrophage. *Mol. Cell Biochem.* **298**, 49–57 (2007).
254. Tang, K. et al. Neoandrographolide inhibits mature osteoclast differentiation to alleviate bone loss and treat osteoporosis. *Front. Pharmacol.* **16**, 1466057 (2025).
255. Kim, J. H., Lee, C.-H. & Baek, M.-C. Dissecting exosome inhibitors: therapeutic insights into small-molecule chemicals against cancer. *Exp. Mol. Med.* **54**, 1833–1843 (2022).
256. Yan, D. & Wan, L. WTAP modulates macrophage polarization in rheumatoid arthritis by targeting exosomal circ-CBLB via m6A modification. *Front. Immunol.* **16**, 1601259 (2025).
257. Cao, Y. et al. Extracellular vesicles in infrapatellar fat pad from osteoarthritis patients impair cartilage metabolism and induce senescence. *Adv. Sci.* **11**, e2303614 (2024).
258. Dai, J. et al. Osteoclast-derived exosomal miR-212-3p suppressed the anabolism and accelerated the catabolism of chondrocytes in osteoarthritis by targeting TGF- β 1/Smad2 signaling. *Arch. Biochem. Biophys.* **751**, 109827 (2024).
259. Essandoh, K. et al. Blockade of exosome generation with GW4869 dampens the sepsis-induced inflammation and cardiac dysfunction. *Biochim. Biophys. Acta* **1852**, 2362–2371 (2015).
260. Xia, M. et al. Transcription factor EB coordinates environmental cues to regulate T regulatory cells' mitochondrial fitness and function. *Proc. Natl Acad. Sci. USA* **119**, e2205469119 (2022).
261. Piranavan, P. & Perl, A. Improvement of renal and non-renal SLE outcome measures on sirolimus therapy - A 21-year follow-up study of 73 patients. *Clin. Immunol.* **229**, 108781 (2021).
262. Mao, Z. et al. Renal mTORC1 activation is associated with disease activity and prognosis in lupus nephritis. *Rheumatology* **61**, 3830–3840 (2022).
263. Jiang, N. et al. Sirolimus versus tacrolimus for systemic lupus erythematosus treatment: results from a real-world CSTAR cohort study. *Lupus Sci. Med.* **9**, e000617 (2022).
264. Lin, Y. et al. A small-molecule drug inhibits autophagy gene expression through the central regulator TFEB. *Proc. Natl Acad. Sci. USA* **120**, e2213670120 (2023).
265. Scheinberg, P., Singulane, C. C., Barbosa, L. S. G. & Scheinberg, M. Successful platelet count recovery in lupus-associated thrombocytopenia with the thrombopoietin agonist eltrombopag. *Clin. Rheumatol.* **33**, 1347–1349 (2014).
266. Li, W. et al. Eltrombopag in severe immune thrombocytopenia secondary to connective tissue disease: a report of 17 patients and literature review. *Clin. Rheumatol.* **42**, 1451–1457 (2023).

267. Garra, W., Carmi, O., Kivity, S. & Levy, Y. Catastrophic antiphospholipid syndrome in lupus-associated immune thrombocytopenia treated with eltrombopag A case series and literature review. *Medicine* **102**, e32949 (2023).
268. Schrezenmeier, E. & Dörner, T. Mechanisms of action of hydroxychloroquine and chloroquine: implications for rheumatology. *Nat. Rev. Rheumatol.* **16**, 155–166 (2020).
269. Zhang, L., Cui, J. Y. & Zhang, L. Clinical efficacy and safety of sirolimus in childhood-onset systemic lupus erythematosus in real world. *Medicine* **101**, e31551 (2022).
270. Yap, D. Y. H. et al. Longterm data on sirolimus treatment in patients with lupus nephritis. *J. Rheumatol.* **45**, 1663–1670 (2018).
271. Sevim, E. et al. Mammalian target of rapamycin pathway assessment in antiphospholipid antibody-positive patients with livedo. *J. Rheumatol.* **49**, 1026–1030 (2022).
272. Eriksson, P., Wallin, P. & Sjöwall, C. Clinical experience of sirolimus regarding efficacy and safety in systemic lupus erythematosus. *Front. Pharmacol.* **10**, 82 (2019).
273. Hayes, G. M. et al. Antitumor activity of an anti-CD98 antibody. *Int. J. Cancer* **137**, 710–720 (2015).
274. Qureshi, O. S. et al. Constitutive clathrin-mediated endocytosis of CTLA-4 persists during T cell activation. *J. Biol. Chem.* **287**, 9429–9440 (2012).
275. Donaldson, J. G., Johnson, D. L. & Dutta, D. Rab and Arf G proteins in endosomal trafficking and cell surface homeostasis. *Small GTPases* **7**, 247–251 (2016).
276. Feng, S. et al. The mechanism of ferroptosis and its related diseases. *Mol. Biomed.* **4**, 33 (2023).
277. Kato, H. & Perl, A. Mechanistic target of rapamycin complex 1 expands Th17 and IL-4⁺ CD4-CD8- double-negative T cells and contracts regulatory T cells in systemic lupus erythematosus. *J. Immunol.* **192**, 4134–4144 (2014).
278. Kato, H. & Perl, A. Blockade of treg cell differentiation and function by the interleukin-21-mechanistic target of rapamycin axis via suppression of autophagy in patients with systemic lupus erythematosus. *Arthritis Rheumatol.* **70**, 427–438 (2018).
279. Zhou, X. et al. mTORC2 contributes to systemic autoimmunity. *Immunology* **168**, 554–568 (2023).
280. Hailey, D. W. et al. Mitochondria supply membranes for autophagosome biogenesis during starvation. *Cell* **141**, 656–667 (2010).
281. Mandal, T. et al. Mechano-regulation by clathrin pit-formation and passive cholesterol-dependent tubules during de-adhesion. *Cell Mol. Life Sci.* **81**, 43 (2024).
282. Srivatsan, S., Swiecki, M., Otero, K., Cella, M. & Shaw, A. S. CD2-associated protein regulates plasmacytoid dendritic cell migration, but is dispensable for their development and cytokine production. *J. Immunol.* **191**, 5933–5940 (2013).
283. Blasius, A. L. et al. Slc15a4, AP-3, and Hermansky-Pudlak syndrome proteins are required for Toll-like receptor signaling in plasmacytoid dendritic cells. *Proc. Natl Acad. Sci. USA* **107**, 19973–19978 (2010).
284. Moffett, J. R. et al. Quinolate as a marker for kynurenine metabolite formation and the unresolved question of NAD⁺ synthesis during inflammation and infection. *Front. Immunol.* **11**, 31 (2020).
285. Zhao, Y. J., Lam, C. M. C. & Lee, H. C. The membrane-bound enzyme CD38 exists in two opposing orientations. *Sci. Signal.* **5**, ra67 (2012).
286. Xie, N. et al. NAD⁺ metabolism: pathophysiologic mechanisms and therapeutic potential. *Signal Transduct. Target. Ther.* **5**, 227 (2020).
287. Lane, D. J. R. et al. Cellular iron uptake, trafficking and metabolism: key molecules and mechanisms and their roles in disease. *Bioch. Biophys. Acta Mol. Cell Res.* **1853**, 1130–1144 (2015).

Acknowledgements

The work of the author has been supported in part by grants A1072648 and A1122176 from the National Institutes of Health, the Phillips Lupus and Autoimmunity Centre of Excellence, and the Central New York Community Foundation.

Competing interests

The author declares no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41584-025-01336-4>.

Peer review information *Nature Reviews Rheumatology* thanks Hu Zeng, who co-reviewed with Xingxing Zhu, and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

© Springer Nature Limited 2025

Inflammation and pain as interconnected targets in axial spondyloarthritis

Xenofon Baraliakos¹✉, Victoria Navarro-Compán², Nelly Ziade³ & Denis Poddubnyy^{4,5}

Abstract

Axial spondyloarthritis (axSpA) is a chronic inflammatory disease characterized by complex pain mechanisms that extend beyond inflammation. Although inflammatory nociceptive pain – primarily mediated by pro-inflammatory cytokines – represents the classic pathway and therapeutic target, many patients continue to experience pain despite suppression of inflammation. This residual pain often reflects non-inflammatory processes, including nociplastic and neuropathic pain. Central sensitization, a key mechanism of nociplastic pain, contributes to pain amplification and poor response to treatment. Fibromyalgia, considered the typical phenotype of nociplastic pain, can co-occur with axSpA and is associated with increased symptom burden and reduced efficacy of anti-inflammatory therapies. Neuropathic pain, albeit less common, can result from structural complications and requires targeted therapeutic approaches. In addition, biological sex differences further influence pain perception and treatment outcomes: female patients report more widespread pain, show higher rates of central sensitization and have a worse response to biologic therapies than male patients. Current treatment paradigms are effective for inflammation-driven symptoms but often fail to address the broader spectrum of pain phenotypes in axSpA. Future work should include the development of biomarkers to differentiate pain mechanisms, the refinement of assessment tools and the evaluation of multimodal therapies that target both inflammation and pain processes. This evolving understanding necessitates a shift from an inflammation-centric to a mechanism-informed approach to pain management in axSpA.

Sections

Introduction

Characteristics of pain in axial spondyloarthritis

Drivers of pain in axial spondyloarthritis

Assessment of pain in axial spondyloarthritis

Interconnection between the different types of pain in axial spondyloarthritis

Treatment options for pain in axial spondyloarthritis

Future directions and research needs

Conclusion

¹Rheumazentrum Ruhrgebiet Herne, Ruhr Universität Bochum, Bochum, Germany. ²Department of Rheumatology, La Paz University Hospital, IdiPaz, Madrid, Spain. ³Hotel-Dieu de France Hospital, Saint-Joseph University, Beirut, Lebanon. ⁴Division of Rheumatology, Department of Medicine, University of Toronto and Schroeder Arthritis Institute, University Health Network, Toronto, ON, Canada. ⁵Department of Gastroenterology, Infectiology and Rheumatology (including Nutrition Medicine), Charité — Universitätsmedizin Berlin, Berlin, Germany.

✉e-mail: baraliakos@me.de

Key points

- Pain in axial spondyloarthritis (axSpA) is multifactorial. Nociceptive, nociplastic and neuropathic components frequently coexist, and central sensitization is present in about 45–60% of patients, amplifying symptoms beyond their inflammatory origin.
- Fibromyalgia and widespread pain occur in up to one-quarter of patients with axSpA, confound disease activity scores and are linked to worse response to biologic therapies and overall disease burden.
- Sex differences are clinically relevant; women more often present with widespread and thoracic pain, higher central sensitization and worse treatment response, whereas men develop more structural damage but report lower disease activity.
- Cytokines such as TNF and IL-17 drive inflammatory nociceptive pain but also contribute to central and peripheral neuroinflammation, which links inflammation with neuropathic and nociplastic pain mechanisms in axSpA.

Introduction

Axial spondyloarthritis (axSpA) is a chronic inflammatory rheumatic disease that primarily affects the axial skeleton, including the sacroiliac joints and spine. AxSpA includes non-radiographic axSpA, which is defined by the absence of definitive sacroiliac joint changes on conventional radiographs, and radiographic axSpA (formerly referred to as ankylosing spondylitis), for which there is evidence of definite radiographic damage¹. The hallmark symptom of axSpA is chronic back pain, which often presents before 40 years of age. In axSpA, back pain is frequently described as ‘inflammatory back pain’ (IBP) (Box 1). IBP refers to a cluster of clinical features that raise suspicion of underlying inflammatory disease. Three sets of IBP criteria – the Calin², Berlin³ and Assessment of Spondyloarthritis International Society (ASAS)⁴ criteria – all rely on the characteristic features of IBP; however, the presence of IBP alone does not indicate the presence of underlying rheumatic inflammatory conditions.

Historically, pain in axSpA was viewed almost exclusively through a unidimensional lens, as a direct consequence of inflammation. Although other mechanisms were known, they were largely neglected in clinical practice and research. The advent of highly effective anti-inflammatory therapies shows that despite well-controlled inflammation, many patients continue to experience substantial symptoms. This residual pain often reflects mechanisms beyond inflammation, such as structural damage, altered central pain processing or neuropathic factors⁵. The recognition of these alternative pain mechanisms has prompted a paradigm shift towards a more nuanced, multidimensional understanding of pain in axSpA.

This Review is aimed at providing a comprehensive overview of the different mechanisms that contribute to pain in axSpA and highlights the clinical implications of this broader perspective. By promoting a multidimensional view of pain, we aim to refine disease assessment and management strategies, moving beyond an inflammation-centric model to improve patient outcomes.

Characteristics of pain in axial spondyloarthritis

Following axSpA diagnosis and the initiation of anti-inflammatory therapy, patients might continue to report pain. Pain in axSpA arises

from multiple, often overlapping, mechanisms that include all types of pain: nociceptive pain, nociplastic pain and neuropathic pain (Table 1).

Nociceptive pain

Traditionally, axSpA pain was attributed solely to inflammation of the sacroiliac joints, spine or peripheral musculoskeletal structures (arthritis, enthesitis and dactylitis), owing to nociceptive pain⁵. This type of pain is typically localized, deep and aching, and has inflammatory characteristics in most patients (Box 1). Nociceptive pain is mediated by peripheral nociceptors activated by tissue injury or inflammation^{6,7}. Inflammatory nociceptive pain responds to anti-inflammatory agents, including NSAIDs, biologic or targeted synthetic DMARDs (bDMARDs and tsDMARDs). However, nociceptive pain can also arise from non-inflammatory sources such as structural damage, concomitant degenerative or mechanical changes or muscle strain^{5,8}.

In axSpA, inflammation drives structural changes over time⁹, including new bone formation leading to syndesmophytes and ankylosis, and bone loss as a result of osteoporosis. These processes alter spinal biomechanics and, at advanced stages, can result in a fully ankylosed spine. Interestingly, in some patients, advanced ankylosis itself might not be directly associated with pain, as inflammatory activity diminishes and movement in the affected segments becomes restricted; however, structural damage can indirectly lead to pain through several mechanisms. First, as motion is lost in ankylosed segments, increased mechanical stress is transferred to the remaining mobile areas of the spine, potentially causing persistent nociceptive pain. Furthermore, secondary osteoarthritis might develop in adjacent mobile spinal segments as a long-term consequence of altered biomechanics and chronic mechanical overload. Imaging has a key role in distinguishing pain related to active inflammation from pain caused by mechanical complications secondary to structural damage¹⁰.

Overall, new bone formation in the sacroiliac joints and spine – the primary mechanism underlying structural damage progression in axSpA – does not necessarily translate into increased pain. In analyses of two long-term extension studies of TNF inhibitor trials, patient-reported outcomes, including pain, remained stable despite ongoing progression of structural damage¹¹.

Nociplastic pain

Nociplastic pain arises from altered nociception, typically in the absence of ongoing tissue or nerve injury^{7,12}. Central sensitization, whereby spinal and supraspinal pain pathways amplify input, is considered the core mechanism of nociplastic pain. Signs of central sensitization occur in 45%–60% of patients with axSpA^{13–17}. Fibromyalgia is considered the prototypical nociplastic pain syndrome and can co-occur with axSpA¹⁸.

Currently, nociplastic pain pathophysiology can be broadly divided into two subtypes: descending (top-down) and ascending (bottom-up) pathways. Individuals with top-down nociplastic pain are thought to have a primary brain-based disorder, which often manifests early in life with emerging sleep or memory problems, followed by pain beginning in one body region, then gradually spreading to others into adulthood. The genetic and familial contribution to nociplastic pain seems strongest in these individuals with top-down nociplastic pain^{19–21}.

By contrast, individuals with bottom-up nociplastic pain usually have an identifiable nociceptive or neuropathic pain condition, upon which nociplastic pain develops secondarily. In these patients, treatment is often most effective when it targets both the central sensitization process and the underlying pain condition, in this case, axSpA^{19,22}.

In axSpA, nociplastic pain can be suspected if pain persists despite controlled inflammation, especially if pain is in multiple regions of the body. The presence of fatigue, sleep disturbances, cognitive dysfunction, depression and anxiety, all of which are known to occur in axSpA, increases the probability of nociplastic pain²³. Patients with axSpA who have nociplastic pain frequently show higher scores in axSpA-related patient-reported outcomes²⁴ and worse response to NSAIDs, bDMARDs and tsDMARDs than those patients with nociceptive pain²⁴. For example, a 'possible' or 'likely' nociplastic pain component (defined as a widespread pain index (WPI) of ≥ 4) was reported in 27% of patients (22% 'possible' and 5% 'likely') with radiographic axSpA receiving bDMARDs for at least 6 months with presumably sufficient control of inflammatory activity (although control of inflammation was not verified with MRI²⁵). At the time of assessment, 21% of patients had a Bath Ankylosing Spondylitis Disease Activity Index (BASDAI) of ≥ 4 , indicating a high symptom burden. In multivariable linear regression, WPI was independently associated with the presence of residual symptoms²⁵.

Neuropathic pain

Neuropathic pain results from damage or disease of the somatosensory nervous system⁷. Clinically, neuropathic pain presents as burning, electric shock-like or shooting pain, often with paraesthesia or radicular radiation⁶. Although axSpA is not a condition that directly damages the nervous system, the prevalence of neuropathic pain in axSpA ranges between 22% and 56.9%^{26,27}, and could be caused by structural damage leading to nerve-root or spinal-cord compression, as observed in cauda equina syndrome or concurrent degenerative disease.

Overall, chronic pain in axSpA often involves overlapping phenotypes (Fig. 1), which are not mutually exclusive and might be present at the same time in the same patient⁵. Active inflammation provides 'nociceptive input' that can trigger peripheral sensitization (inflammatory mediators lower the threshold of local nociceptors, causing heightened pain sensitivity in affected tissues). If that input is prolonged, the dorsal horn neurons and pain-processing pathways in the central nervous system undergo neuroplastic changes – a process of central sensitisation – leading to persistent pain perception even after inflammation subsides, which could result in nociplastic pain^{8,23}. At the same time, chronic structural damage might cause nerve irritation (neuropathic pain) and mechanical strain (nociceptive pain), all of which contribute to the total pain burden⁸ (Fig. 2).

The clinical relevance of this complex construct is high; persistent pain in axSpA should prompt an evaluation for predominant pain mechanisms, rather than reflexively assuming inflammation that is refractory to treatment. Identifying a neuropathic or nociplastic component (including fibromyalgia) has direct treatment implications; instead of switching anti-inflammatory treatment, the rheumatologist might need to address factors associated with nociplastic pain such as anxiety, depression, sleep disturbance and modify the management strategy by addressing non-nociceptive pain mechanisms.

Drivers of pain in axial spondyloarthritis

In axSpA, pain arises from a complex interplay of factors, making it difficult to isolate the contribution of each one individually. Traditionally, nociceptive pain has been attributed to pro-inflammatory cytokines and mechanical stress; however, emerging evidence suggests that these factors might also have a role in neuropathic and nociplastic pain. Additionally, coexisting fibromyalgia and biological sex-related differences are increasingly recognized as considerable contributors to the pain experience in patients with axSpA.

Box 1 | Core clinical features of inflammatory back pain

- Insidious (gradual) onset of chronic back pain
- Morning stiffness in the low back (typically ≥ 30 min)
- Improvement with exercise or activity (and no sustained relief from rest)
- No improvement with rest (pain might even worsen with inactivity)
- Pain at night, especially in the second half of the night, often improving upon getting up

The role of cytokines

Cytokines have a central role in the pathogenesis and persistence of pain in axSpA by driving inflammation, sensitizing nociceptors and contributing to central pain-processing abnormalities (Table 1). Among these cytokines, TNF and IL-17 are key mediators in the inflammatory response. In addition, these cytokines enhance the production of prostaglandins and other pain mediators, directly activating nociceptors and increasing pain perception.

The central role of TNF was first identified in biopsy-obtained sacroiliac joint samples from patients with axSpA, in which *TNF* mRNA was abundant²⁸. TNF is recognized as a common downstream effector pathway in several immune-mediated inflammatory diseases, including axSpA. Functionally, TNF is an important activator and product of macrophages that stimulates cytokine production in immune cells and activates fibroblasts, leading to subsequent tissue remodelling^{29,30}. TNF is also produced by neutrophils and activated T cells, which are enriched in the inflamed synovial membrane and enthesal structures³¹. The success of TNF inhibitors, which can reduce both inflammation and pain in axSpA, is the ultimate proof of the pivotal role of this pro-inflammatory cytokine^{32,33}. The next successful studies were observed with IL-17 inhibitors and JAK inhibitors. Conversely, trials of drugs that inhibit IL-6, IL-23 and the IL-1 receptor have failed in axSpA³².

IL-17A constitutes a major cytokine involved in the pathogenesis of axSpA; several IL-17A-producing cell types, including T helper 17 (T_H17) cells, $\gamma\delta$ T cells and ILC3s, are present at enthesal vertebral sites. Enthesitis, a key feature of axSpA, arises from robust activation of prostaglandin E₂ (PGE₂) and the IL-23–IL-17 axis. This effect leads to the influx of innate immune cells into the entheses, followed by mesenchymal tissue responses and new bone formation^{34,35}. The sustained production of PGE₂ through cyclooxygenase-2 (COX-2) activation is associated with inflammatory responses, such as vasodilatation and neutrophil attraction to tissues, leading to spondylitis³⁶. The inhibition of IL-17 and TNF has been shown to be effective for the resolution of enthesitis in axSpA³⁶.

Notably, IL-17A and PGE₂ are important pain mediators, which are expressed in the dorsal root ganglia. In animal studies, a single injection of IL-17A into the knee joint in rats elicited a slow developing and long-lasting sensitization of nociceptive C fibres of the joint in response to mechanical stimuli, which was not attenuated by neutralizing TNF³⁷. Evidence showing successful inhibition of IL-17A–IL-17F heterodimers in axSpA has raised questions about the potential role of IL-17F in pain pathways³¹. This heterodimer shapes the potency, specificity and overall activity of these cytokines. Some studies suggest substantial redundancy between IL-17F and other IL-17 family members, but a distinct function of IL-17F has yet to be clearly defined^{38,39}.

Table 1 | Characteristics and drivers of pain phenotypes in axial spondyloarthritis

Aspect	Nociceptive pain	Neuropathic pain	Nociplastic pain
Mechanism	Tissue inflammation or damage	Compression of or damage to nerves	Central sensitization with amplified pain processing in the central nervous system
Distribution	Usually localized to affected areas (such as the low back and buttock area)	Pain can follow a radicular or peripheral nerve distribution (for example, sciatica) rather than staying localized to the back	Diffuse and widespread, not confined to sites of inflammation and can be multifocal (affecting the neck, back, peripheral joints and muscles)
Type	Often described as a dull, deep ache or throbbing	Burning, shooting, electric shock-like pain with possible numbness or tingling	Exceeds what objective findings would suggest, along with fatigue, poor sleep and symptoms of anxiety or depression
Timing	For inflammation-related nociceptive pain, symptoms are typically worse with rest and overnight and get better with activity Purely mechanical nociceptive pain might worsen with activity	Can be episodic and lancinating, with paraesthesia or weakness in the affected limb	Non-specific
Physical examination	Localized tenderness and swelling of the peripheral joints and entheses	Sensory loss or reflex changes corresponding to nerve damage along with diminished muscle strength	Physical examination is typically unrevealing apart from widespread tenderness (for example, positive tender points)
Imaging	Imaging can reveal active inflammatory changes in the axial skeleton	Inflammatory lesions do not explain this type of pain	Inflammatory lesions do not explain this type of pain
Main drivers of disease	Pro-inflammatory cytokines	Structural lesions in the peripheral nerve structure caused by mechanical damage or inflammation	Possible role of cytokines
Cytokines	Strong association with TNF and IL-17	Minor involvement of IL-17 and an unknown role of the JAK–STAT pathway	Minor involvement of IL-17 and the role of other cytokines is unclear
Main therapeutic interventions	NSAIDs, TNF inhibitors, IL-17 inhibitors and JAK inhibitors, in combination with non-pharmacological therapies	Gabapentinoids, SNRIs, TCAs, in combination with psychological approaches such as CBT and mindfulness	Gabapentinoids, SNRIs, TCAs, in combination with psychological approaches such as CBT and mindfulness
Mechanical stress	Exaggerated mechanical stress response is associated with enhanced IL-17 production	Role is unclear	Manual work increases pain, whereas regular physical activity is associated with better outcomes
Fibromyalgia	Persistent nociceptive pain is a risk factor for fibromyalgia	Fibromyalgia is associated with this type of pain	Fibromyalgia is considered a typical aspect of central sensitization
Female sex	Hormonal fluctuations, body composition and biomechanics are associated with this type of pain	More common in the female sex than in the male sex	More widespread pain pattern, more abnormalities on neuroimaging and more likely to experience diagnostic delay

CBT, cognitive behavioural therapy; GM–CSF, granulocyte–macrophage colony-stimulating factor; JAK, Janus kinase; SNRIs, serotonin–noradrenaline reuptake inhibitors; STAT, signal transducer and activator of transcription; TCAs, tricyclic antidepressants.

In addition, in some patients, persistent pain despite well-controlled inflammation suggests a major shift towards central pain mechanisms. Indeed, beyond nociceptive pain, cytokines might also contribute to neuropathic and nociplastic pain mechanisms. Chronic inflammation and cytokine-induced neuroinflammation can lead to structural and functional changes in peripheral and central pain pathways, potentially causing neuropathic pain^{40,41}. Animal studies have shown that IL-17 is involved in the development and maintenance of neuropathic pain, inflammatory pain and cancer pain^{42,43}. Other preclinical studies also suggest a role for IL-17A in neuropathic pain³⁴.

Additionally, cytokines can influence central sensitization, the hallmark of nociplastic pain, by altering the excitability of neurons in the spinal cord and brain, thereby amplifying pain signals even in the absence of ongoing inflammation⁴⁴. The dependence of central sensitization on the presence of some cytokines such as IL-17, granulocyte–macrophage colony-stimulating factor (GM–CSF) and IL-6 is debated⁴⁵.

Data from the British Society for Rheumatology biologics register indicate that high disease activity and widespread pain concomitant to axSpA can predict the development of secondary fibromyalgia,

whereas low disease activity, absence of widespread pain and initiation of TNF inhibitors predict resolution of fibromyalgia in axSpA⁴⁶. Subsequently, uncontrolled inflammation (that is, nociceptive pain) during the early stages of axSpA might be associated with the development of nociplastic pain in the long term⁴⁶.

Finally, several cytokines, including TNF and IL-17, signal via JAK, which explains the efficacy of JAK inhibitors in axSpA but also the positive effect of these therapies on pain symptoms⁴⁷. The JAK–STAT signalling pathway has been implicated in the processing of pain beyond its role in mediating inflammation. JAK inhibitors can alleviate a broad array of pain outcomes in both axial and peripheral joints⁵. Their effect on nociceptive pain has thus far been assessed using a limited number of questionnaires in cross-sectional studies, restricting firm conclusions; however, the JAK–STAT pathway is receiving increasing attention in the modulation of nociceptive responses given its clear role in cytokine signalling in several immune-mediated inflammatory diseases^{5,45,48}.

The role of mechanical stress

Mechanical stress has a crucial role in driving pain in axSpA by contributing to both inflammatory and structural disease processes.

In the context of nociceptive pain, mechanical stress might function as a trigger for inflammation. Repetitive mechanical strain, particularly at entheses, can lead to microdamage and elicit an immune system-mediated tissue-repair pathway in tendons and ligaments⁴⁹. Mechanistically, an exaggerated mechanical stress response is associated with enhanced IL-17A production at enthesal sites and sustained production of PGE₂ through COX-2 activation, driving chronic inflammation, pain and progressive structural damage^{31,50}. Ongoing inflammation at sites of high mechanical load can result in bone formation, syndesmophytes and ankylosis, leading to spinal stiffness and altered biomechanics^{51–53}. As the spine loses flexibility, adjacent segments can experience compensatory mechanical stress, further perpetuating pain and dysfunction. Indeed, physically demanding jobs were traditionally associated with worse outcomes in axSpA, including functional limitation and more new bone formation⁵⁴. Conversely, studies from the past 5 years indicate that exercise is associated with reduced disease activity⁵⁵. In addition, persistent mechanical stress might also contribute to nociplastic pain mechanisms⁵⁶.

The role of fibromyalgia and illness perception

The presence of fibromyalgia introduces a nociplastic component, thus contributing to increased pain severity, widespread pain distribution and reduced treatment response. Persistent nociceptive pain is itself a risk factor for the development of nociplastic pain⁵⁷ as fibromyalgia is

associated with sustained nociceptive pain in axSpA⁵. Studies suggest that fibromyalgia is more prevalent in patients with axSpA than in the general population; the overall prevalence of fibromyalgia in axSpA ranges from 4% to 25%^{18,58,59}.

In general, the presence of concomitant fibromyalgia in axSpA is associated with higher pain and disease activity scores²⁴, fatigue, sleep disturbances, functional impairment and reduced quality of life, compared with axSpA without fibromyalgia, even in patients with well-controlled inflammation. Statistically significant differences in the levels of C-reactive protein (CRP) or erythrocyte sedimentation rate are not typically observed when comparing patients with and those without fibromyalgia^{58–61}.

Finally, response to treatment is negatively affected by concomitant fibromyalgia. In a prospective observational cohort study, the percentage success after 12 weeks of TNF inhibitor treatment was lower in the group with secondary fibromyalgia for most of the efficacy endpoints (BASDAI 50 of 45.3% with fibromyalgia versus 54.1% without fibromyalgia), except for those endpoints related to CRP change⁶².

In addition, psychological factors, including illness perceptions, pain-related worrying, coping strategies and overall psychological well-being, are central components of the biopsychosocial model of health in rheumatic diseases and might influence disease activity assessment. In axSpA, negative illness perceptions can affect disease activity as assessed by the Ankylosing Spondylitis Disease Activity Score

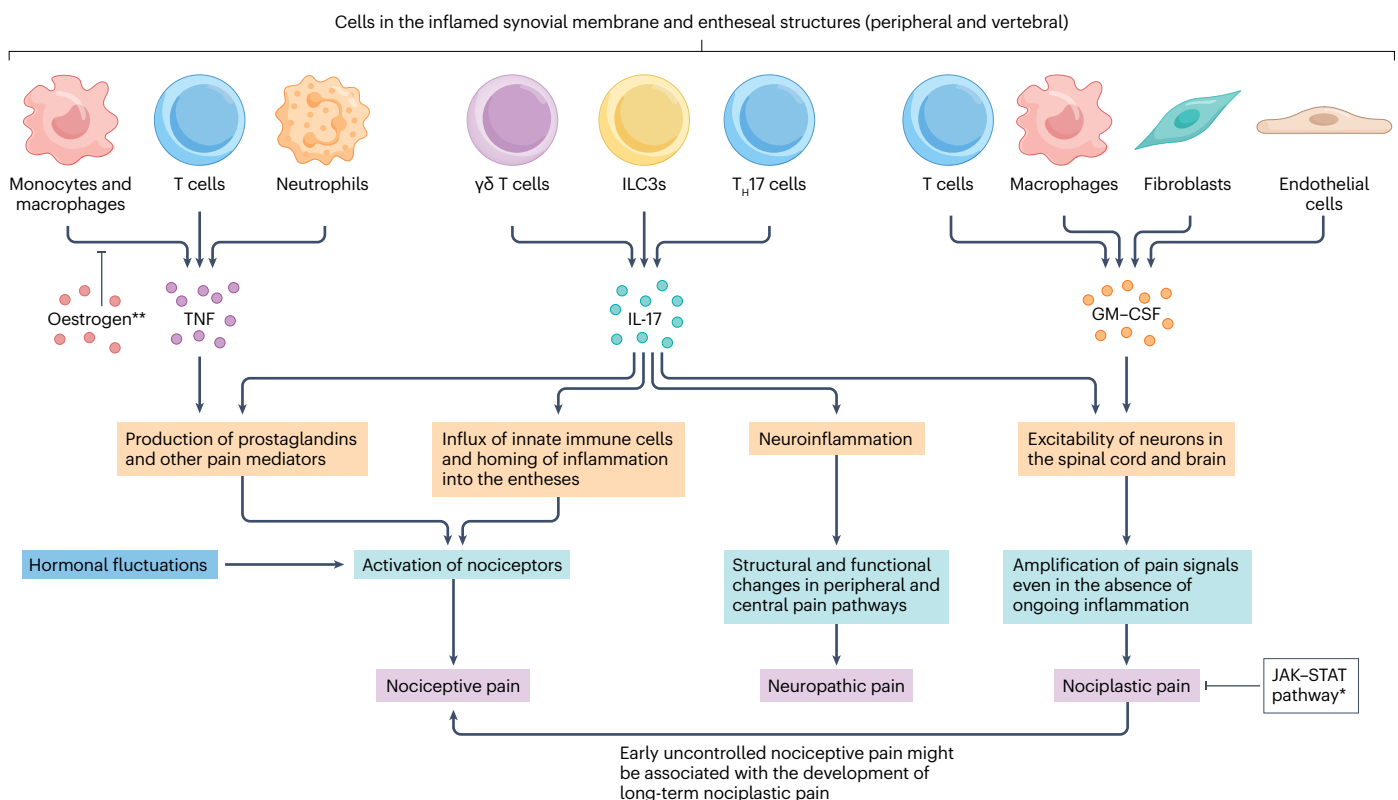


Fig. 1 | Cytokines involved in the development of inflammation in axSpA in relation to pain. Different cytokines are involved in the production and maintenance of high levels of different pro-inflammatory factors and cytokines in axial spondyloarthritis. All these factors drive different pro-inflammatory processes that lead to different types of pain. *For the modulation of neuropathic and nociplastic pain mechanisms, current clinical trials have primarily assessed

the efficacy of targeting JAK–STAT pathways in inflammatory pain contexts, but there are no specific methods to assess their impact on other pain phenotypes. **Oestrogen can modulate TNF activity, for example, by inhibiting TNF production by macrophages and monocytes. ILC, innate lymphoid cell; T_H17 cells, T helper 17 cells; GM–CSF, granulocyte–macrophage colony-stimulating factor.

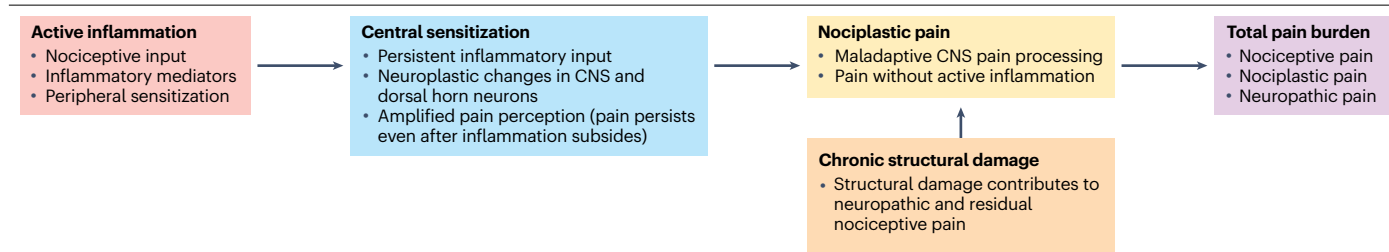


Fig. 2 | Continuum and interconnection of pain mechanisms in inflammatory disease. Active inflammation might lead to different pain types, starting from inflammatory-driven nociceptive input, which develops into neuroplastic

changes of the central nervous system and eventually involves the entire pain spectrum when disease becomes chronic. CNS, central nervous system.

(ASDAS)^{63,64}. Such negative perceptions, together with pain-related worrying, might encourage passive or avoidant coping strategies when managing symptoms such as pain, stiffness and fatigue⁶⁵.

The role of sex differences

Biological sex (that is, sex assigned at birth; throughout this Review, ‘sex’ is used as a shorthand for biological sex) has a substantial role in shaping the pain experience in axSpA, influencing pain location, severity, reporting and response to treatment. Historically, axSpA was considered to affect male individuals more than female individuals, but growing evidence suggests a more sex-balanced prevalence, especially in the earlier stages of the disease.

A study evaluating pain location in 170 patients with axSpA (63.5% of whom were male patients) found that the location and spread of pain were different in male and female patients and were related to worse clinical status. Axial thoracic pain was the least prevalent overall (lumbar, 74.4%; cervical, 47.6%; cervicothoracic, 47.6%; thoracic, 32.4%), but it was about three times more likely in female patients than in male patients (odds ratio (OR), 2.92; $P = 0.009$). Axial cervicothoracic junction pain spread more diffusely in female patients (OR 2.48; $P = 0.018$). Female individuals had a two- to three-fold higher likelihood of widespread axial (OR 3.33; $P = 0.007$) and peripheral articular (OR 2.34; $P = 0.023$) pain. In this study, widespread non-articular pain and low physician global assessment of disease activity was associated with worse Bath Ankylosing Spondylitis Functional Index (BASFI), BASDAI, Hospital Anxiety and Depression Scale (HADS) subscale anxiety and depression in male patients and worse fear of movement (Tampa Scale for Kinesiophobia (11-item version)) and HADS anxiety in female patients⁶⁶. Female patients were also more likely to have diagnoses of depression and fibromyalgia⁶⁷.

Furthermore, similar results were shown in a prospective multi-centre cohort study. Of the 494 patients with early axSpA who were followed-up for 6 years (mean \pm s.d. age of 31.9 ± 7.5 years, symptom duration of 20.7 ± 11.7 months, 50.4% of whom were male), female patients had higher ASDAS and patient global assessment over time than male patients ($P < 0.0001$ for both), with similar levels of CRP overall ($P = 0.089$), whereas structural damage increased more in male patients ($P < 0.001$)⁶⁸.

In addition to high disease activity, female sex in axSpA is associated with increased central sensitization. In a study that compared 116 patients with axSpA with 95 healthy individuals, the Central Sensitization Inventory (CSI) was higher in female individuals than in male individuals¹⁵. This association is also supported by several neuroimaging studies reporting structural and functional changes in the brains of female patients with axSpA compared with those of healthy

individuals⁶⁹. In a study that used modular analysis and machine learning to assess resting-state functional MRI data from 220 individuals with chronic low back pain as a result of axSpA, sex-specific network topological characteristics were observed. Women exhibited atypically higher functional segregation in the mid-cingulate cortex and subgenual anterior cingulate cortex and lower connectivity in the network with the default mode and frontoparietal modules, whereas men exhibited stronger connectivity with the sensorimotor module⁷⁰.

The reporting of higher pain levels, more widespread pain and greater functional impairment, despite similar levels of inflammation, suggests that nociplastic pain might be more prominent in female individuals. In addition, a study that used PET to assess neurotransmission during pain in healthy adults suggested that modulation of the endogenous opioid neurotransmitter circuitry by oestrogen might contribute to sex differences in pain sensitivity, in both male and female patients^{71,72}, which might be of particular interest owing to the high risk of nociplastic pain in axSpA.

In female individuals, low oestrogen was associated with reduced activation of endogenous opioid neurotransmission during a pain stressor and higher pain ratings, whereas a high-oestrogen state showed greater pain-induced regional activation of the endogenous opioid system⁷². Conversely, testosterone might exert anti-nociceptive effects⁷³, which might also explain differences in the pain occurrence and reporting of pain in patients diagnosed with axSpA.

Moreover, differences in body composition, biomechanics and muscle strength might affect spinal loading and pain perception differently in male and female individuals. Although male individuals have a higher body mass index (BMI) on average, female patients with axSpA have been reported to have a higher adipose tissue mass index and are more likely to have obesity than their male counterparts (28.6% compared with 7.1%), which has been linked to higher disease activity and subsequent pain^{71,74}.

These sex-related differences (Box 2) have important clinical implications, as female patients with axSpA might experience delayed diagnosis, more frequent misclassification as having fibromyalgia and different responses to biologic therapies. According to a 2025 systematic literature review and meta-analysis, male patients with axSpA are more likely to achieve an ASAS40 response than female patients for all advanced therapies (OR 1.88, 95% CI 1.44–2.46) and for IL-17A inhibitors (OR 1.82) and TNF inhibitors (OR 2.42). Male patients were also more likely to achieve ‘low disease activity’ or ‘inactive disease’ status on ASDAS scores (OR 2.19, 95% CI 1.47–3.26) across all advanced therapies and for IL-17A inhibitors (OR 2.08) and TNF inhibitors (OR 2.42) individually⁷⁵.

Acknowledging the influence of sex on pain in axSpA can support more personalized approaches to monitoring and treatment.

For example, in female patients, who more often present with widespread pain and features of central sensitization, early screening for fibromyalgia and the use of targeted non-pharmacological interventions such as cognitive behavioural therapy (CBT), exercise programmes and sleep optimization might be prioritized alongside anti-inflammatory therapy. For male patients, who typically present with localized inflammation-driven pain and tend to develop more structural damage than female patients, using imaging to identify the problem and timely initiation or escalation of biologic therapy, where appropriate, might be a better approach. Although these sex-specific considerations could improve treatment selection and reduce misclassification, prospective studies are needed to confirm whether such stratified strategies lead to better outcomes. In summary, a comprehensive understanding of the overlapping pain mechanisms in axSpA is essential for refining disease assessment and implementing effective, multimodal management strategies.

Assessment of pain in axial spondyloarthritis

To ascertain diagnosis, monitoring and treatment evaluation, assessment and measurement of pain in axSpA is crucial both for inflammation-driven but also so-called residual or non-inflammation-driven pain. Owing to the complex and multidimensional nature of pain, various assessments exist, which consider components of pain intensity, pain quality and impact on disease status (Table 2). These tools can be categorized into patient-reported outcome measures, composite indices and functional assessments.

Patient-reported outcome measures

The Visual Analogue Scale (VAS) and the Numerical Rating Scale (NRS) are simple methods for the assessment of general pain. They consist of a scale of 100 mm in length or ranging from 0 to 10, in which the left end represents 'no pain' and the right end represents 'worst pain possible'. Their application in daily practice is considered feasible because of their simplicity¹⁰. Furthermore, they can be easily adapted to specific painful areas of interest; for example, 'general pain' versus 'pain in the low back', 'pain in the buttocks' or 'pain in the hips'.

Composite indices

The BASDAI⁷⁶ comprises six questions that measure disease activity based on the point of view of the patient. Three of the questions in BASDAI refer to pain: overall pain in the neck, back or hip (question 2); pain in joints other than the neck, back or hips (question 3) and discomfort from an area tender to touch or pressure (question 4). Similarly, the ASDAS⁷⁷, which was derived from the BASDAI in a validated data-based process, also assesses pain in the back and in areas other than the neck, back or hips in relation to inflammatory complaints. These composite indices are the two most widely used questionnaires in axSpA and are also recommended by the ASAS for use in clinical studies. Overall, the ASDAS is preferred over the BASDAI as the set of questions included in the BASDAI are susceptible to different reasons for having pain, making it more susceptible to the influence of general pain and less likely to differentiative inflammation-related disease-activity complaints. This lack of objectivity has been mainly shown in patients with concomitant fibromyalgia when compared with those without concomitant fibromyalgia, as mentioned previously^{62,78}.

The PainDETECT questionnaire⁷⁹ is a tool for screening for neuropathic pain in patients with low back pain but is also widely used in patients with axSpA. The questionnaire comprises nine questions, seven of which relate to pain quality and sensory symptoms and

the other two relate to location of pain and pain progression over time. The patient answers using a scale of 0 to 5, in which 0 stands for no pain and 5 for very strong pain. A score of <13 is interpreted as absence of neuropathic pain, a score of 13–18 as possible presence of neuropathic pain and >18 as likely to have a neuropathic component to their pain⁷⁹.

Similarly, the Neuropathic Pain Symptom Inventory (NPSI)⁸⁰ also assesses neuropathic pain, in both a qualitative and a quantitative manner. The NPSI consists of 12 items, 10 of which are descriptors of different symptoms of neuropathic pain symptoms and the other 2 evaluate the duration of continuous and paroxysmal spontaneous pain. The average score (0–100) provides an overall measure of neuropathic pain severity. In addition, the NPSI enables the calculation of five subscores, each representing different aspects of neuropathic pain (burning superficial spontaneous pain, squeezing deep spontaneous pain, paroxysmal pain, evoked pain and paraesthesia or dysaesthesia). These subscores can be used to gain a more comprehensive assessment of the pain status of the patient and guide more targeted treatment decisions.

The McGill Pain Questionnaire (MPQ)⁸¹ is designed to capture qualitative aspects of pain, including sensory, affective and evaluative dimensions. The MPQ consists of 78 descriptors divided into 20 sections, enabling detailed description of the nature of pain. Patients select words from each category in the questionnaire that best describe their pain experience, which have an associated point value. This questionnaire also includes sections on pain intensity and temporal patterns. The advantage of the MPQ is that the results can distinguish between different pain qualities (such as throbbing versus stabbing), leading to identification of neuropathic pain components or central sensitization. However, the MPQ is also time consuming and seems to be complex for some patients, making it better suited to detailed assessments or research purposes.

The CSI⁸² is a questionnaire designed to identify symptoms related to central sensitization. Although not classed as a primary 'pain assessment tool', the CSI is helpful for distinguishing central from peripheral (inflammatory) pain mechanisms. The questionnaire includes 25 items and patients are asked to rate the frequency of symptoms concerning sleep, concentration difficulties or others on a scale from 0 (never) to 4 (always). Total higher scores indicate a greater likelihood of central sensitization.

Box 2 | Sex-based differences in pain reporting in axial spondyloarthritis

Women

- More widespread pain
- More thoracic location
- Higher self-reported disease activity
- More abnormalities on neuroimaging
- Pain levels are influenced by hormonal fluctuations, body composition and biomechanics
- More components of anxiety and depression

Men

- More radiographic damage
- Better self-reported quality of life
- Better response to therapy

Table 2 | Pain assessment instruments in axial spondyloarthritis

Tool	Assessment method	Pain type	Assessment details	Best used for	Additional notes
VAS	Pain intensity, quantitative	Nociceptive	Mark on a 10-cm line, ranging from 'no pain' at one end to 'worst possible pain' at the other	Quick assessments	Simple and visual
NRS	Pain intensity, quantitative	Nociceptive	Rating of 0–10 given verbally or in writing	Routine tracking	Easy to score
BASDAI	Axial and peripheral pain, quantitative	Nociceptive	Composite measure; six-item questionnaire	Measuring disease activity	Includes pain, fatigue and spinal stiffness
ASDAS	Axial and peripheral pain, quantitative	Nociceptive	Composite measure; four questions answered using a 0–10 scale plus levels of CRP or ESR	Disease-activity assessment	Includes objective inflammatory parameter (CRP or ESR)
PainDETECT	Quantitative	Neuropathic	Composite measure; nine questions	Detecting neuropathic pain	Overlap with nociceptive pain
NPSI	Quantitative and qualitative	Neuropathic	Composite measure; questionnaire that consists of 12 items	Comprehensive assessment of pain levels	Different aspects of neuropathic pain can be assessed
MPQ	Multidimensional, qualitative	Multidimensional	Composite measure; descriptor checklist, two versions (short and long) are available	In research settings or to assess complex pain	Time-intensive
CSI	Central-sensitization symptoms, quantitative	Nociplastic	Composite measure; 25 questions that involve rating symptoms	Non-inflammatory pain	Useful for people with high pain but low inflammation
BPI	Pain intensity and interference, quantitative	Function	Composite measure; questions on pain and function, two versions (short and long) are available	Understanding the impact that pain has on the patient	Quick and informative

ASDAS, Axial Spondyloarthritis Disease Activity Score; BASDAI, Bath Ankylosing Spondylitis Disease Activity Index; BPI, Brief Pain Inventory; CRP, C-reactive protein; CSI, Central Sensitization Inventory; ESR, erythrocyte sedimentation rate; MPQ, McGill Pain Questionnaire; NPSI, Neuropathic Pain Symptom Inventory; NRS, Numerical Rating Scale; VAS, Visual Analogue Scale.

The Brief Pain Inventory (BPI)⁸³ assesses both the intensity of pain and its interference with daily functions. It includes items that evaluate pain severity and the extent to which pain hinders activities such as walking, work and sleep, all of which are relevant to axSpA. In the BPI, patients rate their pain at its worst, least, average and current levels over the past 24 h on a scale of 0 (no pain) to 10 (pain as bad as you can imagine).

Overall, the assessment of pain in axSpA requires a combined approach. Different tools capture the different aspects of pain (Table 2), such as inflammatory and non-inflammatory dimensions. Thus far, a single tool that is validated to capture all these aspects does not exist. One in every two patients with axSpA has a high probability of central sensitization, as measured by the CSI. In one study, specific illness perceptions and obesity were independently associated with BASDAI and ASDAS, with CSI scores explaining 38% of the variance in the BASDAI and 23% of the variance in the ASDAS scores⁶⁴. Patients with higher CSI scores include female patients with a history of depression, clinical enthesitis and high scores for ASDAS, and BASDAI¹³. A 'possible' neuropathic component, using the PainDETECT questionnaire, was evident in only 8% and a 'likely' component in 1% of patients. For the 31% of patients with high disease activity, 79.2% showed either a 'possible' or 'likely' nociplastic component and 20.8% a 'possible' or 'likely' neuropathic component. Both neuropathic and nociplastic scores were shown to be independently associated with ASDAS and BASDAI scores²⁵.

Accurate assessment of pain in axSpA requires recognition of its multifactorial nature, including inflammatory, nociplastic and neuropathic components. A range of tools, including simple scales (VAS and NRS) to composite indices (BASDAI and ASDAS) and specific instruments that target pain mechanisms (PainDETECT, CSI and MPQ),

is available to capture different dimensions of pain. Although no single tool comprehensively addresses all aspects, combining multiple assessments can provide a more complete picture of the experience of the patient. Notably, central sensitization is common and might influence composite disease scores substantially.

Interconnection between the different types of pain in axial spondyloarthritis

Although accurate identification of the predominant pain mechanism in patients with axSpA is essential for optimizing management strategies, pain mechanisms are not mutually exclusive and frequently overlap. Bidirectional interactions exist; nociplastic and neuropathic pain might amplify nociceptive pain scores, whereas persistent and inadequately controlled nociceptive pain can facilitate the development of nociplastic and neuropathic pain. Although evidence in axSpA is limited, insights can be extrapolated from other chronic rheumatic and musculoskeletal conditions.

Nociplastic and neuropathic pain as drivers for nociceptive pain scores

In the prospective Groningen Leeuwarden axSpA (GLAS) cohort, which includes 332 patients with axSpA, illness perception, central sensitization (measured by CSI) and BMI had direct effects on ASDAS, beyond inflammatory parameters⁶³. In the GESPIC cohort (78 patients with axSpA who were treated with bDMARDs) 22% of patients had nociplastic pain (measured by WPI) and 9% had neuropathic pain (measured by PainDETECT). Both types of pain were associated with the presence of residual symptoms (defined by BASDAI of ≥ 4)²⁵. In the CorEvitas registry (1,823 patients with psoriatic arthritis), 11.1% of patients met

the definition for fibromyalgia and 20.6% were considered to have widespread pain. These patients reported approximately two-fold higher clinical Disease Activity in Psoriatic Arthritis scores, pain, global assessment and tender joint counts compared with those without fibromyalgia⁸⁴. Nociceptive pain thus confounded disease-activity assessment and impaired treat-to-target strategies.

Nociceptive pain as a precursor to nociceptive pain

Data from patients with rheumatoid arthritis (RA) suggest that systemic inflammation might lead to changes in functional central nervous system pathways, leading to the development of nociceptive pain⁸⁵. Emerging neuroimaging studies have helped to provide insights into how inflammation might interact with central pain pathways. Moreover, a post hoc analysis supported by functional MRI studies showed that the erythrocyte sedimentation rate correlated positively with functional connectivity between the insula and the left inferior parietal lobule in patients with RA and concomitant fibromyalgia but not in patients with RA alone⁸⁶. This finding suggests that nociceptive pain might indeed be an intrinsic part of systemic rheumatic diseases, resulting from peripheral inflammation stimuli driving bottom-up processes that trigger the development of nociceptive pain.

The presence of pro-inflammatory cytokines and mediators can contribute to central sensitization via their effect on neuronal transmission, which is characterized by increased excitatory activity and reduced inhibitory activity⁸⁷. In rodent models, the use of lipopolysaccharide as a pro-inflammatory stimulus induced the development of hyperalgesia⁸⁸ and other centrally mediated symptoms, including fatigue, sleep and cognitive and mood disturbances⁸⁹.

These data indicate a putative role for systemic inflammation in the development of nociceptive pain, in the context of systemic inflammatory disease, offering evidence of a bottom-up nociceptive dimension induced by peripheral inflammatory nociceptive processes that sensitize pronociceptive central nervous system pathways^{90,91}. An effective and swift control of inflammation leading to reduced long-term nociceptive pain in axSpA, and in other inflammatory rheumatic disease, has yet to be demonstrated.

Treatment options for pain in axial spondyloarthritis

Effective management of pain in axSpA is challenging as patients might experience multiple pain types. Clinicians must therefore assess the origin of pain (inflammatory versus non-inflammatory) to adjust treatments appropriately. Here, we provide a guide to help manage different sources of pain in patients with axSpA. Nevertheless, highlighting that recommendations for nociceptive and neuropathic pain rely largely on indirect or preclinical evidence is important. Most of the available data are extrapolated from other conditions such as fibromyalgia, diabetic neuropathy or post-herpetic neuralgia, with only limited, indirect or small observational studies directly addressing axSpA.

Inflammation-related nociceptive pain interventions

Non-pharmacological therapies. Non-pharmacological therapies continue to have a crucial role in the management of pain in patients with axSpA^{92,93}. Although pharmacological treatments are essential for controlling inflammation and disease progression, non-drug interventions complement these treatments by improving function and reducing pain.

Patient education and self-management are key aspects in the management of axSpA. Understanding pain triggers and how to balance

activities can help patients to control their symptoms better. Additionally, smoking cessation is strongly advised, as it exacerbates spinal damage and might worsen pain. Exercise (such as aerobic exercise and range-of-motion or stretching exercises) and physical therapy can also be beneficial for people with axSpA. Regular exercise can improve an inflammatory immune profile, coinciding with pain reduction⁹⁴. In addition, a meta-analysis confirmed that exercise programmes led to considerable improvements in pain and function compared with no exercise⁹⁵.

Pharmacological treatments. NSAIDs are first-line therapy in axSpA. NSAIDs target inflammatory nociceptive pain by reducing prostaglandin-mediated inflammation⁹². NSAIDs are recommended at maximum tolerated doses as initial treatment for active axSpA. Most patients experience substantial pain relief with NSAIDs, often within days. Continuous NSAID therapy is preferred in those with persistent symptoms, whereas on-demand use might suffice for milder disease. Common choices of NSAIDs include etoricoxib, naproxen, indomethacin and ibuprofen, with selection guided by patient comorbidities and risk factors (such as gastrointestinal or cardiovascular risk)⁹². NSAID-responsive pain suggests predominantly inflammatory mechanisms. Conversely, lack of NSAID response should prompt evaluation of other pain-type contributors; however, in longstanding disease, the discriminative capacity of NSAIDs response might disappear⁹⁶.

In clinical practice, bDMARDs, such as TNF and IL-17 inhibitors are recommended after failure of ≥ 2 NSAIDs^{92,97}. Multiple randomized controlled trials have demonstrated that TNF inhibitors (adalimumab, certolizumab pegol, etanercept, infliximab and golimumab) and IL-17 inhibitors (secukinumab, ixekizumab and bimekizumab) quickly improve total spinal and night pain⁹⁸. These agents target the pro-inflammatory cytokines that promote nociceptive pain, often resulting in substantial pain improvement when successful. Not much is known about the impaired pain pathways in these patients and the effects of bDMARDs on these pathways. In patients with RA and SpA, impaired descending pain modulation (assessed by conditioned pain modulation) improved over 6 months following initiation of TNF inhibitor therapy, whereas no statistically significant difference in thermal pain thresholds was observed, suggesting a possible effect of TNF inhibitors on central pain modulation⁹⁹. Patients with certain extra-musculoskeletal manifestations might have specific biologic preferences; for example, TNF inhibitor monoclonal antibodies are preferred if the patient has coexistent inflammatory bowel disease or recurrent uveitis and IL-17 inhibitors are better for those with severe psoriasis⁹².

Treatment with tsDMARDs (such as the JAK inhibitors tofacitinib and upadacitinib) can interrupt multiple cytokine pathways relevant to both inflammation and pain processing⁴⁵. Randomized control trials indicate that JAK inhibitors can produce fast and sustained pain relief in axSpA^{93,100,101} and these drugs have demonstrated clinically relevant improvements in patient global pain as early as week 2 of treatment^{47,102}.

Notably, JAK inhibitors might also modulate neuropathic and nociceptive pain mechanisms via the JAK-STAT pathway. However, current clinical trials (NCT03178487, NCT04169373, NCT03502616) have primarily assessed their efficacy in inflammatory pain contexts, without using specific methodologies to evaluate their impact on other pain phenotypes. Although preclinical and early clinical data suggest a potential role for JAK inhibitors in modulating nociceptive and neuropathic pain, acknowledging that current clinical evidence remains

sparse and exploratory is important. Thus far, studies have not been designed to test the superiority of JAK inhibitors over established treatments, and thus the claim of therapeutic advantage remains unconfirmed by robust clinical data. The UPSTAND trial (NCT04846244) is an ongoing observational study designed to evaluate the real-world effectiveness of upadacitinib in achieving early and sustained pain control in adult patients with axSpA.

Even with optimal anti-inflammatory treatment, some patients still experience residual pain. Studies show that up to 50% of patients with axSpA who are in clinical remission or have low disease activity continue to report substantial pain⁹⁷. This residual pain is often caused by nociplastic or structural mechanisms, which require additional therapies beyond bDMARDs or tsDMARDs, and usually adjunct analgesics are used. Analgesics such as paracetamol (acetaminophen) or opioid-based drugs might be considered for refractory pain after other treatments have failed, are contraindicated or poorly tolerated⁹². Acetaminophen can be useful for mild-to-moderate pain and has a favourable safety profile, although its efficacy in severe axial pain is limited¹⁰³. Tramadol is sometimes used for moderate pain that is not relieved by NSAIDs. Strong opioids (such as morphine or oxycodone) are generally avoided in axSpA owing to modest efficacy for inflammatory pain and considerable long-term risks (including tolerance and dependence)¹⁰⁴.

Interventional and advanced therapies. Intra-articular glucocorticoid injections are particularly useful in managing local pain flares (such as an acutely inflamed sacroiliac joint or a swollen peripheral enthesis) in patients who are on otherwise effective systemic therapy. A 2025 meta-analysis of injectable therapies in SpA found that glucocorticoid injections of the sacroiliac joints led to a large reduction in pain VAS (from -8/10 to -3/10) at short-term follow-up¹⁰⁵. This pain relief tends to persist for several months but might decrease over time and not all patients respond to treatment.

Non-inflammatory nociceptive pain interventions

Non-inflammatory nociceptive pain arises from mechanical or structural abnormalities rather than active inflammation. In axSpA, recognizing factors such as joint damage, altered biomechanics and postural stress is important when selecting the most appropriate treatment as they can contribute to persistent pain, even in the absence of inflammatory activity.

Non-pharmacological therapies. Supervised physical therapy (including hydrotherapy) can adjust an exercise regimen to maintain spinal flexibility, strengthen core muscles and improve posture, all of which help to reduce mechanical pain¹⁰⁶. Advice on posture and joint protection can mitigate pain from daily mechanical stress. In advanced disease with spinal fusion, physical therapists might recommend adaptive strategies and devices (such as a shower chair or long-handled tools for hygiene) to minimize pain with activities. Although not directly analgesic, these interventions prevent exacerbation of pain owing to poor biomechanics¹⁰⁷.

Interventional therapies. Orthopaedic evaluation is important for structural sources of pain that might be corrected via surgical intervention. Total hip arthroplasty in patients with severe hip-joint arthritis has shown pain relief and function improvement⁹². Patients with fused spines and a severe kyphotic deformity causing pain and functional limitations might benefit from spinal osteotomy procedures to realign

the spine⁹². Additionally, if a patient has a spinal fracture, surgical stabilization or decompression could be urgently required to relieve pain and prevent neurological injury.

Nociplastic pain interventions

Importantly, recognizing nociplastic pain prevents unnecessary escalation of anti-inflammatory therapy, which might be ineffective and carry adverse effects.

Non-pharmacological therapies. Exercise is the most effective therapy in the management of fibromyalgia, underscoring its importance for nociplastic pain. Adequate sleep and stress reduction can mitigate central sensitization and fatigue. Mindfulness-based stress reduction and meditative movement therapies have shown improvements in fibromyalgia pain¹⁰⁴.

Chronic pain in axSpA can lead to distress, anxiety, depression and maladaptive coping, which in turn amplify pain perception¹⁰⁸. Psychological therapies target maladaptive coping and central pain amplification. Established approaches include CBT, acceptance and commitment therapy and interventions such as mindfulness-based stress reduction^{109–112}. These methods can reduce fear avoidance, catastrophizing and improve coping strategies, although evidence in inflammatory conditions remains limited¹¹³.

Pain-reprocessing therapy (PRT) is a newer psychological approach designed specifically for chronic nociplastic pain. It focuses on retraining brain pathways that perpetuate pain by combining education, cognitive reframing, somatic tracking (mindful, non-fearful attention to pain sensations) and gradual exposure to feared movements. In a randomized trial of chronic low back pain, PRT led to marked and sustained reductions in pain intensity¹¹³. However, there is currently no direct evidence supporting the use of PRT in inflammatory diseases such as axSpA, in which pain mechanisms also include active inflammation and structural damage.

Overall, existing evidence supports the importance of combining lifestyle interventions, physical activity and psychological therapies as part of a multimodal strategy in patients with predominantly nociplastic pain, while emphasizing the urgent need for studies specific to inflammatory rheumatic conditions.

Pharmacological therapies. Neurotransmitters have a central role in pain perception by transmitting signals between neurons in the pain pathway, influencing whether pain is amplified or dampened. Excitatory neurotransmitters such as glutamate and substance P enhance pain signalling, whereas inhibitory neurotransmitters such as GABA, serotonin and noradrenaline suppress it. Specifically targeting these neurotransmitters with drugs can reduce pain intensity and alter the way in which the brain processes painful stimuli, which makes neurotransmitter-targeted therapies a scientifically grounded strategy for pain relief. Pharmacological therapies should be considered for those with persistent nociplastic pain. However, evidence of the use of these interventions to improve pain and other symptoms is limited and slow titration of these medications and monitoring for adverse effects (such as sedation and dizziness) is essential^{114,115}.

Serotonin and noradrenaline reuptake inhibitors (SNRIs; such as duloxetine and milnacipran) have demonstrated analgesic effects in addition to mood-stabilizing properties¹¹⁵. Tricyclic anti-depressant medications (such as low-dose amitriptyline) and muscle relaxants (such as cyclobenzaprine) can improve sleep disturbance^{116,117}.

Gabapentinoids (gabapentin or pregabalin) can improve pain and sleep disturbance¹¹⁸.

Neuropathic pain interventions

Neuropathic pain arises from dysfunction or injury to the somatosensory nervous system and can be observed in axSpA. Recognizing neuropathic components is crucial for selecting the appropriate pharmacological interventions, such as gabapentinoids or certain anti-depressant medications, which target neural sensitization mechanisms.

Non-pharmacological therapies. Non-pharmacological therapies are vital in the management of neuropathic pain, complementing pharmacological treatments⁸⁵. Physical therapy and structured exercise help to reduce pain and improve function. Psychological approaches such as CBT and mindfulness address emotional aspects and pain perception. Patient education and self-management strategies enhance understanding and adherence. Complementary therapies, including acupuncture and yoga, might provide additional relief^{119,120}.

Pharmacological therapies. For patients with a clear neuropathic pain component (for example, radiculopathy or peripheral neuropathy), adjuvant neuropathic pain medications are indicated. Gabapentinoids (gabapentin or pregabalin) are useful for nerve pain and are recommended, especially if there are radicular symptoms. In patients with axSpA, pregabalin can help refractory pain while facilitating opioid tapering¹²¹. SNRIs (such as duloxetine and milnacipran) reduce pain by increasing serotonin and noradrenaline levels in the central nervous system, thereby strengthening the descending inhibitory pathways that suppress pain-signal transmission in the spinal cord¹²². Tricyclic anti-depressant medications (such as low-dose amitriptyline) might be used for certain individuals. Clinical evidence supports the efficacy of amitriptyline in conditions such as diabetic neuropathy and postherpetic neuralgia, often showing moderate pain relief¹²³.

Interventional and advanced therapies. Interventional and advanced therapies might be considered on a case-by-case basis by pain specialists, especially if imaging shows advanced structural changes that might be causing pain. Epidural steroid injections can be offered for severe radicular pain (for example, disc herniation or spinal stenosis in the context of ankylosis)¹²⁴. If facet joint arthritis or enthesal pain contributes to chronic back pain, local anaesthetic blocks can help. If the latter intervention is effective, longer-term relief can be achieved with radiofrequency ablation¹²⁵.

Neuromodulatory therapies, such as transcutaneous electrical nerve stimulation and spinal-cord stimulation, are additional options for difficult-to-treat pain. Transcutaneous electrical nerve stimulation is a non-invasive modality whereby surface electrodes deliver mild electrical currents to modulate pain signalling, which might provide short-term relief for muscle pain or localized back pain. For truly intractable pain for which all conventional therapies have failed, spinal-cord stimulation (implantation of electrodes in the epidural space to deliver electrical pulses) can be considered¹²⁶.

In summary, axSpA pain is often multifactorial and requires a multimodal and mechanism-based treatment approach. Recognizing and treating the entire spectrum, from inflammatory to nociceptive, neuropathic and nociplastic pain, require a paradigm shift from inflammation-centric to mechanism-based, individualized therapy. A multidisciplinary, patient-centred approach remains the cornerstone of effective pain management in axSpA.

Future directions and research needs

Although anti-inflammatory therapies have revolutionized axSpA treatment, pain control remains suboptimal for many patients. Future research should be aimed at identifying reliable biomarkers to distinguish pain phenotypes and stratify patients according to predominant pain mechanisms, nociceptive, nociplastic or neuropathic. Stratified treatment algorithms tailored to these profiles might improve outcomes. Novel therapies that target central sensitization and neural mechanisms are especially needed for patients with features of nociplastic or neuropathic pain, who often do not respond to anti-inflammatory agents.

Validated tools to distinguish pain subtypes in clinical practice remain limited and should be prioritized in future work, which is especially relevant in the context of the evolving difficult-to-manage and treatment-refractory axSpA concepts¹²⁷. Moreover, the role of sex differences in pain perception and response to treatment requires further exploration to inform sex-specific management approaches. Digital technologies such as wearable devices and app-based symptom tracking offer opportunities for real-time pain monitoring and might support more personalized, responsive care strategies. In parallel, research into multimodal and multidisciplinary care models, including psychological support and rehabilitation, should be expanded. Altogether, these efforts could bridge the current gap between inflammatory disease control and comprehensive pain relief in axSpA.

Conclusion

Although anti-inflammatory therapies effectively address inflammation-driven nociceptive pain, many patients continue to report pain despite achieving remission or low disease activity by conventional metrics. The achievement of such treatment outcomes underscores the role of additional pain mechanisms, including central sensitization and structural or nerve-related factors. In clinical practice, moving beyond the traditional inflammation-centred model and adopting a multidimensional framework that considers the heterogeneity of pain experiences is crucial for the decision of which pharmacological or non-pharmacological strategy is appropriate for each patient. However, further research on inflammation and pain as interconnected targets in axSpA needs to be considered a high priority in the field of axSpA and beyond. This research should include the improvement of screening and diagnostic tools, tailored interventions and digital solutions. The ultimate goal for axSpA management should be optimal targeting of both aspects of disease burden, inflammation and pain, and their interconnected qualities.

Published online: 21 January 2026

References

1. van der Heijde, D. et al. Goodbye to the term 'ankylosing spondylitis', hello 'axial spondyloarthritis': time to embrace the ASAS-defined nomenclature. *Ann. Rheum. Dis.* **83**, 547–549 (2024).
2. Calin, A., Porta, J., Fries, J. F. & Schurman, D. J. Clinical history as a screening test for ankylosing spondylitis. *JAMA* **237**, 2613–2614 (1977).
3. Rudwaleit, M., Metter, A., Listing, J., Sieper, J. & Braun, J. Inflammatory back pain in ankylosing spondylitis: a reassessment of the clinical history for application as classification and diagnostic criteria. *Arthritis Rheum.* **54**, 569–578 (2006).
4. Sieper, J. et al. New criteria for inflammatory back pain in patients with chronic back pain: a real patient exercise by experts from the Assessment of SpondyloArthritis International Society (ASAS). *Ann. Rheum. Dis.* **68**, 784–788 (2009).
5. Horbal, N. & Maksymowych, W. P. Nociplastic pain in axial spondyloarthritis and psoriatic arthritis: role of JAK kinases in immunopathology and therapeutic impact of JAK inhibitors. *Expert Rev. Clin. Immunol.* **21**, 137–152 (2025).
6. Minhas, D. & Clauw, D. J. Pain mechanisms in patients with rheumatic diseases. *Rheum. Dis. Clin. North. Am.* **47**, 133–148 (2021).
7. Cohen, S. P., Vase, L. & Hooten, W. M. Chronic pain: an update on burden, best practices, and new advances. *Lancet* **397**, 2082–2097 (2021).

8. Murphy, A. E., Minhas, D., Clauw, D. J. & Lee, Y. C. Identifying and managing nociplastic pain in individuals with rheumatic diseases: a narrative review. *Arthritis Care Res.* **75**, 2215–2222 (2023).
9. Poddubnyy, D. & Sieper, J. Mechanism of new bone formation in axial spondyloarthritis. *Curr. Rheumatol. Rep.* **19**, 55 (2017).
10. Sieper, J. et al. The Assessment of SpondyloArthritis international Society (ASAS) handbook: a guide to assess spondyloarthritis. *Ann. Rheum. Dis.* **68**, ii1–ii44 (2009).
11. Poddubnyy, D. et al. Physical function and spinal mobility remain stable despite radiographic spinal progression in patients with ankylosing spondylitis treated with TNF- α inhibitors for up to 10 years. *J. Rheumatol.* **43**, 2142–2148 (2016).
12. Fitzcharles, M. A. et al. Nociplastic pain: towards an understanding of prevalent pain conditions. *Lancet* **397**, 2098–2110 (2021).
13. Kieskamp, S. C. et al. Central sensitization has major impact on quality of life in patients with axial spondyloarthritis. *Semin. Arthritis Rheum.* **52**, 151933 (2022).
14. Aykurt Karlibel, I. & Kasapoğlu Aksoy, M. The relationship between central sensitization and disease activity, quality of life, and sleep quality among patients with axial spondyloarthritis. *Ir. J. Med. Sci.* **192**, 481–489 (2023).
15. Sas, S., Cengiz, G. & Kaplan, H. The effect of central sensitization on disease activity measures, quality of life and clinical parameters in axial spondyloarthritis: a cross-sectional study. *J. Rheum. Dis.* **30**, 176–184 (2023).
16. Sariyildiz, A., Coskun Benlidayi, I., Turk, I., Zengin Acemoglu, S. S. & Unal, I. Biopsychosocial factors should be considered when evaluating central sensitization in axial spondyloarthritis. *Rheumatol. Int.* **43**, 923–932 (2023).
17. Yücel, F. N. & Duruöz, M. T. Central sensitization in axial spondyloarthritis: an explorative study with quantitative sensory testing and clinical scales. *Mod. Rheumatol.* **32**, 1137–1145 (2022).
18. Jones, G. T., Mallawaarachchi, B., Shim, J., Lock, J. & Macfarlane, G. J. The prevalence of fibromyalgia in axial spondyloarthritis. *Rheumatol. Int.* **40**, 1581–1591 (2020).
19. Clauw, D. J. From fibrositis to fibromyalgia to nociplastic pain: how rheumatology helped get us here and where do we go from here? *Ann. Rheum. Dis.* **83**, 1421–1427 (2024).
20. Dutta, D. et al. Heritability of the fibromyalgia phenotype varies by age. *Arthritis Rheumatol.* **72**, 815–823 (2020).
21. Kato, K., Sullivan, P. F., Evengard, B. & Pedersen, N. L. A population-based twin study of functional somatic syndromes. *Psychol. Med.* **39**, 497–505 (2009).
22. Glass, J. M. et al. The effect of brief exercise cessation on pain, fatigue, and mood symptom development in healthy, fit individuals. *J. Psychosom. Res.* **57**, 391–398 (2004).
23. Kaplan, C. M. et al. Deciphering nociplastic pain: clinical features, risk factors and potential mechanisms. *Nat. Rev. Neurol.* **20**, 347–363 (2024).
24. Tsiami, S. et al. Performance of standardized patient reported outcomes developed for spondyloarthritis in primary and concomitant forms of fibromyalgia. *Arthritis Res. Ther.* **26**, 141 (2024).
25. Al Mohamad, F. et al. Association of nociplastic and neuropathic pain components with the presence of residual symptoms in patients with axial spondyloarthritis receiving biological disease-modifying antirheumatic drugs. *RMD Open* **10**, e004009 (2024).
26. Kim, T. W., Son, S. M. & Lee, J. S. Neuropathic pain in ankylosing spondylitis: a meta-analysis. *J. Rheumatol.* **79**, 95–102 (2020).
27. Borman, P., Kaygisiz, F. & Yaman, A. Neuropathic component of low back pain in patients with ankylosing spondylitis. *Mod. Rheumatol.* **31**, 462–467 (2021).
28. Braun, J. et al. Use of immunohistologic and in situ hybridization techniques in the examination of sacroiliac joint biopsy specimens from patients with ankylosing spondylitis. *Arthritis Rheum.* **38**, 499–505 (1995).
29. Chen, S. et al. Macrophages in immunoregulation and therapeutics. *Signal. Transduct. Target. Ther.* **8**, 207 (2023).
30. Jang, D. I. et al. The role of tumor necrosis factor alpha (TNF- α) in autoimmune disease and current TNF- α inhibitors in therapeutics. *Int. J. Mol. Sci.* **22**, 2719 (2021).
31. Schett, G., McInnes, I. B. & Neurath, M. F. Reframing immune-mediated inflammatory diseases through signature cytokine hubs. *N. Engl. J. Med.* **385**, 628–639 (2021).
32. Braun, J., Sieper, J. & Dougados, M. The history of ankylosing spondylitis/axial spondyloarthritis — what is the driving force of new knowledge? *Semin. Arthritis Rheum.* **71**, 152611 (2025).
33. Baraliakos, X., Kiltz, U., Kononenko, I. & Ciurea, A. Treatment overview of axial spondyloarthritis in 2023. *Best. Pract. Res. Clin. Rheumatol.* **37**, 101858 (2023).
34. McGonagle, D. G., McInnes, I. B., Kirkham, B. W., Sherlock, J. & Moots, R. The role of IL-17A in axial spondyloarthritis and psoriatic arthritis: recent advances and controversies. *Ann. Rheum. Dis.* **78**, 1167–1178 (2019).
35. Ceribelli, A. et al. Clinical trials supporting the role of the IL-17/IL-23 axis in axial spondyloarthritis. *Front. Immunol.* **12**, 622770 (2021).
36. Schett, G. et al. Enthesitis: from pathophysiology to treatment. *Nat. Rev. Rheumatol.* **13**, 731–741 (2017).
37. Richter, F. et al. Interleukin-17 sensitizes joint nociceptors to mechanical stimuli and contributes to arthritic pain through neuronal interleukin-17 receptors in rodents. *Arthritis Rheum.* **64**, 4125–4134 (2012).
38. Ruiz de Morales, J. M. G. et al. Critical role of interleukin (IL)-17 in inflammatory and immune disorders: an updated review of the evidence focusing in controversies. *Autoimmun. Rev.* **19**, 102429 (2020).
39. Chang, S. H. & Dong, C. IL-17F: regulation, signaling and function in inflammation. *Cytokine* **46**, 7–11 (2009).
40. Shamash, S., Reichert, F. & Rotshenker, S. The cytokine network of Wallerian degeneration: tumor necrosis factor- α , interleukin-1 α , and interleukin-1 β . *J. Neurosci.* **22**, 3052–3060 (2002).
41. Liedtke, W. STING-ing pain: how can pro-inflammatory signaling attenuate pain? *Neurosci. Bull.* **37**, 1075–1078 (2021).
42. Zhao, J., Huh, Y., Bortsov, A., Diatchenko, L. & Ji, R. R. Immunotherapies in chronic pain through modulation of neuroimmune interactions. *Pharmacol. Ther.* **248**, 108476 (2023).
43. Jiang, X. et al. Interleukin-17 as a potential therapeutic target for chronic pain. *Front. Immunol.* **13**, 999407 (2022).
44. Atta, A. A., Ibrahim, W. W., Mohamed, A. F. & Abdelkader, N. F. Microglia polarization in nociplastic pain: mechanisms and perspectives. *Inflammopharmacology* **31**, 1053–1067 (2023).
45. Selmi, C. et al. Pain in axial spondyloarthritis: role of the JAK/STAT pathway. *Front. Immunol.* **15**, 1341981 (2024).
46. Provan, S. A., Dean, L. E., Jones, G. T. & Macfarlane, G. J. The changing states of fibromyalgia in patients with axial spondyloarthritis: results from the British Society of Rheumatology biologics register for ankylosing spondylitis. *Rheumatology* **60**, 4121–4129 (2021).
47. McInnes, I. B. et al. Effect of upadacitinib on reducing pain in patients with active psoriatic arthritis or ankylosing spondylitis: post hoc analysis of three randomised clinical trials. *RMD Open* **8**, e002049 (2022).
48. Crispino, N. & Ciccia, F. JAK/STAT pathway and nociceptive cytokine signalling in rheumatoid arthritis and psoriatic arthritis. *Clin. Exp. Rheumatol.* **39**, 668–675 (2021).
49. Gracey, E. et al. Tendon and ligament mechanical loading in the pathogenesis of inflammatory arthritis. *Nat. Rev. Rheumatol.* **16**, 193–207 (2020).
50. Cambre, I. et al. Mechanical strain determines the site-specific localization of inflammation and tissue damage in arthritis. *Nat. Commun.* **9**, 4613 (2018).
51. Iyer, P., Hwang, M., Ridley, L. & Weisman, M. M. Biomechanics in the onset and severity of spondyloarthritis: a force to be reckoned with. *RMD Open* **9**, e003372 (2023).
52. Fassio, A. et al. Osteoimmunology of spondyloarthritis. *Int. J. Mol. Sci.* **24**, 14924 (2023).
53. Perrotta, F. M., Lories, R. & Lubrano, E. To move or not to move: the paradoxical effect of physical exercise in axial spondyloarthritis. *RMD Open* **7**, e001480 (2021).
54. Ward, M. M., Weisman, M. H., Davis, J. C. Jr. & Reveille, J. D. Risk factors for functional limitations in patients with long-standing ankylosing spondylitis. *Arthritis Rheum.* **53**, 710–717 (2005).
55. Sveaas, S. H. et al. High intensity exercise for 3 months reduces disease activity in axial spondyloarthritis (axSpA): a multicentre randomised trial of 100 patients. *Br. J. Sports Med.* **54**, 292–297 (2020).
56. Carbo, M. et al. Higher levels of physical activity are associated with less evasive coping, better physical function and quality of life in patients with axial spondyloarthritis. *PLoS ONE* **19**, e0301965 (2024).
57. Kosek, E. et al. Chronic nociplastic pain affecting the musculoskeletal system: clinical criteria and grading system. *Pain* **162**, 2629–2634 (2021).
58. Duffield, S. J., Miller, N., Zhao, S. & Goodson, N. J. Concomitant fibromyalgia complicating chronic inflammatory arthritis: a systematic review and meta-analysis. *Rheumatology* **57**, 1453–1460 (2018).
59. Macfarlane, G. J. et al. Co-Occurrence and characteristics of patients with axial spondyloarthritis who meet criteria for fibromyalgia: results from a UK national register. *Arthritis Rheumatol.* **69**, 2144–2150 (2017).
60. Bello, N., Etcheto, A., Beal, C., Dougados, M. & Molto, A. Evaluation of the impact of fibromyalgia in disease activity and treatment effect in spondyloarthritis. *Arthritis Res. Ther.* **18**, 42 (2016).
61. Wach, J., Letroublon, M. C., Coury, F. & Tebib, J. G. Fibromyalgia in spondyloarthritis: effect on disease activity assessment in clinical practice. *J. Rheumatol.* **43**, 2056–2063 (2016).
62. Molto, A. et al. Evaluation of the impact of concomitant fibromyalgia on TNF alpha blockers' effectiveness in axial spondyloarthritis: results of a prospective, multicentre study. *Ann. Rheum. Dis.* **77**, 533–540 (2018).
63. Kieskamp, S. C. et al. Interrelationships of disease activity, central sensitization, psychosocial and lifestyle factors in axial spondyloarthritis. *Rheumatology* **64**, 3547–3555 (2025).
64. Kieskamp, S. C. et al. Central sensitization, illness perception and obesity should be considered when interpreting disease activity in axial spondyloarthritis. *Rheumatology* **60**, 4476–4485 (2021).
65. Leventhal, H., Phillips, L. A. & Burns, E. The Common-Sense Model of Self-Regulation (CSM): a dynamic framework for understanding illness self-management. *J. Behav. Med.* **39**, 935–946 (2016).
66. Swinnen, T. W., Westhovens, R., Dankaerts, W. & de Vlam, K. Widespread pain in axial spondyloarthritis: clinical importance and gender differences. *Arthritis Res. Ther.* **20**, 156 (2018).
67. Webers, C. et al. Gender-attributable differences in outcome of ankylosing spondylitis: long-term results from the Outcome in Ankylosing Spondylitis International Study. *Rheumatology* **55**, 419–428 (2016).
68. Aouad, K. et al. Influence of sex on early axial spondyloarthritis: a six-year longitudinal analysis from a large national cohort. *Arthritis Care Res.* **75**, 2107–2116 (2023).
69. Lee, Y. C., Malfait, A. M. & Ogdie, A. R. Unmet needs in spondyloarthritis: understanding and managing chronic pain. *J. Rheumatol.* **51**, 1235–1240 (2024).
70. Fauchon, C. et al. Sex differences in brain modular organization in chronic pain. *Pain* **162**, 1188–1200 (2021).

71. Rusman, T., van Vollenhoven, R. F. & van der Horst-Bruinsma, I. E. Gender differences in axial spondyloarthritis: women are not so lucky. *Curr. Rheumatol. Rep.* **20**, 35 (2018).
72. Smith, Y. R. et al. Pronociceptive and antinociceptive effects of estradiol through endogenous opioid neurotransmission in women. *J. Neurosci.* **26**, 5777–5785 (2006).
73. Vincent, K. et al. Brain imaging reveals that engagement of descending inhibitory pain pathways in healthy women in a low endogenous estradiol state varies with testosterone. *Pain* **154**, 515–524 (2013).
74. Ibanez Vodnizza, S. et al. Muscle wasting in male TNF- α blocker naive ankylosing spondylitis patients: a comparison of gender differences in body composition. *Rheumatology* **56**, 1566–1572 (2017).
75. Gao, A. et al. Sex-related differences in efficacy and safety outcomes in axial spondyloarthritis randomized clinical trials: a systematic literature review and meta-analysis. *Arthritis Care Res.* **77**, 813–826 (2025).
76. Garrett, S. et al. A new approach to defining disease status in ankylosing spondylitis: the Bath Ankylosing Spondylitis Disease Activity Index. *J. Rheumatol.* **21**, 2286–2291 (1994).
77. Machado, P. et al. Ankylosing spondylitis disease activity score (ASDAS): defining cut-off values for disease activity states and improvement scores. *Ann. Rheum. Dis.* **70**, 47–53 (2010).
78. Baraliakos, X. et al. Patients with fibromyalgia rarely fulfil classification criteria for axial spondyloarthritis. *Rheumatology* **57**, 1541–1547 (2018).
79. Freynhagen, R., Baron, R., Gockel, U. & Tolle, T. R. painDETECT: a new screening questionnaire to identify neuropathic components in patients with back pain. *Curr. Med. Res. Opin.* **22**, 1911–1920 (2006).
80. Bouhassira, D. et al. Development and validation of the neuropathic pain symptom inventory. *Pain* **108**, 248–257 (2004).
81. Melzack, R. The McGill Pain Questionnaire: major properties and scoring methods. *Pain* **1**, 277–299 (1975).
82. Mayer, T. G. et al. The development and psychometric validation of the central sensitization inventory. *Pain. Pract.* **12**, 276–285 (2012).
83. Cleeland, C. S. & Ryan, K. M. Pain assessment: global use of the Brief Pain Inventory. *Ann. Acad. Med. Singap.* **23**, 129–138 (1994).
84. Mease, P., Reed, G., Ogdie, A., Pappas, D. A. & Kremer, J. M. Prevalence of fibromyalgia and widespread pain in psoriatic arthritis: association with disease severity assessment in a large US registry. *Arthritis Care Res.* **76**, 1313–1321 (2024).
85. Chaabo, K. et al. Pain sensitisation and joint inflammation in patients with active rheumatoid arthritis. *RMD Open* **10**, e003784 (2024).
86. Kaplan, C. M. et al. Association of inflammation with pronociceptive brain connections in rheumatoid arthritis patients with concomitant fibromyalgia. *Arthritis Rheumatol.* **72**, 41–46 (2020).
87. Vanderwall, A. G. & Milligan, E. D. Cytokines in pain: harnessing endogenous anti-inflammatory signaling for improved pain management. *Front. Immunol.* **10**, 3009 (2019).
88. Kapas, L., Hansen, M. K., Chang, H. Y. & Krueger, J. M. Vagotomy attenuates but does not prevent the somnogenic and febrile effects of lipopolysaccharide in rats. *Am. J. Physiol.* **274**, R406–R411 (1998).
89. Dantzer, R., O'Connor, J. C., Freund, G. G., Johnson, R. W. & Kelley, K. W. From inflammation to sickness and depression: when the immune system subjugates the brain. *Nat. Rev. Neurosci.* **9**, 46–56 (2008).
90. Erickson, M. A. et al. Genetics and sex influence peripheral and central innate immune responses and blood-brain barrier integrity. *PLoS ONE* **13**, e0205769 (2018).
91. Sunzini, F., Schrepf, A., Clauw, D. J. & Basu, N. The biology of pain: through the rheumatology lens. *Arthritis Rheumatol.* **75**, 650–660 (2023).
92. Ramiro, S. et al. ASAS-EULAR recommendations for the management of axial spondyloarthritis: 2022 update. *Ann. Rheum. Dis.* **82**, 19–34 (2023).
93. Ortolan, A. et al. Efficacy and safety of non-pharmacological and non-biological interventions: a systematic literature review informing the 2022 update of the ASAS/EULAR recommendations for the management of axial spondyloarthritis. *Ann. Rheum. Dis.* **82**, 142–152 (2023).
94. Roberts, M. J., Hamrouni, M., Linsley, V., Moorthy, A. & Bishop, N. C. Exercise as an anti-inflammatory therapy in Axial Spondyloarthritis Therapeutic Intervention (EXTASI) study: a randomized controlled trial. *Rheumatol. Adv. Pract.* **8**, rkae062 (2024).
95. Hu, X. et al. Effects of exercise programmes on pain, disease activity and function in ankylosing spondylitis: a meta-analysis of randomized controlled trials. *Eur. J. Clin. Invest.* **50**, e13352 (2020).
96. Baraliakos, X., Bergmann, E., Tsiang, S., Redeker, I. & Braun, J. A good response to nonsteroidal antiinflammatory drugs does not discriminate patients with longstanding axial spondyloarthritis from controls with chronic back pain. *J. Rheumatol.* **51**, 250–256 (2024).
97. Webers, C. et al. Efficacy and safety of biological DMARDs: a systematic literature review informing the 2022 update of the ASAS-EULAR recommendations for the management of axial spondyloarthritis. *Ann. Rheum. Dis.* **82**, 130–141 (2023).
98. Navarro-Compan, V., Sepriano, A., Capelusnik, D. & Baraliakos, X. Axial spondyloarthritis. *Lancet* **405**, 159–172 (2025).
99. Trouvin, A. P. et al. Changes in descending pain modulation during anti-tumor necrosis factor therapy: a prospective study in rheumatoid arthritis and spondyloarthritis. *Arthritis Rheumatol.* **77**, 658–663 (2025).
100. Navarro-Compan, V. et al. Effect of tofacitinib on pain, fatigue, health-related quality of life and work productivity in patients with active ankylosing spondylitis: results from a phase III, randomised, double-blind, placebo-controlled trial. *RMD Open* **8**, e002253 (2022).
101. Navarro-Compan, V. et al. Effect of upadacitinib on disease activity, pain, fatigue, function, health-related quality of life and work productivity for biologic refractory ankylosing spondylitis. *Rheumatol. Ther.* **10**, 679–691 (2023).
102. Navarro-Compan, V. et al. Time to improvement of pain, morning stiffness, fatigue, and disease activity in patients with ankylosing spondylitis treated with tofacitinib: a post hoc analysis. *Arthritis Res. Ther.* **26**, 105 (2024).
103. Machado, G. C. et al. Efficacy and safety of paracetamol for spinal pain and osteoarthritis: systematic review and meta-analysis of randomised placebo controlled trials. *BMJ* **350**, h1225 (2015).
104. Lopez-Medina, C. & Molto, A. Comorbid pain in axial spondyloarthritis, including fibromyalgia. *Ther. Adv. Musculoskelet. Dis.* **12**, 1759720X20966123 (2020).
105. Cerasoli, T. et al. Injective therapies for managing sacroiliac joint pain in spondyloarthropathy: a systematic review and meta-analysis. *J. Clin. Med.* **14**, 1294 (2025).
106. van Wissen, M. A. T. et al. One-year effectiveness of long-term exercise therapy in people with axial spondyloarthritis and severe functional limitations. *Rheumatology* **64**, 1817–1825 (2025).
107. Ward, M. M. et al. 2019 Update of the American College of Rheumatology/Spondylitis Association of America/Spondyloarthritis Research and Treatment Network Recommendations for the treatment of ankylosing spondylitis and nonradiographic axial spondyloarthritis. *Arthritis Rheumatol.* **71**, 1599–1613 (2019).
108. Garrido-Cumbrera, M. et al. Factors associated with poor mental health in patients with axial spondyloarthritis: results from the International Map of Axial Spondyloarthritis (IMAS). *RMD Open* **10**, e004218 (2024).
109. Veehof, M. M., Trompetter, H. R., Bohlmeijer, E. T. & Schreurs, K. M. Acceptance- and mindfulness-based interventions for the treatment of chronic pain: a meta-analytic review. *Cogn. Behav. Ther.* **45**, 5–31 (2016).
110. Feliu-Soler, A. et al. Current status of acceptance and commitment therapy for chronic pain: a narrative review. *J. Pain. Res.* **11**, 2145–2159 (2018).
111. Sharpe, L. Psychosocial management of chronic pain in patients with rheumatoid arthritis: challenges and solutions. *J. Pain. Res.* **9**, 137–146 (2016).
112. Fisher, E. et al. Psychological therapies for the management of chronic and recurrent pain in children and adolescents. *Cochrane Database Syst. Rev.* **9**, CD003968 (2018).
113. Bidonde, J. et al. Effectiveness of non-pharmacological interventions for fibromyalgia and quality of review methods: an overview of Cochrane Reviews. *Semin. Arthritis Rheum.* **63**, 152248 (2023).
114. Macfarlane, G. J. et al. EULAR revised recommendations for the management of fibromyalgia. *Ann. Rheum. Dis.* **76**, 318–328 (2017).
115. Malik, A. et al. Serotonin and norepinephrine reuptake inhibitors (SNRIs) for treatment of fibromyalgia: a comprehensive clinical review. *Psychopharmacol. Bull.* **55**, 24–40 (2025).
116. Moldofsky, H., Harris, H. W., Archambault, W. T., Kwong, T. & Lederman, S. Effects of bedtime very low dose cyclobenzaprine on symptoms and sleep physiology in patients with fibromyalgia syndrome: a double-blind randomized placebo-controlled study. *J. Rheumatol.* **38**, 2653–2663 (2011).
117. Lederman, S., Arnold, L. M., Vaughn, B., Kelley, M. & Sullivan, G. M. Efficacy and safety of sublingual cyclobenzaprine for the treatment of fibromyalgia: results from a randomized, double-blind, placebo-controlled trial. *Arthritis Care Res.* **75**, 2359–2368 (2023).
118. Hauser, W., Bernady, K., Uceyler, N. & Sommer, C. Treatment of fibromyalgia syndrome with gabapentin and pregabalin — a meta-analysis of randomized controlled trials. *Pain* **145**, 69–81 (2009).
119. Cohen, S. P. & Hooten, W. M. Advances in the diagnosis and management of neck pain. *BMJ* **358**, j3221 (2017).
120. Staud, M. D. et al. Current strategies for the management of painful diabetic neuropathy. *J. Diabetes Sci. Technol.* **16**, 341–352 (2022).
121. Kontoangelos, K. A. et al. Pregabalin for opioid-refractory pain in a patient with ankylosing spondylitis. *Case Rep. Psychiatry* **2013**, 912409 (2013).
122. Onutu, A. H. Duloxetine, an antidepressant with analgesic properties — a preliminary analysis. *Rom. J. Anaesth. Intensive Care* **22**, 123–128 (2015).
123. Ayub, S. et al. Non-opioid psychiatric medications for chronic pain: systematic review and meta-analysis. *Front. Pain. Res.* **5**, 1398442 (2024).
124. Borton, Z. M., Oakley, B. J., Clamp, J. A., Birch, N. C. & Bateman, A. H. Cervical transforaminal epidural steroid injections for radicular pain: a systematic review. *Bone Jt J.* **104-B**, 567–574 (2022).
125. Manchikanti, L. et al. Equivalent outcomes of lumbar therapeutic facet joint nerve blocks and radiofrequency neurotomy: comparative evaluation of clinical outcomes and cost utility. *Pain. Physician* **25**, 179–192 (2022).
126. Hofmeister, M. et al. Effectiveness of neurostimulation technologies for the management of chronic pain: a systematic review. *Neuromodulation* **23**, 150–157 (2020).
127. Poddubnyy, D. et al. The Assessment of SpondyloArthritis International Society (ASAS) consensus-based expert definition of difficult-to-manage, including treatment-refractory, axial spondyloarthritis. *Ann. Rheum. Dis.* **84**, 538–546 (2025).

Author contributions

The authors contributed equally to all aspects of the article.

Competing interests

X.B. has consulted for and/or participated in scientific advisory boards for AbbVie, Advanz, Alexion, Alphasigma, Amgen, AstraZeneca, BMS, CESAS, Celltrion, Clarivate, Dr. Reddy's, Galapagos, Greywolfe, Globemed, Johnson and Johnson, Lilly, Moonlake, Novartis, Peervoice, Pfizer, Roche, Sandoz, Springer, STADA, Takeda, UCB and Zuellig; has received research grants from AbbVie, Celltrion, Janssen, Moonlake and Novartis; is EULAR president and past president of ASAS. V.N.-C. has consulted for and/or participated in scientific advisory boards for AbbVie, Alfasigma, BMS, Galapagos, Eli Lilly, Johnson and Johnson, Moonlake, Novartis, Pfizer, Roche and UCB; has received research grants from AbbVie and Novartis; is also the president of ASAS. N.Z. has received speaker fees from Roche, AbbVie, Eli Lilly, Pfizer, Janssen, Novartis, Pierre Fabre, Apotex and Pharmaline; has participated in advisory boards for Roche, AbbVie, Eli Lilly, Pfizer, Janssen, Novartis, Gilead and NewBridge; has received research grants from AbbVie, NewBridge, Pfizer and Algorithm/Celgene. D.P. has received research support from AbbVie, Eli Lilly, Johnson and Johnson, Novartis, Pfizer and UCB; has received consulting fees from AbbVie, Eli Lilly, GlaxoSmithKline, Greywolf Therapeutics, Johnson and Johnson, Merk, Moonlake, Novartis, Pfizer and UCB; has received speaker fees from AbbVie, Canon, Celltrion, Eli Lilly, Globemed, Johnson and Johnson,

Medscape, Novartis, Peervoice, Pfizer and UCB; serves as a member of the executive committee of ASAS and is a member of the steering committee of GRAPPA.

Additional information

Peer review information *Nature Reviews Rheumatology* thanks Francesco Ciccia and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

© Springer Nature Limited 2026, modified publication 2026

Cam morphology and the development of femoroacetabular impingement syndrome and hip osteoarthritis

Joshua Heerey¹✉, Joanne Kemp¹, Pim van Klij², Vasco Mascarenhas³, Mark Scholes¹, Fleur Boel⁴, Paul Dijkstra^{5,6}, Kay Crossley¹, Sita Bierma-Zeinstra^{4,7} & Rintje Agricola⁴

Abstract

Hip morphology has emerged as an important factor in the development of hip osteoarthritis (OA). Cam morphology is one of the most common hip morphologies, characterized by a bony prominence around the femoral head–neck junction of the hip that alters the normal shape of the femoral head. Cam morphology can contribute to intra-articular joint damage by generating abnormal contact stresses at this junction, initiating femoroacetabular impingement (FAI) syndrome and eventually leading to hip OA. Cam morphology is a causal risk factor for hip OA, but not everybody with this morphology will develop FAI syndrome or OA. The pathogenesis of hip disease is probably driven by the interplay between cam morphology, other coexisting hip morphologies (such as pincer morphology), femoral version, spinopelvic parameters and biomechanical and environmental factors. Early identification of FAI syndrome could enable timely, multidisciplinary intervention and offers the potential to modify the trajectory of disease. Cam morphology can develop during skeletal maturation, particularly in adolescents participating in high-joint-load physical activity, raising important questions about preventative approaches. Management of FAI syndrome includes both surgical and non-surgical approaches. Emerging insights into the pathogenesis and detection of cam morphology are paving the way for more targeted interventions and a deeper understanding of its role in FAI syndrome and hip OA development.

Sections

Introduction

Clinical definitions and terminology

Diagnosis and classification

Pathogenesis

Progression to hip OA

Management

Conclusion

¹La Trobe Sport and Exercise Medicine Research Centre, School of Allied Health, Human Services and Sport, La Trobe University, Melbourne, Victoria, Australia. ²Department of Orthopaedic Surgery and Sports Medicine, Amsterdam UMC, Amsterdam, The Netherlands. ³Musculoskeletal Imaging Unit, Hospital da Luz, Lisbon, Portugal. ⁴Department of Orthopaedics and Sports Medicine, Erasmus MC University Medical Centre, Rotterdam, The Netherlands. ⁵Department of Medical Education, Aspetar Orthopaedic and Sports Medicine Hospital, Doha, Qatar. ⁶Weill Cornell Medicine – Qatar, Qatar Foundation, Doha, Qatar. ⁷Department of General Practice, Erasmus MC University Medical Center, Rotterdam, The Netherlands. ✉e-mail: j.heerey@latrobe.edu.au

Key points

- Cam morphology is a causal risk factor for hip osteoarthritis (OA) and is characterized by excess bone at the anterolateral head–neck junction of the proximal femur.
- Cam morphology probably develops during adolescence as a physiological response to high-joint-load activity, and can lead to femoroacetabular impingement (FAI) syndrome, a symptomatic, motion-related condition linked to hip OA development.
- Specific subtypes of cam morphology and individual characteristics could expedite the development of OA; pincer morphology in the presence of hip pain also seems to be relevant to hip OA development.
- Radiography is the first-line imaging technique for assessing cam and pincer morphology in FAI syndrome, with magnetic resonance imaging providing complementary detail on hip anatomy and intra-articular soft tissues.
- The pathogenesis of FAI syndrome probably results from an imbalance between biological, physical and environmental factors.
- Surgical and non-surgical treatments could be appropriate for people with FAI syndrome.

Introduction

An estimated one in four people will develop hip osteoarthritis (OA) in their lifetime¹. Hip OA increases the risk of other chronic diseases (such as cardiovascular disease) and imposes a substantial societal and personal burden, including poor quality of life, persistent pain and disability^{1–3}. Structural variations in hip morphology are one of the most important risk factors in the development of hip OA^{4–7}. Changes in hip morphology are thought to induce pathological tissue forces within the hip joint that can generate clinical symptoms and expedite OA^{6,8–10}. Historically, clinical interest has focused on conditions that produce distinct changes to hip morphology, such as hip dysplasia; however, cam morphology (characterized by the presence of additional bone on the anterolateral head–neck junction of the proximal femur) has now been studied in greater detail and seems to have a central causal role in OA pathogenesis^{5,8,9,11,12}.

Cam morphology can impinge against the acetabulum in positions of hip flexion and internal rotation, causing changes to the articular cartilage and acetabular labrum^{7,10,13,14}. However, emerging evidence suggest that pincer morphology, femoral version and spinopelvic alignment also have key roles in modulating the interaction between cam morphology and intra-articular structures^{8,15–18}. In some people, this mechanical process can cause femoroacetabular impingement (FAI) syndrome, a symptomatic motion-related condition that can occur in young to middle-aged active adults^{10,13}. Over time, this abnormal joint contact might contribute to the development of OA. Although this mechanical pathway towards hip OA might hold true in many individuals, cam morphology is also present in individuals without pain and a large proportion of these people will remain asymptomatic and/or will not develop OA^{19–24}. Given that FAI syndrome contributes to the development of hip OA in older adults, early detection in clinical settings could support collaborative management and help to

modify disease progression. This Review offers rheumatologists an in-depth update on cam morphology and FAI syndrome, concentrating on three key areas. First, we outline the recommended definitions and terminology for cam morphology and FAI syndrome. Second, we highlight advances in understanding of cam morphology and FAI syndrome, including diagnostic approaches, insights into its pathogenesis (including the involvement of pincer morphology, femoral version and spinopelvic parameters) and its role in the development of hip OA. Third, we summarize the latest clinical trial findings that inform current approaches to the management of FAI syndrome and FAI syndrome with hip OA, as well as potential opportunities for cam morphology prevention.

Clinical definitions and terminology

Consistent use of recommended definitions and terminology improves communication between patients and clinicians, enhancing understanding and contributing to a more effective and positive healthcare experience²⁵. In this section, we summarize current definitions and terminology related to cam morphology, pincer morphology and FAI syndrome.

Cam morphology

As our understanding of the contribution of cam morphology to FAI syndrome and hip OA has evolved, so have the associated definitions and recommended terminology^{25–27} (Box 1). Cam morphology is defined, according to a concept analysis²⁶ and the Oxford Consensus study²⁵, as a cartilage or bony prominence (bump) of varying size at any location around the femoral head–neck junction of the hip that changes the shape of the femoral head from spherical to aspherical. Various forms or subtypes of cam morphology probably exist²⁸. These variants of cam morphology may differ substantially in shape and the extent to which each is associated with FAI syndrome and/or OA²⁸. An important step forwards in the understanding of cam morphology has been the distinction between primary and secondary forms. Primary cam morphology (the focus of this Review article and hereinafter referred to as cam morphology) develops during skeletal maturation in young adolescents (with no current or previous hip disease), as a normal physiological response to high-joint-load sporting activity and other unconfirmed risk factors^{25,26,29–32}. By contrast, secondary cam morphology arises as a consequence of pre-existing hip disease or acute trauma, including Legg–Calvé–Perthes disease, slipped capital femoral epiphysis and healed proximal femoral fractures²⁵. Distinguishing between these two subtypes of cam morphology could be important for informing prognosis, guiding treatment decisions and facilitating patient discussions about clinical relevance^{25,27}.

Pincer morphology

Pincer morphology has yet to be formally defined by consensus studies, but generally refers to focal or global overcoverage of the femoral head by the acetabulum^{10,33}. Pincer morphology can result from increased acetabular coverage and/or acetabular retroversion³³ (Fig. 1). As part of its effort to standardize the diagnosis and management of FAI, the Warwick Agreement on FAI syndrome provided recommended terminology for describing pincer morphology¹⁰ (Box 1).

FAI syndrome

Femoroacetabular impingement syndrome is now the widely accepted term to describe a clinical disorder present in young to middle-aged adults (Box 1). The Warwick Agreement defined FAI syndrome as a

Box 1 | Evidence-informed terminology for cam and pincer morphology and FAI syndrome

The terminology surrounding cam and pincer morphology, as well as femoroacetabular impingement (FAI) syndrome, has varied considerably. For cam and pincer morphology, terms such as ‘lesion’, ‘abnormality’ or ‘deformity’ are frequently used^{10,26}. However, these terms imply pathological change, which is inappropriate given that cam morphology is also present in asymptomatic individuals^{19,23,24,26} and probably reflects a physiological adaptation to high-joint-load activity during adolescence^{10,25,27}. Pincer morphology should be considered similarly, although its developmental origins are less well understood. The term ‘FAI syndrome’ emphasizes the role of symptoms in diagnosis to avoid confusion associated with previous terminology, such as ‘asymptomatic FAI’. Evidence-informed terminology for describing cam and pincer morphology, and FAI syndrome, are outlined below.

Recommended terminology:

- Cam morphology
- Pincer morphology

- FAI syndrome
- FAI syndrome with cam morphology (when describing the clinical disorder with cam morphology)
- FAI syndrome with pincer morphology (when describing the clinical disorder with pincer morphology)
- FAI syndrome with mixed morphology (when describing the clinical disorder with cam morphology and pincer morphology)

Terminology to avoid:

- Symptomatic femoroacetabular impingement
- Asymptomatic femoroacetabular impingement
- Cam-type morphology
- Pincer-type morphology
- Cam-type femoroacetabular impingement
- Femoroacetabular impingement morphology
- Deformity, lesion, abnormality or pathology when referring to cam or pincer morphology

motion-related clinical disorder of the hip with a triad of symptoms, clinical signs and imaging findings. It represents symptomatic premature contact between the proximal femur and the acetabulum¹⁰. FAI syndrome is classified into three subtypes: cam morphology, pincer morphology and mixed morphology. Of these subtypes, FAI syndrome with cam morphology is considered to be the most prevalent^{16,24,25,34}. Distinguishing those with FAI syndrome from individuals with either cam or pincer morphology but without symptoms is essential to avoid misdiagnosis and inform treatment. Femoral version and spinopelvic parameters can interact with cam morphology and/or pincer morphology and contribute to the pathogenesis of FAI syndrome. The anatomical, radiological and clinical characteristics of each of these factors, as well as their contribution to FAI syndrome and hip OA, are detailed in the following sections wherever we considered it relevant to the rheumatologist.

Diagnosis and classification

Diagnosis serves as the foundation of clinical care, integrating clinical signs, symptoms and investigations to determine the cause of disease³⁵. Conversely, classification is the process of using standardized criteria to create homogeneous patient populations for clinical research^{33,36,37}. In this section we discuss the available diagnostic and classification criteria for cam morphology and FAI syndrome, as well as pincer morphology.

Cam morphology

The alpha angle is the most frequently used radiologic metric for assessing cam morphology and is defined as the angle between the femoral neck axis and a line drawn from the centre of the femoral head to the point where the contour diverges from a perfect circle^{25,26,38,39} (Fig. 1). This metric quantifies the sphericity of the femoral head–neck junction at specific locations and is widely applicable across various imaging modalities and views^{25,38}. Evidence supports a non-sex-specific alpha angle threshold of 60° (at any location around the anterosuperior femoral head–neck junction) (Fig. 2) as a useful discriminator

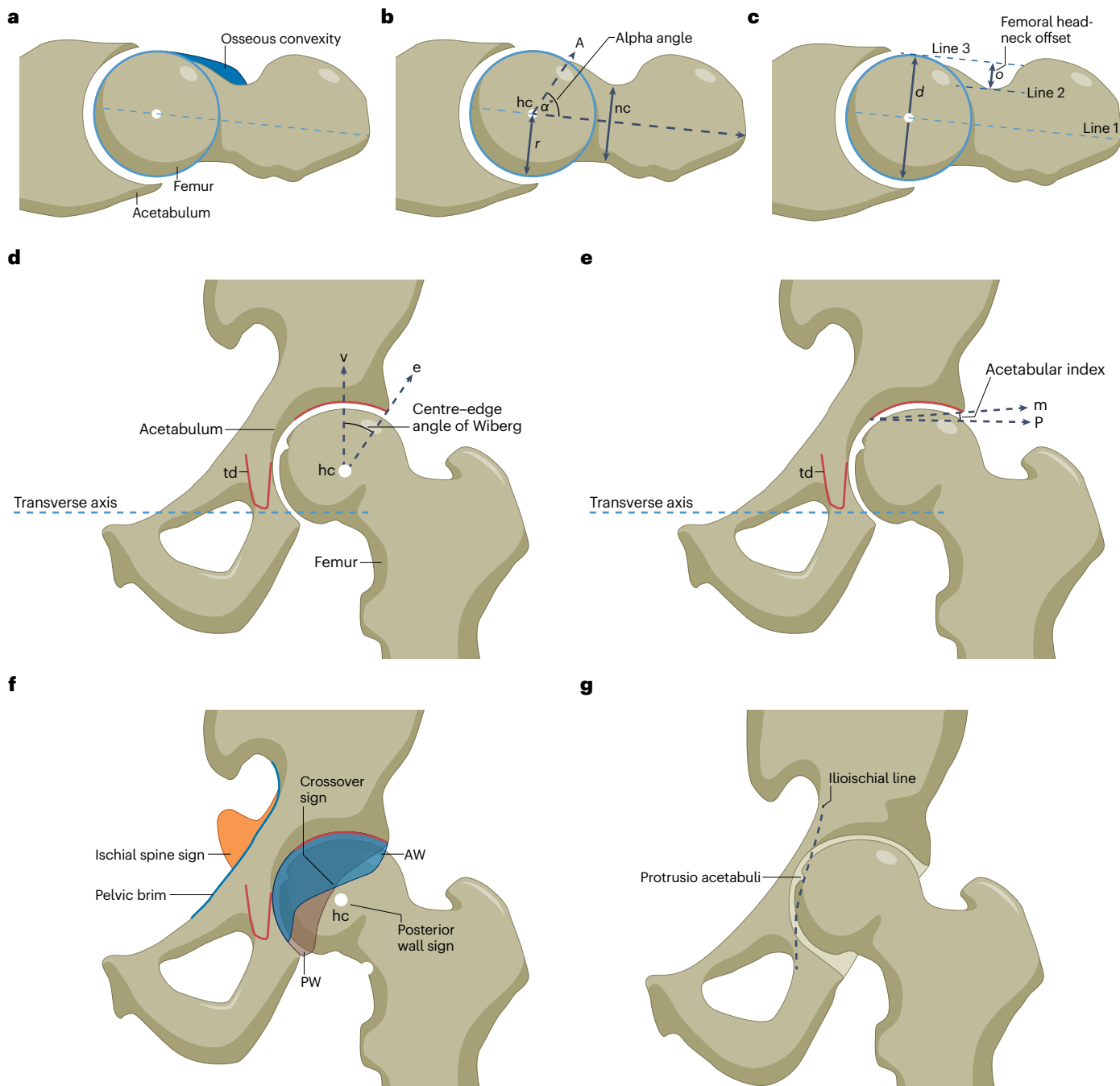
between people with FAI syndrome and pain-free individuals; however, some overlap between these groups still remains around this threshold^{16,38,40}. Magnetic resonance imaging (MRI)-based studies have identified additional threshold values for specific locations on the femoral neck, with an alpha angle of 57–60° at the 1:30–2:00 position on the femoral head–neck junction, showing a strong association with symptoms¹⁶.

Pincer morphology

Researchers have proposed various radiologic measures and diagnostic criteria for identifying pincer morphology, but these approaches often produce inconsistent and conflicting findings^{33,37,38}. Evaluating acetabular coverage and version is essential, as variations in either can result in pincer morphology^{33,37,38}. The centre–edge angle of Wiberg (W-CEA) and the acetabular index are the most used radiologic measures for evaluating the superolateral coverage of the acetabulum^{33,37,38} (Figs. 1 and 2). The presence of protrusio acetabuli should also be noted owing to its association with OA⁴¹ (Fig. 1). When evaluating acetabular version, the crossover sign, ischial spine sign and posterior wall sign (Figs. 1 and 2 and Supplementary Table 1) can help to determine the presence of acetabular retroversion (either focal or global)^{33,37,38}.

FAI syndrome

The diagnosis of FAI syndrome requires a comprehensive approach that integrates both clinical evaluation (Box 2) and imaging assessment (Box 3). Pain is the predominant symptom of FAI syndrome and is often exacerbated with repetitive activity or positions. The location, anatomical spread and severity of pain varies widely, with people often reporting groin pain, but also discomfort in the lateral hip, anterior or posterior thigh, buttock or lower back^{10,13}. Mechanical symptoms (clicking, catching and locking) can also be present and often coexist with pain. In symptomatic people presenting with clinical findings, imaging has a pivotal role in the diagnostic work-up for rheumatologists. When cam morphology is used as a diagnostic criterion for FAI syndrome, additional variables must be considered^{16,33,37}. For example,



some people exhibit impingement symptoms with an alpha angle below 60° , as this simple radiographic measure might not capture the complexity of FAI syndrome^{16,38,42}. A comprehensive assessment should therefore include other anatomical and functional variations – such as pincer morphology, femoral version and spinopelvic parameters, sex and athletic performance – to better understand the diverse clinical phenotypes of FAI syndrome^{10,13,42,43}.

Classification criteria for clinical research

In the past few years, The Lisbon Agreement has provided imaging-based criteria for classifying cam and pincer morphology (irrespective of

symptoms)^{33,36,37}. For cam morphology, recommended parameters included the alpha angle, femoral head-neck (FHN) offset, the FHN offset ratio and the osseous convexity of the FHN junction³³ (Fig. 1 and Supplementary Table 1). For pincer morphology, key indicators included the W-CEA, acetabular index, protrusio acetabuli and three measures of retroversion (crossover, ischial spine and posterior wall sign)³³.

Pathogenesis

The formation of cam morphology, and in some cases the subsequent onset of FAI syndrome, are recognized as critical stages within the disease continuum of hip OA^{6,40,44}. Understanding the mechanisms

Fig. 1 | Recommended imaging measures for classifying cam and pincer morphology. This figure illustrates key radiographic measures used to identify hip morphology associated with femoroacetabular impingement syndrome, including cam and pincer morphologies. **a**, Osseous convexity is identified by a visible bony prominence at the femoral head–neck junction. **b**, The alpha angle α is measured at the anterior point (A) where the contour of the femoral head exceeds its radius r . The angle is formed between the centre of the femoral head (hc), the point of deviation and the centre of the femoral neck at its narrowest point (nc). **c**, The femoral head–neck offset o is defined as the distance (in millimetres) between the anterior cortex of the femoral neck (line 2) and the outer edge of the femoral head (line 3), measured parallel to the neck (line 1). The offset ratio is calculated by dividing this distance o by the diameter d of the femoral head. **d**, The centre–edge angle of Wiberg is the angle between a vertical line (v) drawn from the centre of the femoral head (hc) and a line extending to the lateral edge of the acetabular sourcil (e), and is used to assess acetabular

coverage. This vertical reference line v is perpendicular to the transverse axis, which is established by connecting the acetabular tear drops (td). **e**, The acetabular index is measured as the angle between a line (m) drawn from the medial aspect of the sclerotic sourcil to the lateral edge of the acetabular weight-bearing surface and a reference line (p) drawn parallel to the transverse axis from the medial aspect of the acetabular weight-bearing surface. **f**, Crossover sign, ischial spine sign and posterior wall sign are indicators of acetabular retroversion. Crossover sign is present when the anterior acetabular wall (AW) crosses over and becomes lateral to the posterior wall (PW). Posterior wall sign is present when the posterior wall lies medial to the centre of the femoral head (hc). Ischial spine sign is present when the triangular shape of the ischial spine protrudes and is visible medially to the pelvic brim. **g**, Protrusio acetabuli is considered to be present when the femoral head crosses or touches the ilioischial line. Parts **d–g** adapted with permission from ref. 38, Thieme Medical Publishers.

underlying their formation is essential for identifying opportunities for early intervention and disease modification.

Formation of cam morphology

Existing evidence indicates that cam morphology develops during adolescence through an interplay between femoral growth plate physiology and mechanical loading of the hip joint (such as that caused by athletic activity) that ultimately result in additional bone formation around the femoral head–neck junction^{25,26,29,30,32}. Typical high-joint-load activities associated with cam morphology formation include football, ice hockey and basketball⁴⁵. Bone and cartilage are highly responsive to mechanical and physiological stimuli during adolescence, but this adaptive capacity diminishes after growth-plate closure, essentially limiting cam morphology development to the period of active skeletal growth^{26,30,32}.

Despite considerable advances in understanding, the exact tissue-based mechanism underpinning cam morphology formation remains incompletely understood. Longitudinal studies of physically active, mainly male adolescents have identified increased epiphyseal extension and hypertrophy of the fibrochondro-osseous tissue area at the femoral head–neck junction as important mechanisms in cam morphology development^{30,32,46} (Fig. 3). However, the exact contribution of each mechanism differs between studies and populations, and might not be true for cam morphology development in female adolescents^{30,32,46}. The developmental time frame for cam morphology is currently best described in male athletes^{29–32,46} (Fig. 3). From birth until approximately 8 years of age, the proximal femur typically develops in a physiological manner. Between ages 8 and 10 years, the first signs of soft-tissue hypertrophy arise, primarily of fibrochondro-osseous origin and typically located at the lateral head–neck junction of the femur²⁹. Around ages 9 to 10 years, the femoral epiphysis begins to assume an increasingly oblique orientation with progressive lateral extension. From ages 9 to 12 years – depending on sex and occurring just before the second growth spurt – the first osseous changes emerge, marking the earliest visible signs of cam morphology formation^{30,31}. At this stage, the anterolateral head–neck junction gradually begins to lose its spherical contour and develops a subtle bony asphericity. As growth-plate closure approaches, the capacity for further osseous remodelling declines³⁰. After epiphyseal closure (around age 13 years in women and between ages 13 and 18 years in men), the skeleton loses the capacity for further cam morphology development^{30,32,47}. Epiphyseal closure can occur earlier in some

individuals. The lower prevalence of cam morphology in women might partly reflect sex-related differences in skeletal maturity during adolescence, as women typically experience earlier femoral growth-plate closure than men. This earlier closure might limit the period of vulnerability during which mechanical loading could induce cam morphology. Notably, the location and size of cam morphology also seems to differ by sex, which might suggest overarching differences in its pathogenesis⁴⁸.

The primary causal risk factor for cam morphology development is environment (that is, engagement in high-joint-load sports during adolescence)^{29–31,49,50}. In male athletes, both the amount and type of sport played during adolescence are linked to an increased risk of developing cam morphology. For example, one study of professional football players identified a dose–response relationship whereby higher activity levels during adolescence increased the odds of developing cam morphology⁵⁰. Male athletes participating in sports that induce repetitive hip internal rotation and flexion (for example, football and ice hockey) seem to be particularly susceptible^{30,32,46,49}. Ethnicity might also influence an individual's risk of developing cam morphology^{20,51–53}. For instance, cam morphology is less common in East Asian populations compared with white or Black populations^{20,51,52}. These ethnic differences suggest that genetic or cultural factors could be important in the development of cam morphology. Growing interest surround the roles of growth hormones, oestrogen and vitamin D in regulating growth-plate closure⁵⁴. Although the interrelationship between hormonal influence, growth-plate closure and cam morphology remains unclear, specific imbalances or variations during adolescence might affect the growth plate's response to mechanical loading, potentially influencing the formation of cam morphology.

Pathogenesis of FAI syndrome

Considerable interest surrounds the timing and mechanisms by which cam morphology contributes to the development of FAI syndrome^{40,44}. The pathogenesis of FAI syndrome probably results from a complex interplay of biological, physical and environmental factors^{40,44}. Distinct clinical phenotypes probably exist, with the relevance of each factor varying between individuals and over time. Pincer morphology also contributes to FAI syndrome pathogenesis by increasing bony coverage of the femoral head^{7,10,15,33}. When combined with cam morphology, the resulting anatomical configuration leads to earlier bony impingement during hip movement¹⁵.

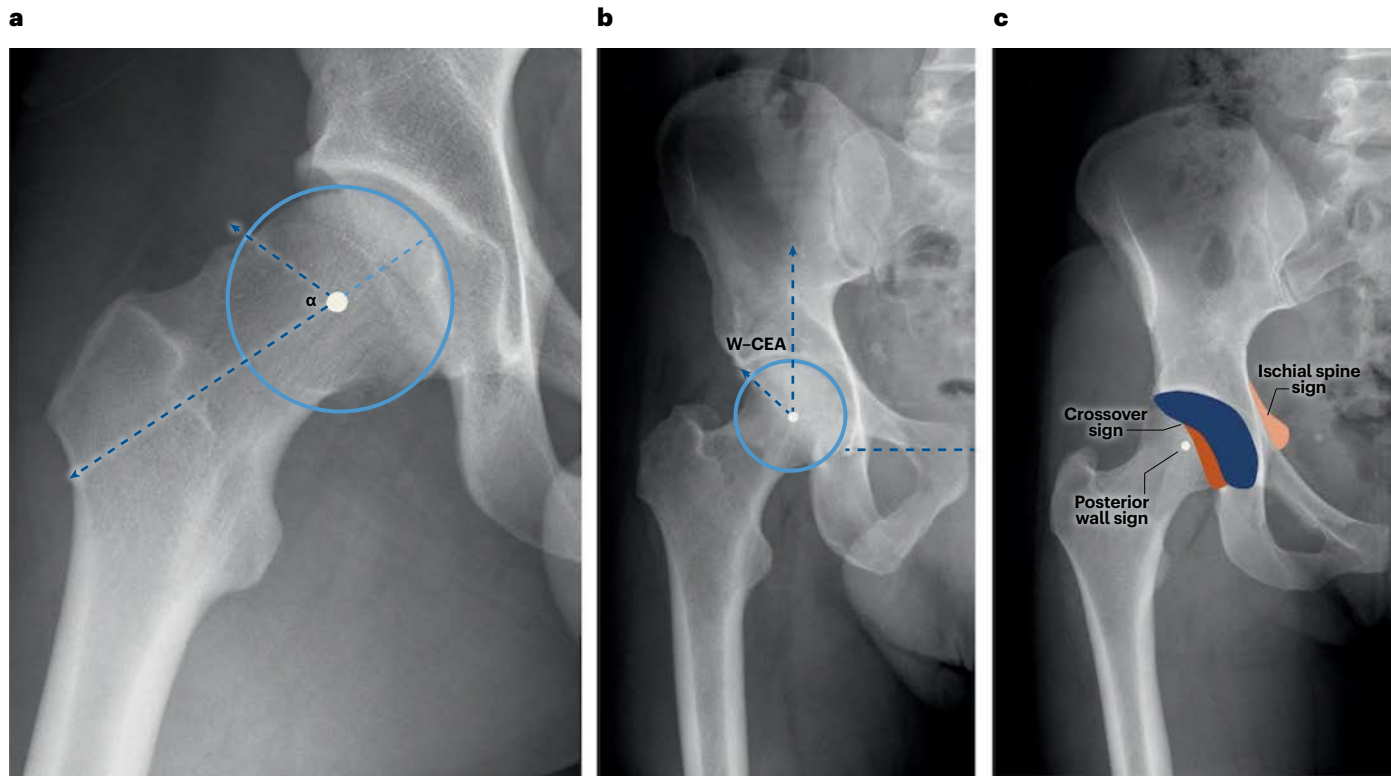


Fig. 2 | Cam morphology and pincer morphology. a. Cam morphology of the right hip, defined by an alpha angle $\alpha \geq 60^\circ$, as visualized on a Dunn 45° lateral radiograph. **b.** Global pincer morphology of the right hip, indicated by a centre-edge angle of Wiberg (W-CEA) $\geq 40^\circ$, on an anteroposterior pelvis radiograph.

c. Global pincer morphology of the right hip, characterized by the presence of crossover sign, posterior wall sign and ischial spine sign, on an anteroposterior pelvis radiograph.

Emerging evidence highlights the notable role of spinopelvic parameters and variations in femoral version in the development of FAI syndrome^{16,17,38,55–57}. The alignment of the spinopelvic complex influences the acetabular orientation and the degree of impingement-free hip motion^{16,58}. Several imaging parameters have been used to understand spinopelvic alignment, including pelvic incidence, sacral slope and pelvic tilt (Supplementary Fig. 1). Higher pelvic incidence^{16,17} and sacral slope¹⁶ (indicating greater lumbopelvic mobility and increased anterosuperior acetabular coverage) seem to predict symptomatic hip status, although findings remain inconsistent across studies⁵⁹. High or low pelvic incidence has been implicated in the development of FAI syndrome, albeit via different mechanisms^{55–57}. For instance, low pelvic incidence (associated with limited lumbopelvic motion) can lead to greater reliance on hip motion during movement and compensatory forward tilt of the pelvis, which can change the functional orientation of the acetabulum and reduce impingement-free hip range motion^{55–57}. When paired with cam and/or pincer morphology, these altered biomechanics might heighten the risk of FAI syndrome. Conversely, those individuals with high pelvic incidence are less reliant on hip motion because they possess greater lumbopelvic mobility^{55–57}. This increase in mobility might induce compensatory backward tilting of the pelvis, resulting in functional undercoverage of the anterior femoral head and increased loading of the anterior joint structures^{55–57}.

Femoral version (the amount of rotation between the proximal and distal aspect of the femur; Supplementary Fig. 1) might also have

an important role in the pathomechanics of FAI syndrome^{18,34,38,60}. Decreased, normal or increased femoral version can be present in hips with cam morphology³⁴. Decreased version (known as retroversion) limits hip internal rotation⁶⁰ and increases the likelihood of both intra-articular and extra-articular impingement during extreme hip flexion compared to hips with FAI syndrome and normal version¹⁸. However, the role of femoral version in early-stage pathology remains uncertain, because lower femoral version is linked to better hip cartilage metabolism⁶¹. Moreover, isolated pathology within pain-generating tissues (such as the acetabular labrum) is unlikely to be the sole mechanism underlying the development of FAI syndrome^{8,62}. For example, one large-scale study identified a similar prevalence, location and severity of labral tears in pain-free athletes with cam morphology and in athletes with FAI syndrome⁸.

Inflammatory and immunologic mechanisms might also have an important role in FAI syndrome pathogenesis, as shown in OA⁶³. Specifically, the synovial tissue of people with FAI syndrome shows elevated levels of synovitis and pro-inflammatory cytokines, which might contribute to symptom development and the degradation of articular cartilage^{61,63}. Furthermore, athletes with FAI syndrome also have elevated levels of C-reactive protein compared with pain-free athletes, suggesting the presence of systematic inflammation⁵¹. However, future exploration is still needed to determine the potential of these and other biomarkers and to identify individuals at risk of FAI syndrome.

Progression to hip OA

Our understanding of the relationship between cam and/or pincer morphology and FAI syndrome continues to evolve, particularly in how these hip morphologies alter hip joint mechanics and accelerate the trajectory of hip OA.

Association of cam and pincer morphology with FAI syndrome

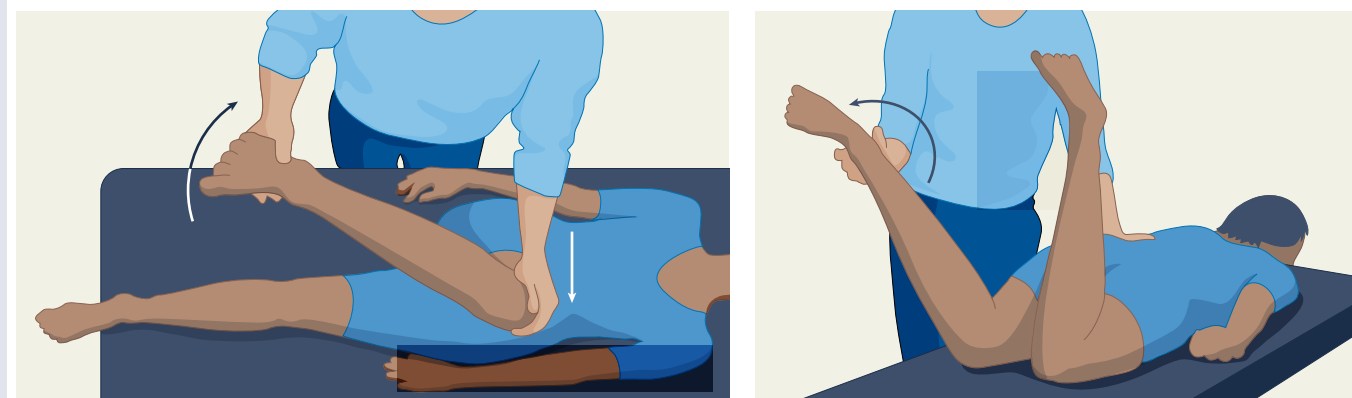
A major limitation of the longitudinal literature is that studies often report on the occurrence of hip pain as a binary variable (that is, present or absent). Hip pain is an umbrella term encompassing various intra-articular conditions, including FAI syndrome, hip dysplasia and labral tears. Longitudinal investigations are also hampered by the slow development of FAI syndrome in those individuals with cam morphology and the difficulty of accurately tracking changes in symptoms over time⁴⁷. The refinement of diagnostic criteria for FAI syndrome (now requiring a triad of symptoms, clinical signs and imaging findings)¹⁰ has also complicated the interpretation of earlier studies that did not use this standardized definition. Finally, most studies investigate hip morphology in isolation rather than the combined effect of femoral, acetabular and spinopelvic anatomy. The following section summarizes evidence linking cam and pincer morphology (and where data permit, femoral version and spinopelvic parameters) to hip pain or FAI syndrome when the appropriate diagnostic criteria have been used.

Longitudinal studies have shown that cam morphology is an inconsistent risk factor for the development of hip pain. A 10-year follow-up of the CHECK study (1,685 hips; mean age 55.9 years) found no association between cam morphology and hip pain in women, and only a weak association in men at the 5-year mark. Cam size also failed to predict symptoms in either sex. However, these findings offer limited insight into younger populations more often affected by FAI syndrome⁶⁴. In a smaller study of 332 asymptomatic hips in younger adults (mean age 29.5 years), MRI-defined cam morphology was associated with an increased risk of developing hip pain within 5 years (relative risk 4.3), although only a small number of hips ($n = 14$) became symptomatic during follow-up⁶⁵. The contralateral pain-free hip of people undergoing hip arthroscopy for ipsilateral FAI syndrome has been used to model the natural history of cam and/or pincer morphology^{66,67}. For example, a prospective investigation (mean follow-up of 7.1 years) found that individuals who later developed hip pain had a decreased FHN offset ratio (indicative of a more pronounced cam morphology) in the contralateral hip; no relationship was observed for imaging measures of pincer morphology and subsequent hip pain⁶⁶. Cross-sectional studies have yielded similarly conflicting results across various populations²⁰. Cam morphology, but not pincer morphology, differentiated people undergoing or seeking hip arthroscopy for FAI syndrome from pain-free individuals^{16,68}. However,

Box 2 | Clinical assessment of FAI syndrome

Clinical tests are often used in the assessment of people suspected of having FAI syndrome^{10,13,112,113}. However, most clinical tests have limited diagnostic accuracy and are best suited for screening or ruling out the condition. Importantly, no single clinical test can confirm a diagnosis of FAI syndrome. The flexion–adduction–internal–rotation (FADIR) test (figure, left) is the most frequently used test in clinical settings; however, although this test has high sensitivity, it lacks specificity and can lead to false-positive findings. For the FADIR test, the symptomatic hip is passively moved to 90° of flexion and the knee is bent to 90° of flexion. From this position, the hip is moved into adduction (avoiding movement of the pelvis) and then internally rotated while maintaining both flexion and adduction. The test is deemed to be positive when it reproduces the pain typically experienced by the patient. Passive joint range of motion should also be assessed, with particular attention to hip internal and external

rotation in both the neutral hip position and at 90° of hip flexion, because these measures might indicate altered acetabular or femoral version¹¹⁴. Notably, reduced hip internal rotation, particularly when assessed in the prone or neutral hip position, is recognized as a useful indicator in the diagnosis of FAI syndrome (figure, right)¹¹⁵. For assessing passive hip internal rotation, the symptomatic hip is moved to a neutral position (with the knee bent to 90° of flexion). From this position, the hip is moved into internal rotation. A positive test is indicated by a limited range of motion, regardless of whether the person's familiar pain is reproduced. A comprehensive clinical assessment should also include evaluation of hip, core and lower-limb muscle strength, movement control and palpation of surrounding anatomical structures (such as the hip adductors, iliopsoas and pubic symphysis) because pain arising from these structures can present similarly to FAI syndrome^{14,113,116}.



Box 3 | Imaging assessment of FAI syndrome

Imaging is pivotal for making informed treatment decisions and determining prognosis in FAI syndrome. Imaging modalities enable detailed characterization of cam and pincer morphology to diagnose the condition, assessment of acetabular and femoral version, evaluation of spinopelvic parameters and identification of labral tears and chondral loss. Imaging also aids in differential diagnoses^{10,33,38,40}. Radiography is the crucial first step in assessing hip anatomy and is often paired with MRI as standard imaging modalities for assessing hip disease associated with cam and/or pincer morphology^{33,38}.

Plain radiographs

The anteroposterior pelvis and lateral views are fundamental to the assessment of cam morphology. The anteroposterior view (figure, panel a) is important for the assessment of pincer morphology (for example, acetabular depth, version and coverage). The lateral view enables assessment of the femoral head–neck junction (best shown with a Dunn 45° view; hips at 45° flexion and 20° abduction (figure, panel b), allowing accurate visualization of cam morphology^{33,36,37}.

However, these two-dimensional radiographic measurements capture only approximately 50% of proximal femur shape variation, often necessitating advanced imaging for comprehensive evaluation of cam morphology^{33,36,37}. Spinopelvic parameters (such as pelvic incidence, sacral slope and pelvic tilt) can be evaluated using lateral lumbosacral radiography³⁸.

Advanced imaging techniques

MRI and computed tomography imaging provide enhanced three-dimensional (3D) assessment of cam and pincer morphology^{36–38}. MRI enables detailed 3D evaluation of hip morphology, including version (acetabular and femoral) and soft tissue structures (for example, cartilage loss and labral tears)³⁸. The Lisbon Agreement categorizes MRI with radial imaging and small field-of-view sequences as the gold standard for non-invasive assessment of cam morphology and associated hip joint soft tissues³⁶. Computed tomography provides high-resolution 3D visualization of bony detail and is mainly used for surgical planning and virtual range-of-motion simulations^{33,36}.

a Anteroposterior view



b Lateral view



neither cam nor pincer morphology distinguished individuals with hip pain from those without hip pain in a large general population sample ($n = 3,202$)⁶⁹ or in a cohort of young adult football players¹⁹. Altered femoral version (whether decreased or increased) exists in both pain-free individuals and individuals with FAI syndrome^{16,34}. By contrast, higher pelvic incidence and sacral slope distinguished individuals with FAI syndrome and cam morphology from pain-free individuals with or without cam morphology¹⁶.

As a three-dimensional structure, variations in cam morphology size, location and magnitude probably also influence its reported relationships with hip pain and FAI syndrome. For example, in one study, larger cam morphology size could discriminate collegiate National Football League draftees⁷⁰ with prior or current hip pain from players without hip pain. Maximum alpha angle measurements are frequently observed in the anterosuperior region of the femoral head–neck junction^{16,37,71,72}, with individuals undergoing surgery for

FAI consistently demonstrating larger anterosuperior values than pain-free individuals^{16,68}. In a separate study of football players with FAI syndrome, larger anterosuperior, but not superior cam morphology, was modestly associated with worse symptom severity⁷³. Finally, greater severity of cam morphology (that is, bony extension over a greater portion of the femoral head–neck junction) has also been associated with FAI syndrome¹⁶. Evidence on the role of cam and pincer morphology in hip pain is conflicting, and longitudinal studies are needed to clarify their contribution (along with femoral version and spinopelvic parameters) to FAI syndrome pathogenesis.

Association of FAI syndrome with hip OA

FAI syndrome has historically been considered a more clinically relevant predictor of hip OA risk than the presence of cam morphology alone. However, until recently little causal evidence has been available to support this perception. A 10-year longitudinal study of older adults whose hips meet the diagnostic criteria for FAI syndrome (defined by the presence of hip pain, restricted internal rotation ($\leq 25^\circ$ in 90° of hip flexion) and cam morphology), reported a markedly increased risk of incident end-stage OA, with an adjusted odds ratio of 47.82 (ref. 12). Although only a small proportion of hips (approximately 1.5%) fulfilled the criteria for FAI syndrome, the strength of the association, along with

the positive and negative predictive values for OA development, was substantially greater than for cam morphology alone. The distinction between cam morphology and FAI syndrome has important clinical implications when considering hip OA risk. Longitudinal investigations of FAI syndrome that incorporate pincer morphology, femoral version or spinopelvic parameters remain scarce, limiting insight into their role in hip OA development.

Association of cam and pincer morphology with hip OA severity

Hip OA is best conceptualized as a disease continuum that commences early in life (often without clinical symptoms), with initial structural changes occurring within the acetabular labrum and articular cartilage^{74–76}. Structural changes can emerge in the absence of radiographic evidence of disease and, although early-stage OA remains undefined for the hip joint, these changes increasingly serve as indicators of early disease development^{74,75,77}. Emerging evidence has provided insight into how cam and pincer morphology contribute to early-stage OA. Cross-sectional findings from a large study of adolescent male football players and a general population comparison group suggest that cam morphology does not affect articular cartilage until later in adolescence when the femoral growth plate

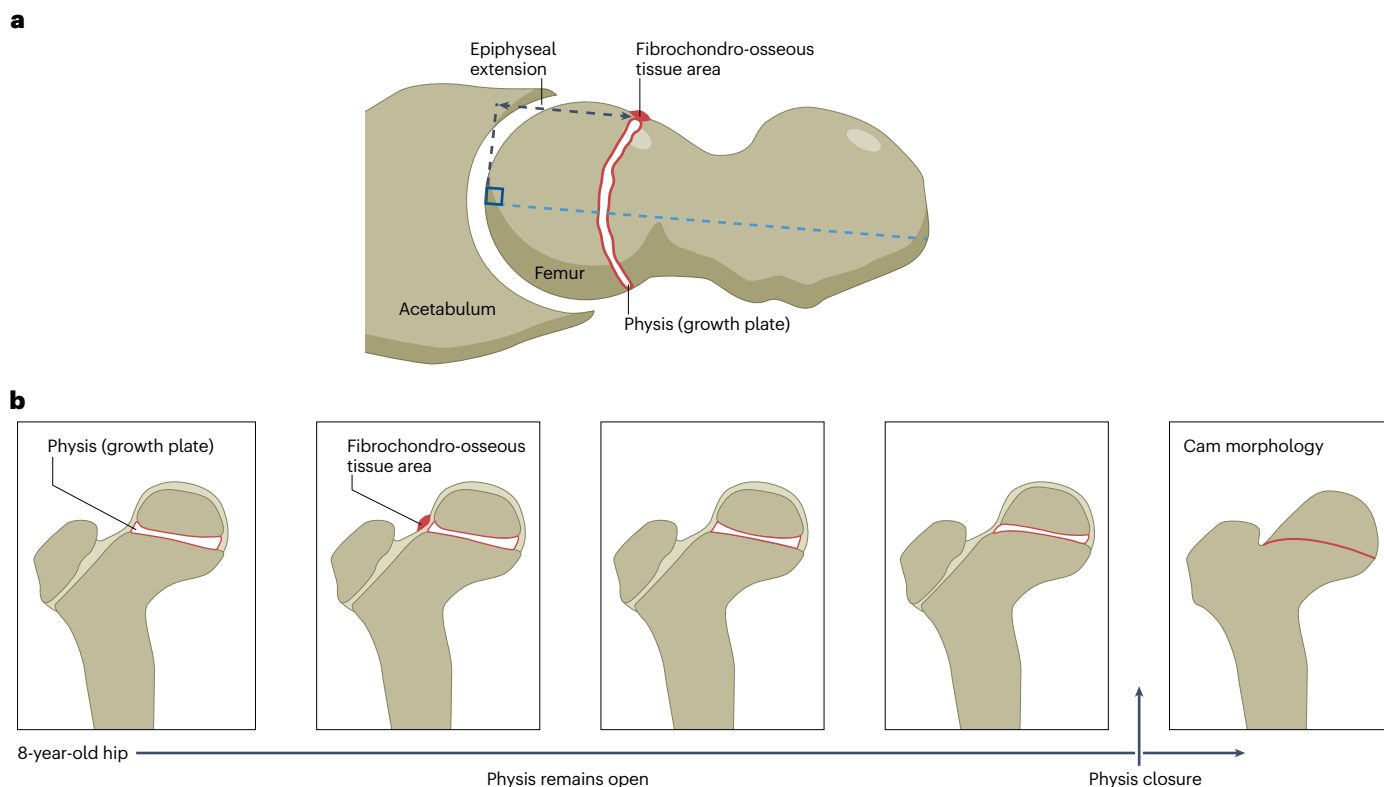


Fig. 3 | Tissue-based mechanisms and development of cam morphology. **a**, Epiphyseal extension and hypertrophy of the fibrochondro-osseous tissue area at the femoral head–neck junction are mechanisms implicated in the development of cam morphology. The fibrochondro-osseous tissue area represents a composite anatomical structure comprising the ossification groove of Ranvier and the perichondral fibrous ring of La Croix. Epiphyseal extension refers to growth or expansion of the epiphysis and is quantified by measuring the distance between the femoral head and the most distal aspect of the physis

(growth plate), along a line that is parallel to the long axis of the femoral neck (dashed light blue line). This measure is then normalized to the diameter of the femoral head to account for individual size variation. **b**, At 8 years of age, a normal hip anatomy is observed. Soft-tissue hypertrophy is hypothesized to precede the extension of the femoral epiphysis (red zone), which progresses while the physis remains open. Following physeal closure, cam morphology becomes established. The proposed age ranges for growth-plate closure and cam morphology development might vary between individuals.

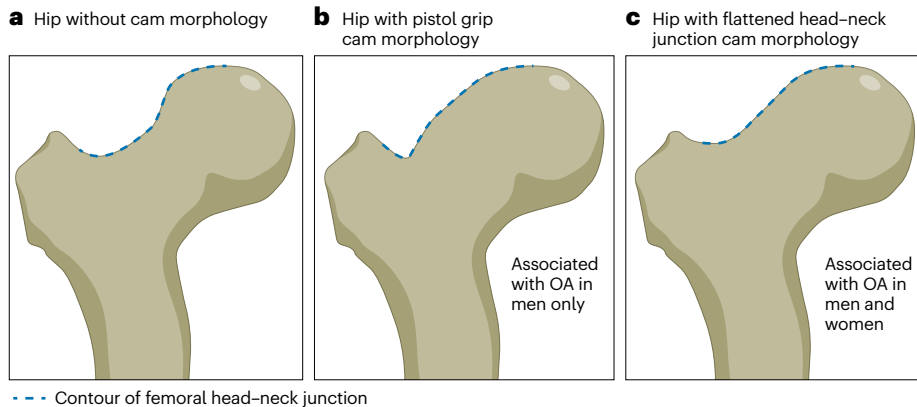


Fig. 4 | Cam morphology subtypes associated with hip osteoarthritis development. Cam morphology is a known risk factor for hip osteoarthritis (OA), with distinct subtypes showing variable associations²⁸. **a**, Hip without cam morphology. **b**, Hip with pistol grip cam morphology (associated with hip OA in men only). **c**, Hip with flattened head-neck junction (associated with hip OA in men and women).

closes⁷⁸. This concept is supported by a number of epidemiological and surgical studies^{8,62,79,80}. Longitudinal studies of high-school athletes and elite male football players have confirmed an association between cam morphology (but not pincer morphology) and early-stage OA^{9,81}. A subsequent cross-sectional study of pain-free elite male Australian football players (mean age 18.6 years) also found that cam rather than pincer morphology is linked to early-stage OA⁶². In adolescents (median age 16.7 years) undergoing surgery for FAI syndrome, greater cam morphology size was associated with surgically identified cartilage loss (with an odds ratio of 1.77 for every 10° increase in alpha angle)⁷⁹. In the same study, the presence of crossover sign seemed to have a protective effect against cartilage loss⁷⁹. Studies of adults aged 20 years and older consistently demonstrate an association between cam morphology and early-stage OA changes, whereas evidence for pincer morphology remains conflicting^{8,82–85}. A cross-sectional investigation of young adult football players with and without hip pain identified a modest dose–response relationship between cam morphology and early-stage OA; each 1° increase in alpha angle was associated with a 3% greater odds of cartilage damage⁸. Meta-analytical evidence from surgical studies has also shown a link between greater alpha angle and increased severity of cartilage damage⁸⁴. The relationship between cam morphology and the severity of early-stage OA should be considered when assessing an individual’s risk of disease progression.

Extensive evidence shows that cam morphology has an important causal role in the development of hip OA, as confirmed through radiographic or surgical assessment^{7,11,20,86}. A meta-analysis of three prospective cohort studies, involving 10,523 hips in older adults aged >45 years, found that cam morphology confers a 2.5-fold increased odds of developing hip OA over a median follow-up of 9 years, relative to hips without cam morphology¹¹. Longitudinal data from the CHECK cohort have highlighted a sustained and consistent association between cam morphology and the development of OA, observed across four follow-up assessments over a 10-year period⁸⁷. Nevertheless, the strength of the association between cam morphology and hip OA varies widely, and many individuals with this hip morphology do not go on to develop the disease.

At least two emerging insights could help rheumatologists to identify the type of cam morphology and specific contexts that increase the risk of future hip OA. First, researchers have proposed distinct subtypes of cam morphology, of which only some expedite hip OA. Cam morphology is a collective term for various forms of extra bone

formation at the femoral head–neck junction. Although researchers first described subtypes of cam morphology (such as pistol grip deformity) over 50 years ago, robust evidence supporting a causal relationship with OA has remained limited⁸⁸. To date, cam morphology is mostly quantified using an alpha angle threshold greater than 60°. However, this parameter only measures one aspect of cam morphology (that is, sphericity) and such threshold values probably encompass a spectrum of morphological subtypes. Analyses from the CHECK study used statistical shape modelling to characterize subtypes of cam morphology and investigate their associations with the development of hip OA over 10 years²⁸. A pistol-grip-shaped femoral head was associated with hip OA exclusively in men, whereas a flattened head–neck junction showed associations with hip OA in both men and women²⁸ (Fig. 4). These findings confirm that specific subtypes of cam morphology could have greater relevance than others in OA development. The unique characteristics of each subtype cannot be fully captured by the alpha angle alone. Therefore, incorporating additional radiologic measures (such as femoral head–neck offset; Fig. 1) might be needed in clinical settings to better capture cam morphology subtypes and more accurately assess future OA risk.

A second emerging insight relates to the importance of stratifying risk by both sex and age. Data from the World COACH Consortium^{5,89} – a global initiative that pooled and harmonized individual participant data from all nine available prospective cohort studies on hip OA, encompassing 23,886 hips – suggests that specific patient subgroups with cam morphology have a greater risk of developing hip OA⁸⁹. Stratified analyses reveal that men with cam morphology have an increased risk of developing hip OA (relative risk 2.50) compared with men without cam morphology. By contrast, women with cam morphology also demonstrate increased risk (relative risk 1.75), but to a lesser extent⁸⁹. Age-specific analyses further identify individuals aged 51–60 years with cam morphology as being at particularly high risk (relative risk of 2.15), with this subgroup exhibiting a higher relative risk than other age groups (40–50, 61–70 and >70 years). These findings further highlight the importance of risk stratification when evaluating the relevance of cam morphology in hip OA development.

Meta-analytical evidence from three prospective cohort studies (mean age >55 years, including 10,481 hips) indicates that pincer morphology does not increase the likelihood of OA development (odds ratio of 1.08) when compared with individuals without pincer morphology¹¹. However, newly analysed data from the CHECK study indicates that hip pain might modify the relationship between pincer

morphology and OA⁹⁰. For example, individuals presenting with a combination of baseline hip pain and pincer morphology (defined by an anterior centre edge angle >40°) had an increased odds of developing hip OA at 5-year (odds ratio of 3.41), 8-year (odds ratio of 2.36) and 10-year (1.97) follow-up, compared with individuals who had pincer morphology but no hip pain⁹⁰. Finally, evidence of an association between protrusio acetabuli and hip OA development remains inconclusive. Two population-based prospective studies have reported conflicting evidence: the Johnston County OA Project⁴¹ found a fourfold increased risk of hip OA in women with protrusio acetabuli, but not in men, whereas the Chingford Study⁹¹ did not find any association.

Management

Over the past decade, considerable progress has been made in the management of FAI syndrome, with emerging clinical trial evidence supporting both surgical and non-surgical treatment approaches^{92–99}.

FAI syndrome

FAI syndrome can be managed with surgical and non-surgical treatments^{10,100}. The current evidence, discussed in this section, is derived from studies involving people with FAI syndrome without coexisting radiographic hip OA. Hip arthroscopy is the most common surgical technique for FAI syndrome, with its use largely concentrated in high-income countries. The procedure typically involves resection of the cam and/or pincer morphology, and is frequently accompanied by treatment of co-existing cartilage and labral pathology¹⁰. Common non-surgical strategies include activity modification, education, intra-articular injections and physiotherapy-led incremental exercise rehabilitation^{92,93,100}. Patient education is a cornerstone of non-surgical treatment and centres on activity modification, recommending avoidance of positions and activities usually associated with pain (for example, squatting, leg crossing, pivoting, high-joint-load activity and floor sitting)¹⁰¹. Patients should also be made aware of the high prevalence of cam and pincer morphology in pain-free individuals to help them contextualize their imaging findings²³. Therapeutic intra-articular injections are often used in clinical settings in conjunction with exercise rehabilitation or in those with high pain levels^{10,92}. Both corticosteroids and hyaluronic acid can provide modest short-term pain relief, although longer-term effects seem to vary considerably between individuals⁹². Importantly, the potential for either injection to modify disease progression remains unclear. Despite growing interest in orthobiologic injections (for example, platelet-rich plasma and mesenchymal stem cells), at present no clinical trial evidence is available to support the effectiveness of such approaches in people with FAI syndrome⁹².

A growing body of research is focused on examining the effectiveness of hip arthroscopy and physiotherapy-led treatments in improving patient-reported outcomes and modifying disease progression in individuals with FAI syndrome. Three randomized controlled trials have compared surgery to physiotherapist-led treatment for FAI syndrome, examining the effects on pain and quality of life^{94,95,99}. Although both treatment groups showed improvements in pain, the superiority of arthroscopy compared over physiotherapy was modest at 6–12 months. At two-year follow-up, one trial reported no advantage of arthroscopy over physiotherapist-led treatment in terms of pain relief or quality-of-life outcomes⁹³. However, a more recent randomized controlled trial reported that, at 3-year follow-up, arthroscopy was superior to physiotherapist-led treatment, resulting in an 8.9-point improvement (out of 100; 95% confidence interval 7.0–10.8) in activities

of daily living compared with physiotherapy-led treatment, among the 77% of participants who completed follow-up⁹⁶.

Consensus from the International Hip Pain Research Network recommends physiotherapist-led treatment as the first-line approach for managing FAI syndrome¹⁰⁰. Several small-scale studies have compared different physiotherapy-led treatments¹⁰⁰, and a full-scale randomized controlled trial (PhysioFIRST)¹⁰² is under way. This trial is comparing a targeted strengthening intervention with standardized stretching in 154 individuals with FAI syndrome¹⁰², and is expected to provide robust evidence on the comparative effect of these approaches on patient outcomes. Optimal treatment (surgical or non-surgical) of FAI syndrome should involve shared decision-making, enabling patients to make informed choices. Clinicians should work within a multidisciplinary team with access to all treatment options.

Although both surgical and non-surgical treatments are primarily aimed at reducing pain and improving quality of life, growing interest has emerged in their potential to modify the course of OA^{27,98,103}. So far, two randomized controlled trials have investigated the effects of arthroscopy versus physiotherapy on MRI-defined hip joint structure: one study with a 12-month follow-up⁹⁸ and the other with a 3-year follow-up⁹⁶. At 12 months, no difference was observed between the surgery and physiotherapy groups in terms of hip joint structure⁹⁸. At 3-year follow-up, radiographic joint space width remained similar across both groups, but the Scoring Hip OA with MRI^{104,105} score was lower in the arthroscopy group (mean 9.22, standard deviation 11.43)

Glossary

Acetabular labrum

An intra-articular fibrocartilaginous triangle that traverses the anterior and posterior acetabular rim, joining with the transverse acetabular ligament inferiorly to create a continuous ring. It functions to improve joint stability, distribute joint forces and preserve articular cartilage.

Acetabular version

The orientation of the acetabulum in the horizontal plane.

Acetabulum

The cup-shaped bony socket in the pelvis that forms the hip joint by articulating with the head of the femur.

Crossover sign

A radiographic indicator of acetabular retroversion; present when the anterior acetabular wall crosses over the posterior acetabular wall.

Femoral version

The amount of rotation or torsion between the proximal and distal parts of the femur.

Ischial spine sign

A radiographic feature associated with acetabular retroversion; considered present when the triangular ischial spine protrudes medially into the pelvic cavity.

Posterior wall sign

A radiographic sign of insufficient posterior acetabular coverage; considered present when the projection of the posterior acetabular wall is medial to the centre of the femoral head.

Protrusio acetabuli

A radiographic finding indicating medial displacement of the acetabulum; occurs when the medial wall of the acetabulum projects into the pelvic cavity, resulting in the femoral head crossing or touching the ilioischial line.

Spinopelvic parameters

A set of radiographic measurements used to assess the relationship between the hip, pelvis and lumbar spine in the sagittal plane (namely pelvic incidence, sacral slope and spinopelvic tilt).

compared with the physiotherapy group (mean 22.76, standard deviation 15.26), suggesting that arthroscopy might be associated with reduced MRI-detected joint damage relative to physiotherapy⁹⁶. No clinical trials have examined the effects of exercise-based treatments on the trajectory of hip OA in individuals with FAI syndrome. Hip muscle weakness, altered movement and changes to physical activity are usually seen in individuals with FAI syndrome¹⁰⁶, and might also alter hip joint forces, influencing the trajectory to hip OA¹⁰⁷. Exercise-based treatment programmes that target hip muscle weakness, enhance movement control and improve physical activity might be especially beneficial in altering the risk of future hip OA. However, these interventions must be delivered at a sufficient dosage to effectively target impairments and alter joint forces¹⁰⁸, an aspect that might have been suboptimal in the reported randomized controlled trials^{94,96,98}.

FAI syndrome with hip OA

People with FAI syndrome and established radiographic hip OA present a complex therapeutic challenge in rheumatology practice¹⁰⁹. An expert-panel Delphi study identified genuine uncertainty – clinical equipoise – among experts regarding which treatment is superior for this patient population¹¹⁰. The expert group recommended that physiotherapy-led treatments should consist of core and hip muscle strengthening, enhancement of lumbo-pelvic mobility, avoidance of high-joint-load positions and activity modification. Hip arthroscopy could be considered for younger patients and/or those with a normal body mass index¹¹⁰. By contrast, people with severe joint space narrowing, advanced radiographic disease (that is, Tönnis grade 3 OA) or bilateral cartilage damage involving both the acetabular and femoral head as identified on MRI should be regarded as poor candidates for hip arthroscopy¹¹⁰. For young adults who are not candidates for arthroscopy and do not respond to physiotherapy-led interventions, total hip arthroplasty might be considered as a treatment option¹¹¹.

Opportunities for cam morphology prevention

Growing interest surrounds the prevention of cam morphology in children and adolescents, because of its associated increased risk of FAI syndrome and hip OA in later life⁴⁴. However, despite this increased risk, not all people with cam morphology develop hip OA^{11,28,86,89}. Young athletes with open femoral epiphyseal growth plates represent a critical target population for primary prevention strategies^{29,30,32,46}. The cornerstone of a prevention programme would probably involve reducing sporting load during periods of skeletal growth, particularly in young athletes engaged in high-joint-load sports linked to cam morphology formation (such as football and ice hockey). However, findings from the Oxford Consensus study (parts 1 and 2), which included input from parents, athletes, coaches, clinicians and scientists, emphasized the need for further research to identify risk factors for cam morphology in youth athletes before implementing prevention programmes^{25,27}. Without such knowledge, the benefits of sports participation during youth far outweigh the risks and consequences of cam morphology, and thus primary prevention interventions for cam morphology cannot currently be recommended⁴⁴.

Conclusion

Cam morphology is present in individuals with and without FAI syndrome and is associated with an increased risk of developing early hip OA. Evidence suggests that exposure to high-joint-load physical activity during skeletal maturation has an important role in cam

morphology formation. Imaging remains a cornerstone of the diagnosis of hip morphology but should always be interpreted with caution and in conjunction with clinical symptoms and physical findings. Cam morphology might contribute to the onset of early-stage hip OA from late adolescence, whereas additional factors such as pincer morphology, femoral version and spinopelvic parameters probably influence FAI syndrome development. Longitudinal mechanistic evidence supports cam morphology as a well defined causal risk factor for hip OA, and emerging data also highlight the importance of FAI syndrome in disease development. Specific subtypes of cam morphology and demographic characteristics seem to be more strongly associated with hip OA than others and could inform patient stratification. Although both surgical and non-surgical interventions are available for managing FAI syndrome, existing clinical trial evidence supporting these approaches is predominantly derived from people without coexisting hip OA. At present, strategies to prevent cam morphology development (primary prevention) are not considered feasible. Continued efforts are needed to advance diagnostic approaches for FAI syndrome and to individualize surgical, non-surgical and orthobiologic treatments to enhance clinical outcomes and reduce the future burden of hip OA.

Published online: 26 November 2025

References

1. Murphy, L. B. et al. One in four people may develop symptomatic hip osteoarthritis in his or her lifetime. *Osteoarthr. Cartil.* **18**, 1372–1379 (2010).
2. Hunter, D. J., Schofield, D. & Callander, E. The individual and socioeconomic impact of osteoarthritis. *Nat. Rev. Rheumatol.* **10**, 437–441 (2014).
3. Hunter, D. J. & Bierma-Zeinstra, S. Osteoarthritis. *Lancet* **393**, 1745–1759 (2019).
4. Murphy, N. J., Eyles, J. P. & Hunter, D. J. Hip osteoarthritis: etiopathogenesis and implications for management. *Adv. Ther.* **33**, 1921–1946 (2016).
5. Van Buuren, M. M. A. et al. Cohort profile: Worldwide Collaboration on Osteoarthritis Prediction for the Hip (World COACH) – an international consortium of prospective cohort studies with individual participant data on hip osteoarthritis. *BMJ Open* **14**, e077907 (2024).
6. Agricola, R. et al. Cam impingement of the hip – a risk factor for hip osteoarthritis. *Nat. Rev. Rheumatol.* **9**, 630–634 (2013).
7. Ganz, R. et al. Femoroacetabular impingement: a cause for osteoarthritis of the hip. *Clin. Orthop.* **417**, 112–120 (2003).
8. Heerey, J. et al. Cam morphology is associated with MRI-defined cartilage defects and labral tears: a case-control study of 237 young adult football players with and without hip and groin pain. *BMJ Open Sport Exerc. Med.* **7**, e001199 (2021).
9. Claes, P. A. M. et al. The association between the development of cam morphology during skeletal growth in high-impact athletes and the presence of cartilage loss and labral damage in adulthood: a prospective cohort study with a 12-year follow-up. *Am. J. Sports Med.* **52**, 2555–2564 (2024).
10. Griffin, D. R. et al. The Warwick agreement on femoroacetabular impingement syndrome (FAI syndrome): an international consensus statement. *Br. J. Sports Med.* **50**, 1169–1176 (2016).
11. Casartelli, N. C. et al. Is hip morphology a risk factor for developing hip osteoarthritis? A systematic review with meta-analysis. *Osteoarthr. Cartil.* **29**, 1252–1264 (2021).
12. Agricola, R. et al. Femoroacetabular impingement syndrome in middle-aged individuals is strongly associated with the development of hip osteoarthritis within 10-year follow-up: a prospective cohort study (CHECK). *Br. J. Sports Med.* **58**, 1061–1067 (2024).
13. Reiman, M. P. et al. Consensus recommendations on the classification, definition and diagnostic criteria of hip-related pain in young and middle-aged active adults from the International Hip-related Pain Research Network, Zurich 2018. *Br. J. Sports Med.* **54**, 631–641 (2020).
14. Reiman, M. P. & Thorborg, K. Clinical examination and physical assessment of hip joint-related pain in athletes. *Int. J. Sports Phys. Ther.* **9**, 737–755 (2014).
15. Lerch, T. D. et al. Location of intra- and extra-articular hip impingement is different in patients with pincer-type and mixed-type femoroacetabular impingement due to acetabular retroversion or protrusio acetabuli on 3D CT-based impingement simulation. *Am. J. Sports Med.* **48**, 661–672 (2020).
16. Mascarenhas, V. V. et al. Can we discriminate symptomatic hip patients from asymptomatic volunteers based on anatomic predictors? A 3-dimensional magnetic resonance study on cam, pincer, and spinopelvic parameters. *Am. J. Sports Med.* **46**, 3097–3110 (2018).
17. Grammatopoulos, G. et al. Acetabular and spino-pelvic morphologies are different in subjects with symptomatic cam femoro-acetabular impingement: acetabular and spinopelvic morphology in FAI. *J. Orthop. Res.* **36**, 1840–1848 (2018).

18. Lerch, T. D. et al. Hip impingement location in maximal hip flexion in patients with femoroacetabular impingement with and without femoral retroversion. *Am. J. Sports Med.* **50**, 2989–2997 (2022).
19. Heerey, J. et al. The size and prevalence of bony hip morphology do not differ between football players with and without hip and/or groin pain: findings from the FORCe cohort. *J. Orthop. Sports Phys. Ther.* **51**, 115–125 (2021b).
20. van Klij, P., Heerey, J., Waarsing, J. H. & Agricola, R. The prevalence of cam and pincer morphology and its association with development of hip osteoarthritis. *J. Orthop. Sports Phys. Ther.* **48**, 230–238 (2018).
21. van Klij, P. et al. The relationship between cam morphology and hip and groin symptoms and signs in young male football players. *Scand. J. Med. Sci. Sports* **30**, 1221–1231 (2020).
22. Scholes, M. J. et al. Does femoroacetabular impingement syndrome affect self-reported burden in football players with hip and groin pain? *Sports Health* **14**, 920–931 (2022).
23. Dickenson, E. et al. Prevalence of cam hip shape morphology: a systematic review. *Osteoarthr. Cartil.* **24**, 949–961 (2016).
24. Mascarenhas, V. V. et al. Imaging prevalence of femoroacetabular impingement in symptomatic patients, athletes, and asymptomatic individuals: a systematic review. *Eur. J. Radiol.* **85**, 73–95 (2016).
25. Dijkstra, H. P. et al. Oxford Consensus on primary cam morphology and femoroacetabular impingement syndrome. Part 1 — definitions, terminology, taxonomy and imaging outcomes. *Br. J. Sports Med.* **57**, 325–341 (2023).
26. Dijkstra, H. P. et al. Primary cam morphology: bump, burden or bog-standard? A concept analysis. *Br. J. Sports Med.* **55**, 1212–1221 (2021).
27. Dijkstra, H. P. et al. Oxford Consensus on primary cam morphology and femoroacetabular impingement syndrome. Part 2 — research priorities on conditions affecting the young person's hip. *Br. J. Sports Med.* **57**, 342–358 (2023).
28. Tang, J. et al. The different subtypes of cam morphology as defined by statistical shape modeling and their relationship with the development of hip osteoarthritis: a nationwide prospective cohort study (CHECK) with 10 years follow-up. *Osteoarthr. Cartil.* **32**, 1647–1654 (2024).
29. Palmer, A. et al. Physical activity during adolescence and the development of cam morphology: a cross-sectional cohort study of 210 individuals. *Br. J. Sports Med.* **52**, 601–610 (2018).
30. Fernquest, S. et al. A longitudinal cohort study of adolescent elite footballers and controls investigating the development of cam morphology. *Sci. Rep.* **11**, 18567 (2021).
31. Agricola, R. et al. A cam deformity is gradually acquired during skeletal maturation in adolescent and young male soccer players: a prospective study with minimum 2-year follow-up. *Am. J. Sports Med.* **42**, 798–806 (2014).
32. van Klij, P. et al. Cam morphology in young male football players mostly develops before proximal femoral growth plate closure: a prospective study with 5-year follow-up. *Br. J. Sports Med.* **53**, 532–538 (2019).
33. Mascarenhas, V. V. et al. The Lisbon agreement on femoroacetabular impingement imaging — part 1: overview. *Eur. Radiol.* **30**, 5281–5297 (2020).
34. Lerch, T. D. et al. Prevalence of femoral and acetabular version abnormalities in patients with symptomatic hip disease: a controlled study of 538 hips. *Am. J. Sports Med.* **46**, 122–134 (2018).
35. Aggarwal, R. et al. Distinctions between diagnostic and classification criteria? *Arthritis Care Res.* **67**, 891–897 (2015).
36. Castro, M. O. et al. The Lisbon agreement on femoroacetabular impingement imaging — part 3: imaging techniques. *Eur. Radiol.* **31**, 4652–4668 (2021).
37. Mascarenhas, V. V. et al. The Lisbon agreement on femoroacetabular impingement imaging — part 2: general issues, parameters, and reporting. *Eur. Radiol.* **31**, 4634–4651 (2021).
38. Mascarenhas, V. V. et al. Imaging methodology for hip preservation: techniques, parameters, and thresholds. *Semin. Musculoskelet. Radiol.* **23**, 197–226 (2019).
39. Notzli, H. P. et al. The contour of the femoral head-neck junction as a predictor for the risk of anterior impingement. *J. Bone Jt. Surg. Br.* **84**, 556–560 (2002).
40. Mascarenhas, V. V., Caetano, A., Dantas, P. & Rego, P. Advances in FAI imaging: a focused review. *Curr. Rev. Musculoskelet. Med.* **13**, 622–640 (2020).
41. Nelson, A. E. et al. Measures of hip morphology are related to development of worsening radiographic hip osteoarthritis over 6 to 13 year follow-up: the Johnston County osteoarthritis project. *Osteoarthr. Cartil.* **24**, 443–450 (2016).
42. van Klij, P. et al. Classifying cam morphology by the alpha angle: a systematic review on threshold values. *Orthop. J. Sports Med.* **8**, 2325967120938312 (2020).
43. Thirumaran, A. J., Murphy, N. J., Fu, K. & Hunter, D. J. Femoroacetabular impingement — what the rheumatologist needs to know. *Best. Pract. Res. Clin. Rheumatol.* **38**, 101932 (2024).
44. Heerey, J. J. et al. Preventing hip osteoarthritis in athletes: is it really a mission impossible? *Br. J. Sports Med.* **58**, 465–467 (2024).
45. Westermann, R. W. et al. Activity level and sport type in adolescents correlate with the development of cam morphology. *JBJS Open Access.* **6**, e21.00059 (2021).
46. Hanke, M. S. et al. A cam morphology develops in the early phase of the final growth spurt in adolescent ice hockey players: results of a prospective MRI-based study. *Clin. Orthop.* **479**, 906–918 (2021).
47. Ayeni, O. R. et al. Prospective evaluation of sport activity and the development of femoroacetabular impingement in the adolescent hip (PREVIEW): results of the pilot study. *Pilot Feas. Stud.* **8**, 201 (2022).
48. Hanzlik, S. et al. The prevalence of cam morphology: a cross-sectional evaluation of 3,558 cadaveric femora. *Front. Surg.* **7**, 588535 (2020).
49. Doran, C., Pettit, M., Singh, Y., Sunil Kumar, K. H. & Khanduja, V. Does the type of sport influence morphology of the hip? A systematic review. *Am. J. Sports Med.* **50**, 1727–1741 (2022).
50. Tak, I. et al. The relationship between the frequency of football practice during skeletal growth and the presence of a cam deformity in adult elite football players. *Br. J. Sports Med.* **49**, 630–634 (2015).
51. Mosler, A. B. et al. Ethnic differences in bony hip morphology in a cohort of 445 professional male soccer players. *Am. J. Sports Med.* **44**, 2967–2974 (2016).
52. Zheng, J. et al. Comparison between UK Biobank and Shanghai Changfeng suggests distinct hip morphology may contribute to ethnic differences in the prevalence of hip osteoarthritis. *Osteoarthr. Cartil.* **33**, 473–481 (2025).
53. Edwards, K. et al. Differences between race and sex in measures of hip morphology: a population-based comparative study. *Osteoarthr. Cartil.* **28**, 189–200 (2020).
54. Mirtz, T. A., Chandler, J. P. & Eysers, C. M. The effects of physical activity on the epiphyseal growth plates: a review of the literature on normal physiology and clinical implications. *J. Clin. Med. Res.* **3**, 1–7 (2011).
55. Vaswani, R., White, A. E., Feingold, J. & Ranawat, A. S. Hip–spine syndrome in the nonarthritic patient. *Arthroscopy* **38**, 2930–2938 (2022).
56. Rivière, C. et al. Spine–hip relations add understandings to the pathophysiology of femoro-acetabular impingement: a systematic review. *Orthop. Traumatol. Surg. Res.* **103**, 549–557 (2017).
57. Torabian, K. A. et al. The effect of pelvic incidence on outcomes after hip arthroscopy for femoroacetabular impingement and acetabular labral tears. *Am. J. Sports Med.* **52**, 631–642 (2024).
58. Ross, J. R. et al. Effect of changes in pelvic tilt on range of motion to impingement and radiographic parameters of acetabular morphologic characteristics. *Am. J. Sports Med.* **42**, 2402–2409 (2014).
59. Saltychev, M., Perna, K., Seppänen, M., Mäkelä, K. & Laimi, K. Pelvic incidence and hip disorders: a systematic review and quantitative analysis. *Acta Orthop.* **89**, 66–70 (2018).
60. Kraeutler, M. J., Chadayammuri, V., Garabekyan, T. & Mei-Dan, O. Femoral version abnormalities significantly outweigh effect of cam impingement on hip internal rotation. *J. Bone Jt Surg.* **100**, 205–210 (2018).
61. Murphy, N. J. et al. Combined femoral and acetabular version and synovitis are associated with dGEMRIC scores in people with femoroacetabular impingement (FAI) syndrome. *J. Orthop. Res.* **41**, 2484–2494 (2023).
62. Heerey, J. et al. Are hip joint imaging findings associated with symptoms and early hip osteoarthritis features in elite male Australian football league draftees? *Sci. Med. Footb.* **9**, 341–348 (2024).
63. Tagliero, A. J., Foster, M. J., Melugin, H. P. & Su, C. A. Inflammatory and immunologic contributions in femoroacetabular impingement syndrome. *J. Am. Acad. Orthop. Surg.* **31**, 1097–1102 (2023).
64. Tang, J. et al. The association between cam morphology and hip pain in males and females within 10 years: a national prospective cohort study (CHECK). *Semin. Arthritis Rheum.* **69**, 152539 (2024).
65. Khanna, V., Caragianis, A., Diprimio, G., Rakhra, K. & Beaulieu, P. E. Incidence of hip pain in a prospective cohort of asymptomatic volunteers: is the cam deformity a risk factor for hip pain? *Am. J. Sports Med.* **42**, 793–797 (2014).
66. Khan, A. Z. et al. Factors associated with disease progression in the contralateral hip of patients with symptomatic femoroacetabular impingement: a minimum 5-year analysis. *Am. J. Sports Med.* **50**, 3174–3183 (2022).
67. Dzdizshvili, L. et al. Increased alpha angles and younger age increase the risk of contralateral femoral acetabular impingement syndrome symptom development and surgical intervention: a systematic review. *Arthroscopy* **40**, 1358–1365 (2024).
68. Sutter, R., Dietrich, T. J., Zingg, P. O. & Pfirrmann, C. W. How useful is the alpha angle for discriminating between symptomatic patients with cam-type femoroacetabular impingement and asymptomatic volunteers? *Radiology* **264**, 514–521 (2012).
69. Gosvig, K. K., Jacobsen, S., Sonne-Holm, S., Palm, H. & Troelsen, A. Prevalence of malformations of the hip joint and their relationship to sex, groin pain, and risk of osteoarthritis: a population-based survey. *J. Bone Jt Surg. Am.* **92**, 1162–1169 (2010).
70. Larson, C. M. et al. Increasing alpha angle is predictive of athletic-related ‘hip’ and ‘groin’ pain in collegiate National Football League prospects. *Arthroscopy* **29**, 405–410 (2013).
71. Saito, M., Tsukada, S., Yoshida, K., Okada, Y. & Tasaki, A. Correlation of alpha angle between various radiographic projections and radial magnetic resonance imaging for cam deformity in femoral head-neck junction. *Knee Surg. Sports Traumatol. Arthrosc.* **25**, 77–83 (2017).
72. Domayer, S. E. et al. Femoroacetabular cam-type impingement: diagnostic sensitivity and specificity of radiographic views compared to radial MRI. *Eur. J. Radiol.* **80**, 805–810 (2011).
73. Scholes, M. J. et al. Are cam morphology size and location associated with self-reported burden in football players with FAI syndrome? *Scand. J. Med. Sci. Sports* **32**, 737–753 (2022).
74. Siebelt, M., Agricola, R., Weinans, H. & Kim, Y. J. The role of imaging in early hip OA. *Osteoarthr. Cartil.* **22**, 1470–1480 (2014).
75. Chu, C. R. et al. Transformative approaches for effective clinical trials to reduce the disease burden of osteoarthritis. *Semin. Arthritis Rheum.* **71**, 152652 (2025).
76. Heerey, J. et al. Prevalence of early hip OA features on MRI in high-impact athletes. The Femoroacetabular Impingement and Hip Osteoarthritis Cohort (FORCe) study. *Osteoarthr. Cartil.* **29**, 323–334 (2021).

77. Mahmoudian, A., Lohmander, L. S., Mobasheri, A., Englund, M. & Luyten, F. P. Early-stage symptomatic osteoarthritis of the knee — time for action. *Nat. Rev. Rheumatol.* **17**, 621–632 (2021).
78. Fernquest, S. et al. The response of hip joint cartilage to exercise in children: an MRI study using T2-mapping. *Cartilage* **13**, 1761S–1771S (2020).
79. McClincy, M. P. et al. Clinical and radiographic predictors of acetabular cartilage lesions in adolescents undergoing hip arthroscopy. *Am. J. Sports Med.* **46**, 3082–3089 (2018).
80. Reichenbach, S. et al. Association between cam-type deformities and magnetic resonance imaging-detected structural hip damage: a cross-sectional study in young men. *Arthritis Rheum.* **63**, 4023–4030 (2011).
81. Wyles, C. C. et al. Cam deformities and limited hip range of motion are associated with early osteoarthritic changes in adolescent athletes: a prospective matched cohort study. *Am. J. Sports Med.* **45**, 3036–3043 (2017).
82. Ishøi, L. et al. Demographic and radiographic factors associated with intra-articular hip cartilage injury: a cross-sectional study of 1511 hip arthroscopy procedures. *Am. J. Sports Med.* **47**, 2617–2625 (2019).
83. Dumont, G. D. et al. Patient-specific factors are associated with severity of chondrolabral injury in patients with femoroacetabular impingement. *Knee Surg. Sports Traumatol. Arthrosc.* **28**, 3376–3381 (2020).
84. Martins, E. C., Gomes, D. A., Fernandes, D. A. & De Brito Fontana, H. Alpha angle values predict the severity of hip chondral damage in patients with cam-type femoroacetabular impingement syndrome: a systematic review and meta-analysis. *Arthrosc. J. Arthrosc. Relat. Surg.* **41**, 1539–1553.e5 (2025).
85. Kapron, A. L., Aoki, S. K., Weiss, J. A., Krych, A. J. & Maak, T. G. Isolated focal cartilage and labral defects in patients with femoroacetabular impingement syndrome may represent new, unique injury patterns. *Knee Surg. Sports Traumatol. Arthrosc.* **27**, 3057–3065 (2019).
86. Agricolà, R. et al. Cam impingement causes osteoarthritis of the hip: a nationwide prospective cohort study (CHECK). *Ann. Rheum. Dis.* **72**, 918–923 (2013).
87. Tang, J. et al. Cam morphology is strongly and consistently associated with development of radiographic hip osteoarthritis throughout 4 follow-up visits within 10 years. *Osteoarthr. Cartil.* **31**, 1650–1656 (2023).
88. Beaulé, P. E. et al. Unravelling the hip pistol grip/cam deformity: origins to joint degeneration. *J. Orthop. Res.* **36**, 3125–3135 (2018).
89. Tang, J. et al. Cam morphology and the risk of developing radiographic hip osteoarthritis within 8 years: an individual participant data meta-analysis of 23,886 hips from the world COACH consortium. *Br. J. Sports Med.* <https://doi.org/10.1136/bjsports-2025-101444> (2025).
90. Riedstra, N. S. et al. Pincer morphology is not associated with hip osteoarthritis unless hip pain is present: follow-up data from a prospective cohort study. *Arthritis Care Res.* **76**, 644–651 (2024).
91. Nicholls, A. S. et al. The association between hip morphology parameters and nineteen-year risk of end-stage osteoarthritis of the hip: a nested case-control study. *Arthritis Rheum.* **63**, 3392–3400 (2011).
92. Probst, D. T. et al. What is the rate of response to nonoperative treatment for hip-related pain? A systematic review with meta-analysis. *J. Orthop. Sports Phys. Ther.* **53**, 286–306 (2023).
93. Kemp, J. L. et al. Improving function in people with hip-related pain: a systematic review and meta-analysis of physiotherapist-led interventions for hip-related pain. *Br. J. Sports Med.* **54**, 1382–1394 (2020).
94. Griffin, D. R. et al. Hip arthroscopy versus best conservative care for the treatment of femoroacetabular impingement syndrome (UK FASHIoN): a multicentre randomised controlled trial. *Lancet* **391**, 2225–2235 (2018).
95. Palmer, A. J. R. et al. Arthroscopic hip surgery compared with physiotherapy and activity modification for the treatment of symptomatic femoroacetabular impingement: multicentre randomised controlled trial. *BMJ* **364**, l185 (2019).
96. Palmer, A. et al. Medium-term results of arthroscopic hip surgery compared with physiotherapy and activity modification for the treatment of femoroacetabular impingement syndrome: a multi-centre randomised controlled trial. *Br. J. Sports Med.* **59**, 109–117 (2025).
97. Femoroacetabular Impingement Randomized Controlled Trial (FIRST) Investigators. Osteochondroplasty and labral repair for the treatment of young adults with femoroacetabular impingement: a randomized controlled trial. *Am. J. Sports Med.* **49**, 25–34 (2021).
98. Hunter, D. J. et al. Multi-centre randomised controlled trial comparing arthroscopic hip surgery to physiotherapist-led care for femoroacetabular impingement (FAI) syndrome on hip cartilage metabolism: the Australian FASHIoN trial. *BMC Musculoskelet. Disord.* **22**, 697 (2021).
99. Mansell, N. S., Rhon, D. I., Meyer, J., Slevin, J. M. & Marchant, B. G. Arthroscopic surgery or physical therapy for patients with femoroacetabular impingement syndrome: a randomized controlled trial with 2-year follow-up. *Am. J. Sports Med.* **46**, 1306–1314 (2018).
100. Kemp, J. L. et al. Physiotherapist-led treatment for young to middle-aged active adults with hip-related pain: consensus recommendations from the International Hip-related Pain Research Network, Zurich 2018. *Br. J. Sports Med.* **54**, 504–511 (2020).
101. Ensek, K. R. et al. Hip pain and movement dysfunction associated with nonarthritic hip joint pain: a revision. *J. Orthop. Sports Phys. Ther.* **53**, CPG1–CPG70 (2023).
102. Kemp, J. L. et al. Physiotherapist-led treatment for femoroacetabular impingement syndrome (the PhysioFIRST study): a protocol for a participant and assessor-blinded randomised controlled trial. *BMJ Open.* **11**, e041742 (2021).
103. Bell, E. et al. What are participant beliefs regarding physical therapy led treatment? A qualitative study of people living with femoroacetabular impingement syndrome. *Braz. J. Phys. Ther.* **28**, 101077 (2024).
104. Lee, S. et al. Scoring hip osteoarthritis with MRI (SHOMRI): a whole joint osteoarthritis evaluation system. *J. Magn. Reson. Imaging* **41**, 1549–1557 (2015).
105. Heerey, J. J. et al. Defining hip osteoarthritis feature prevalence, severity, and change using the Scoring of Hip Osteoarthritis with MRI (SHOMRI). *Skelet. Radiol.* **53**, 1599–1609 (2024).
106. Freke, M. et al. Physical impairments in symptomatic femoroacetabular impingement: a systematic review of the evidence. *Br. J. Sports Med.* **50**, 1180 (2016).
107. Lewis, C. L., Sahrman, S. A. & Moran, D. W. Anterior hip joint force increases with hip extension, decreased gluteal force, or decreased iliopsoas force. *J. Biomech.* **40**, 3725–3731 (2007).
108. Kemp, J. L. et al. Is exercise therapy for femoroacetabular impingement in or out of fashion? We need to talk about current best practice for the non-surgical management of FAI syndrome. *Br. J. Sports Med.* **53**, 1204–1205 (2019).
109. Whittaker, J. L. & Roos, E. M. A pragmatic approach to prevent post-traumatic osteoarthritis after sport or exercise-related joint injury. *Best Pract. Res. Clin. Rheumatol.* **33**, 158–171 (2019).
110. Andronic, O. et al. Clinical equipoise in the management of patients with femoroacetabular impingement syndrome and concomitant Tönnis grade 2 hip osteoarthritis or greater: an international expert-panel Delphi study. *Arthrosc. J. Arthrosc. Relat. Surg.* **40**, 2029–2038.e1 (2024).
111. Galloway, R. et al. Satisfaction rates, function, and return to activity following young adult total hip arthroplasty. *Bone Jt Open.* **5**, 304–311 (2024).
112. Reiman, M. P., Goode, A. P., Cook, C. E., Hölmich, P. & Thorborg, K. Diagnostic accuracy of clinical tests for the diagnosis of hip femoroacetabular impingement/labral tear: a systematic review with meta-analysis. *Br. J. Sports Med.* **49**, 811 (2015).
113. Thorborg, K. et al. Clinical examination, diagnostic imaging, and testing of athletes with groin pain: an evidence-based approach to effective management. *J. Orthop. Sports Phys. Ther.* **48**, 239–249 (2018).
114. Chadayammuri, V. et al. Passive hip range of motion predicts femoral torsion and acetabular version. *J. Bone Jt Surg.* **98**, 127–134 (2016).
115. Pålsson, A., Kostogiannis, I. & Ageberg, E. Combining results from hip impingement and range of motion tests can increase diagnostic accuracy in patients with FAI syndrome. *Knee Surg. Sports Traumatol. Arthrosc.* **28**, 3382–3392 (2020).
116. Mosler, A. B. et al. Standardised measurement of physical capacity in young and middle-aged active adults with hip-related pain: recommendations from the first International Hip-related Pain Research Network (HiPRN) meeting, Zurich, 2018. *Br. J. Sports Med.* **54**, 702–710 (2020).

Author contributions

All authors researched data for the article and wrote the article. J.H., J.K., P.v.K., V.M., M.S., K.C., F.B., P.D., S.B.-Z. and R.A. contributed substantially to discussion of the content and reviewed and/or edited the manuscript before submission.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41584-025-01328-4>.

Peer review information *Nature Reviews Rheumatology* thanks Nicholas J. Murphy, Antony Palmer and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

© Springer Nature Limited 2025

Author Correction: Inflammation and pain as interconnected targets in axial spondyloarthritis

Correction to: *Nature Reviews Rheumatology*
<https://doi.org/10.1038/s41584-025-01348-0>,
published online 21 January 2026.

Xenofon Baraliakos , **Victoria Navarro-Compán** , **Nelly Ziade** 
& **Denis Poddubnyy** 

<https://doi.org/10.1038/s41584-026-01363-9>

Published online: 16 February 2026

 Check for updates

In the version of the article initially published, in the “Nociplastic pain” section, the sentence “Patients with axSpA who have nociplastic pain frequently show higher scores in axSpA-related patient-reported outcomes and worse response to NSAIDs, bDMARDs and tsDMARDs than those patients with nociplastic pain” should have read “Patients with axSpA who have nociplastic pain frequently show higher scores in axSpA-related patient-reported outcomes and worse response to NSAIDs, bDMARDs and tsDMARDs than those patients with nociceptive pain.” In the “Nociplastic and neuropathic pain as drivers for nociceptive pain scores” section, the text “In the GESPIC cohort (78 patients with axSpA who were treated with bDMARDs) 22% of patients had nociceptive pain (measured by WPI)...” should have read “In the GESPIC cohort (78 patients with axSpA who were treated with bDMARDs) 22% of patients had nociplastic pain (measured by WPI)...”. These corrections have been made to the HTML and PDF versions of the article.

© Springer Nature Limited 2026